

# Mécanismes moléculaires de la survie à long terme chez Propionibacterium freudenreichii

Flavia Figueira Aburjaile

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# RÉSUMÉ ABSTRACT

sée par l'industrie laitière. Elle appartient aux Actinomycètes connus pour leur survie pendant de longues périodes, dans des conditions environnementales défavorables. Pour mieux comprendre ce phénomène, la caractérisation phénotypique de 8 souches de *P. freudenreichii* a été réalisée sur 11 jours dans un milieu en carence nutritionnelle. Le taux de survie tion et évaluation de la viabilité cellulaire. En outre, l'absence de lyse cellulaire a été évaluée par PCR quantitative. La crois-sance de *P. freudenreichii* a été décrite en phases exponen-tielle, stationnaire, stationnaire tardive et survie à long terme. Dans nos conditions expérimentales pendant la période de survie à long terme, les bactéries sont restées viables. La caractérisation phénotypique a montré que P. freudenreichii CIRM-BIA138 était la plus résistante à la carence nutrition nelle et entrait dans un état viable mais non-cultivable. Cette souche a été utilisée pour une étude fonctionnelle par RNA-Seq ainsi que pour des analyses biochimiques sur les surna-geants de culture, en phases exponentielle et stationnaire. L'association de ces données transcriptomiques et métabolomiques a permis de déduire les stratégies impliquées dans la natives d'énergie semblent impliquées dans l'adaptation et la persistence de *P. freudenreichii* CIRM-BIA138 en carence nutritionnelle durant de longues périodes.

Mots-clés : P. freudenreichii, survie à long terme, RNA-Seq,

periods in harsh environmental conditions. In order to investi-gate the long-term survival phenomenon in P. freudenreichii, 8 strains were phenotypically characterized for a period of 11 days in nutrient shortage condition. Bacterial survival rate was assessed by optical density, CFU counting and live-dead cellular as exponential, stationary, late stationary and long-term survival. Moreover, it was observed that bacterial viability was maintained during long-term survival. Phenotypical characterization indica-ted that P. freudenreichii CIRM-BIA138 was more resistant to nutrient shortage being able to enter into a viable but nonculturable dormant state. In addition, functional studies of this strain and stationary growth phases. Concomitantly, several biochemical analyses were carried out on the culture supernatant. An integrative approach of metabolomic and transcriptomic dat integrative approach of metabolomic and transcriptomic data allowed us to infer strategies associated with the survival of this bacterium, such as preparation for the dormant state, slow down of metabolic activity and utilization of alternative sources of ene to adapt and persist through nutrient shortage for long periods.

Keywords: Propionibacterium freudenreichii, long-term survival, RNA-Seq, metabolomics, viable but nonculturable.

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Mécanismes moléculaires de la survie à long terme chez Propionibacterium freudenreichii





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# UNIVERSIDADE FEDERAL DE MINAS GERAIS INSTITUTO DE CIÊNCIAS BIOLÓGICAS PROGRAMA INTERUNIDADES DE PÓS-GRADUAÇÃO EM BIOINFORMÁTICA

Tese

# Mecanismos moleculares de sobrevivência em longo prazo em Propionibacterium freudenreichii

Orientada: Flavia Figueira Aburjaile

Orientadores: Prof. Dr. Vasco Ariston de Carvalho Azevedo

Dr. Yves Le Loir

Rennes

2015



Flavia Figueira Aburjaile



# Mecanismos moleculares de sobrevivência em longo prazo em Propionibacterium freudenreichii

Tese apresentada ao Programa Interunidades de Pós-Graduação em Bioinformática da Universidade Federal de Minas Gerais, como requisito parcial para obtenção do título de Doutor em Bioinformática.

Orientadores: Prof. Dr. Vasco Azevedo Dr. Yves Le Loir

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"Foi o tempo que dedicaste à tua rosa que a fez tão importante".

« C'est le temps que tu as perdu pour ta rose qui fait ta rose si importante ».

Antoine de Saint-Exupéry

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#### **RESUMO**

Propionibacterium freudenreichii é uma bactéria amplamente utilizada na indústria de lactícinios, na síntese de biomoléculas e na fabricação de produtos probióticos. Esta bactéria é pertencente ao grupo de Actinobacterias, que são conhecidas por sua sobrevivência por períodos prolongados a condições ambientais adversas. Para uma melhor compreensão do fenômeno de sobrevivência à longo-prazo, a caracterização fenotípica de 8 linhagens de P. freudenreichii foi realizada neste trabalho durante um período de 11 dias em um meio de carência nutricional. A taxa de sobrevivência bacteriana foi monitorada através da densidade óptica, da enumeração de colônias e da viabilidade celular (kit live/dead). Além disso, a ausência de lise celular foi avaliada utilizando-se a Real-time quantitative PCR (qPCR). Neste trabalho, as fases de crescimento de P. freudenreichii foram descritas e classificadas em: exponencial, estacionária, estacionária tardia e sobrevivência à longo-prazo. Em nossas condições experimentais, observou-se que durante o período de sobrevivência à longo prazo, as bactérias mantiveram-se viáveis. A caracterização fenotípica indica que, dentre as 8 linhagens, P. freudenreichii CIRM-BIA 138 demonstrou maior resistência a escassez nutricional e suas células entraram em um estado de dormência denominado viáveis, mas não cultiváveis (VBNC). Esta linhagem CIRM-BIA 138 foi utilizada para um estudo funcional através da abordagem de RNA-Seq, em fase de crescimento exponencial e estacionária. Concomitante, um conjunto de análises bioquímicas (dosagem de aminoácidos, de ácidos orgânicos e de açucares) foi realizado a partir do sobrenadante durante a cinética bacteriana. A associação destes dados transcriptômicos e metabolômicos permitiu inferir estratégias envolvidas na sobrevivência desta bactéria. A preparação para um estado de dormência, a diminuição do metabolismo e a utilização de fontes alternativas de energia estão envolvidas na adaptação e na persistência de P. freudenreichii CIRM-BIA 138 em escassez nutricional durante longos períodos.

**Palavras-chave:** *Propionibacterium freudenreichii*, mecanismos de sobrevivência à longo prazo, RNA-Seq, metabolômica, VBNC.

#### ABSTRACT

Propionibacterium freudenreichii is a bacterium widely used in the dairy industry, for biomolecule synthesis, and in the probiotic market. This species belongs to the Actinobacteria group, which is known to survive for long periods in harsh environmental conditions. Therefore, in order to better understand the long-term survival phenomenon of *P. freudenreichii*, 8 strains were phenotypically characterized in a laboratory medium for a period of 11 days, in condition of nutrient shortage. Bacterial survival rate was assessed by optical density, plate counting, and by assessing cellular viability (live/dead). In addition, the absence of cell lysis was evaluated by quantitative PCR (qPCR). The P. freudenreichii growth phases were classified as exponential, stationary, late stationary and long-term survival. Under the experimental conditions, it was observed that bacterial viability was maintained during the long-term survival. Phenotypical characterization indicated that P. freudenreichii CIRM-BIA138 was more resistant to nutrient shortage and that this strain was able to enter into a viable but nonculturable dormant state (VBNC). Therefore, functional studies of this strain were conducted by RNA-Sequencing on cultures sampled in the exponential and stationary growth phases. Concomitantly, a series of biochemical studies (quantitation of amino acids, organic acids and sugars) were carried out on the bacterial supernatant. The integration of metabolome and transcriptome data allowed us to infer strategies associated with the survival of this bacterium, such as preparation for the dormant state, slow down of the metabolic activity and utilization of alternative sources of energy, which altogether might allow P. freudenreichii CIRM-BIA 138 to adapt and persist throughout nutrient shortage for long periods.

**Keywords:** *Propionibacterium freudenreichii*, long-term survival mechanism, RNA-Seq, metabolomics, VBNC.

#### RÉSUMÉ

La Propionibacterium freudenreichii est une bactérie largement utilisée dans l'industrie laitière, dans la synthèse de biomolécules et dans la fabrication de produits probiotiques. Cette bactérie appartient au groupe des Actinomycètes, qui sont connues pour leur survie pendant des périodes prolongées dans des conditions environnementales défavorables. Pour mieux comprendre ce phénomène de survie à long terme, la caractérisation phénotypique de 8 souches de P. freudenreichii a été réalisée dans ce travail sur une période de 11 jours dans un milieu en carence nutritionnelle. Le taux de survie bactérienne a été mesuré par densité optique, par énumération et évaluation de la viabilité cellulaire (live/dead). En outre, l'absence de lyse cellulaire a été évaluée par PCR quantitative (qPCR). Dans ce travail, la croissance de P. freudenreichii ont été a été décrite et divisée en phases exponentielle, stationnaire, stationnaire tardive et survie à long terme. Dans nos conditions expérimentales, il a été observé que, pendant la période de survie à long terme, les bactéries restaient viables. La caractérisation phénotypique des 8 souches a montré que P. freudenreichii CIRM-BIA138 était la plus résistante à la carence nutritionnelle et que les cellules bactériennes entraient dans un état de dormance qualifié d'état viable mais non-cultivable (VBNC). Cette souche CIRM-BIA138 a été utilisée pour une étude fonctionnelle par une approche RNA-Seq, sur les phases de croisssance exponentielle et stationnaire. Parallèlement, un ensemble d'analyses biochimiques (dosage d'acides aminés, d'acides organiques et de sucres) a été réalisé sur les surnageants de culture au cours de la cinétique. L'association de ces données transcriptomiques et métabolomiques a permis de déduire quelques stratégies impliquées dans la survie de cette bactérie. La préparation à l'état de dormance, la diminution du métabolisme et l'utilisation de sources alternatives d'énergie semblent impliqués dans l'adaptation et la persistence de P. freudenreichii CIRM-BIA138 en carence nutritionnelle pendant de longues périodes.

**Mots-clés :** *Propionibacterium freudenreichii,* mécanismes de survie à long terme, RNA-Seq, métabolomique, VBNC.

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#### LISTA DE ABREVIATURAS

	Inglês	Português
ATP	adenosine triphosphate	adenosina trifosfato
BLAST	basic local alignment search tool	ferramenta de busca de alinhamento local básico
bp	base pairs	pares de bases
cDNA	complementary DNA	DNA complementar
CFU	colony forming unit	unidade formadoras de colônias
CIRM-BIA	biological resource centre fully dedicated to bacteria of food interest	centro de recursos biológicos especializado em bactérias de interesse alimentar
DNA	deoxyribonucleic acid	ácido desoxirribonucleico
DHNA	1.4-dihydroxy-2-naphthoic acid	ácido 1,4-dihidróxi 2-naftóico
EMBL-EBI	European Molecular Biology Laboratory - European Bioinformatics Institute	laboratório europeu de biologia molecular - instituto europeu de bioinformática
FAO	food and agriculture organization	organização de alimentação e da agricultura
GOLD	genomes online database	banco de dados de genomas disponíveis ( <i>online</i> )
GRAS	generally recognised as safe	geralmente reconhecido como seguro
INRA	national institute of agronomic research	instituto nacional de pesquisa agronômica
KEGG	kyoto encyclopedia of genes and genomes	enciclopédia kyoto de genes e genomas
LGCM	laboratory of cellular and molecular genetics	laboratório de genética celular e molecular
mRNA	messenger RNA	RNA mensageiro
NGS	next-generation sequencing	sequenciamento de próxima geração
OD	optical density	densidade óptica

ORFs	open reading frame	quadro aberto de leitura
PCR	polymerase chain reaction	reação em cadeia da polimerase
PPC	cooked pressed paste	massa prensada cozida
qPCR	real-time quantitative PCR	PCR quantitativa em tempo real
QPS	qualified presumption of safety	presunção qualificada de segurança
RNA-Seq	RNA sequencing	sequenciamento de cDNA
rRNA	ribosomal ribonucleic acid	ácido ribonucleico ribossômico
RT-PCR	reverse transcription - polymerase chain reaction	reverse transcriptase - réaction en chaîne par polymérase
SCFA	short-chain fatty acids	ácidos graxos de cadeia curta
STLO	science and technology of milk and eggs	ciência e tecnologia do leite e dos ovo
tRNA	transfer ribonucleic acid	ácido ribonucleico transportador
VBNC	viable but non-culturable	viáveis mas não cultiváveis
WHO	world health organization	organização mundial da saúde
YEL	yeast extract lactate	extrato de levedura-lactato

#### LISTE DE ABREVIATIONS UTILISEES

A	nglais	Français
ATP	adenosine triphosphate	adénosine-5 -triphosphate
BLAST	basic local alignment search tool	outil de recherche de base d'alignement local
bp	base pairs	paires de bases
cDNA	complementary DNA	ADN complémentaire
CFU	colony forming unit	unité formant colonie (UFC)
CIRM-BIA	biological resource centre fully dedicated to bacteria of food interest	centre international de ressources microbiennes - bactéries d'intérêt alimentaire
DNA	deoxyribonucleic acid	acide désoxyribonucléique (ADN)
DHNA	1.4-dihydroxy-2-naphthoic acid	l'acide 1,4-dihydroxy -2-naphtoïque
EMBL-EBI	european molecular biology laboratory - european bioinformatics institute	laboratoire européen de biologie moléculaire - Institut européen de bio- informatique
FAO	food and agriculture organization	organisation des Nations Unies pour l'alimentation et l'agriculture
GOLD	genomes online database	base de données de génomes en ligne
GRAS	generally recognised as safe	présentant une inocuité sanitaire / généralement reconnu comme sûr
INRA	national institute for agricultural research	institut national de la recherche agronomique
KEGG	kyoto encyclopedia of genes and genomes	encyclopédie de Kyoto des gènes et des génomes
LGCM	laboratory of cellular and molecular genetics	laboratoire de génétique cellulaire et moléculaire
mRNA	messenger RNA	ARN messager
NGS	next-generation sequencing	séquençage de prochaine génération
OD	optical density	densité optique

ORFs	open reading frame	cadre ouvert de lecture
PCR	polymerase chain reaction	réaction on chaîne de la polymérose
PPC	cooked pressed paste	reaction en chame de la porymerase
aPCR	real-time quantitative PCR	pâte pressée cuite
		PCR quantitative en temps réel
QPS	qualified presumption of safety	présomption d'innocuité reconnue
RNA-Seq	RNA sequencing	séquencage à ARN
rRNA	ribosomal ribonucleic acid	anida ribarual di sua ribara mi
RT-PCR	reverse transcription - polymerase	acide moonucleique moosonnique
	chain reaction	transcriptase reversa seguida da reação em cadeia da polimerase
SCFA	short-chain fatty acids	acidas gras à chaîna courta
STLO	science and technology of milk	actues gras a chame courte
	and eggs	science & technologie du lait & de l'œuf
tRNA	transfer ribonucleic acid	acides ribonucléiques de transfert
VBNC	viable but non-culturable	wichles mais non cultivehles
WHO	world health organization	viables mais non cultivables
VEL		organisation mondiale de la santé
IEL	yeast extract factate	bouillon lactate et extrait de levure

# **APRESENTAÇÃO / PRÉSENTATION**

#### Colaborações

Este trabalho de tese foi realizado em Cotutela de acordo com as convenções propostas pelos estabelecimentos francês e brasileiro respectivamente, *Agrocampus Ouest*, representado pelo Sr. Grégoire Thomas e Universidade Federal de Minas Gerais – UFMG, representado pelo seu Reitor, na época, Professor Clélio Campolina Diniz. A tese foi realizada no laboratório de *Science & technologie du lait et de l'œuf* (STLO) localizado no *Agrocampus Ouest* (UMR 1253 - *Institut National de la Recherche Agronomique*) em Rennes (França) e no laboratório de Genética Celular e Molecular (LGCM) situado no Instituto de Ciências Biológicas na Universidade Federal de Minas Gerais (UFMG) em Belo Horizonte (Brasil). O desenvolvimento das atividades foi coordenado na França pelo Dr. Yves Le Loir e no Brasil pelo Prof. Dr. Vasco Azevedo. O convênio estabelecido em Cotutela permite a obtenção de um duplo diploma entre as instituições conveniadas, que são: Instituto Superior de Ciências Agrícolas, Agro-Alimentaires, Hortícolas e de Paisagem (*Agrocampus Ouest*), como representante francesa, e a UFMG, como representante brasileira.

Desde 1979, vários projetos foram realizados em colaboração franco-brasileira, envolvendo o Ministério da Educação (Brasil) através da CAPES, e o Ministério das Relações Exteriores e Européias e o Ministério do Ensino Superior e da Pesquisa (França), através do COFECUB. O financiamento da tese foi possível graças a este programa CAPES-COFECUB (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – Comitê Francês de Avaliação da Cooperação Universitária com o Brasil) que permitiram o desenvolvimento deste doutorado durante 18 mêses em Rennes (França). Este programa tem como objetivo, o intercâmbio científico de alunos da pós-graduação entre instituições de ensino superior do Brasil e da França, bem como visa a formação de recursos humanos nestes países.

#### Collaborations

Cette thèse a été réalisée en co-tutelle selon les conventions proposées par les établissements français et brésilien, respectivement Agrocampus Ouest, representé par M. Grégoire Thomas et Universidade Federal de Minas Gerais – UFMG, representé par son recteur, à l'époque le Professeur Clélio Campolina Diniz. La thèse a été réalisée au laboratoire de Science & technologie du lait et de l'œuf (STLO) situé à Agrocampus Ouest (UMR 1253, à l'Institut National de la Recherche Agronomique) à Rennes (France) et au laboratoire de Génétique Cellulaire et Moléculaire situé dans l'Institut de Sciences Biologiques à l'UFMG à Belo Horizonte (Brésil). Le développement des activités a été coordonné en France par le Dr. Yves Le Loir et au Brésil par le Prof. Dr Vasco Azevedo. L'accord établi en co-tutelle permet d'obtenir un double diplôme entre les institutions partenaires que sont l'Institut Supérieur des Sciences Agronomiques, Agroalimentaires, Horticoles et du Paysage (Agrocampus Ouest) du côté français, et l'UFMG du côté brésilien.

Depuis 1979, plusieurs projets ont été réalisés dans le cadre de la coopération franco-brésilienne, avec le Ministère de l'Éducation (Brésil), par le biais de la CAPES, et du Ministère des Affaires Etrangères et Européennes et du Ministère de l'Enseignement Supérieur et de la Recherche (France), représenté par COFECUB. Le financement de la thèse a été possible grâce au programme CAPES-COFECUB (Coordination pour le Perfectionnement du Personnel de l'Enseignement Supérieur – Comité français d'évaluation de la coopération universitaire et scientifique avec le Brésil) qui a permis le développement de ce doctorat pendant 18 mois à Rennes (France). Ce programme vise l'échange scientifique des étudiants universitaires dans les établissements d'enseignement supérieur du Brésil et de France, ainsi que la formation des ressources humaines dans ces pays.

#### Prefácio

No final do século XVI, bactérias foram descritas pela primeira vez por Antoine van Leeuwenhoek. Algumas bactérias foram exploradas à partir de experimentos realizados pelo Robert Koch, o qual as descreveu inicialmente como patógenos de animais e de homens, que viria a ser, mais tarde, denominado de Mycobacterium pseudotuberculosis (bacilo de Koch). Na mesma linha, trabalhos do Louis Pasteur, pesquisador francês, contribuíram para a compreensão das bactérias, seres procariotos e ubíquos, capazes de crescer em diferentes temperaturas, ambientes, intensidades de luz e presença e/ou ausência de oxigênio. Isto demonstra a capacidade de adaptação destes seres e sua aptidão de enfrentar condições favoráveis ou desfavoráveis. Um componente chave para esta aptidão é a proliferação e sobrevivência bacteriana por longos períodos (Long-Term Survival). Existem diferentes graus de sobrevivência desde a fase de latência de uma bactéria até o período denominado de "sobrevivência a longo prazo". Apesar de muito utilizado na literatura, este conceito de "sobrevivência a longo prazo" está associado a persistência bacteriana em diferentes frações de tempo, como: horas, dias, meses e até anos. Esta sobrevivência a longo prazo é observada em bactérias isoladas de ambientes variados, tais como: solo, queijo, água, fósseis e de humanos (Greenblatt et al. 2004a; Lemunier et al. 2005; Finkel 2006a).

Dentre as bactérias que apresentam esta capacidade de sobreviver por longos períodos, estão aquelas pertencentes ao Filo Actinobacteria. Estudos de sobrevivência à longo prazo já foram realizados com diferentes espécies, como: *Micrococcus luteus* (Greenblatt *et al.* 2004a), *Mycobacterium bovis* (Fine *et al.* 2011), *M. tuberculosis* (Primm *et al.* 2000), *M. leprae* (Wheat *et al.* 2014), *Bifidobacterium lactis* (Olszewska, Staniewski and Łaniewska-Trokenheim 2012) e *Propionibacterium freudenreichii* (Dalmasso *et al.* 2012a). Dentro deste grupo bacteriano citado acima, temos bactérias que são classificadas como patogênicas e não-patogênicas. As bactérias patogênicas são mais estudadas, uma vez que são agentes de doenças e infecções de importância mundial. Uma pesquisa rápida no PubMed revela que termos como "pathogens bacteria survival" apresentam mais de 8400 resultados, enquanto que "probiotics bacteria survival" o retorno foi de 1074 resultados (acesso em http://www.ncbi.nlm.nih.gov/pubmed em 15-08-2015).

Para que bactérias patogênicas possam sobreviver é necessário que estas desenvolvam certas habilidades como aderir, invadir, colonizar o hospedeiro, evadir ao sistema imune e adquirir nutrientes a partir do hospedeiro, permitindo assim a adaptação

a diversos ambientes, muitas vezes hostis. Esse fenômeno de sobrevivência por longos períodos, já foi relatado em um modelo de infecção por *M. leprae*, o qual demonstrou que estas bactérias são capazes de reter a sua virulência *in vivo (nu/nu mouse model)* e estas permanecerem viáveis por até 8 meses dentro de cistos de ameba. Isto reforça a idéia de que estas bactérias podem persistir em vários hospedeiros ou ambientes (Wheat *et al.* 2014).

Em bactérias não-patogênicas, o estudo da sobrevivência apresenta um interesse biotecnológico, uma vez que as bactérias são utilizadas na agricultura, alimentação, medicina animal e humana. Na indústria alimentícia, sobretudo de laticínios, esta sobrevivência é essencial para manter uma atividade bacteriana nos produtos, por exemplo, probióticos que promovem funções benéficas ao hospedeiro (Lan *et al.* 2007).

Neste contexto, é interessante um bom entendimento sobre as estratégias de sobrevivência em bactérias não-patogênicas, principalmente de interesse biotecnólogico na indústria alimentícia, como por exemplo, bactérias propiônicas. Estas últimas são alvo de pesquisa, desde o final dos anos 80, do laboratório de Science & Technologie du Lait & de l'Oeuf (STLO), uma unidade mista de pesquisa situada no Agrocampus Ouest e INRA, e foi o instituto associado a este trabalho de Tese. Estudos recentemente realizados em P. freudenreichii, demonstraram que esta bactéria resiste a diferentes temperaturas, como nas etapas importantes de maturação dos queijos de massa prensada e cozida (PPC). Estas bactérias mantém-se vivas em concentrações elevadas até o final da maturação e armazenamento destes produtos (Dalmasso et al. 2012a). Nos últimos anos, muitas pesquisas de biologia molecular têm sido desenvolvidas para a caracterização de P. freudenreichii, no intuito de melhorar a nossa compreensão a respeito da biologia desta bactéria, além de aprimorar a sua utilização em processos de fabricação de queijos ou em outras aplicações biotecnológicas (Parayre et al. 2007; Falentin et al. 2010c; Dalmasso et al. 2011). Além disso, P. freudenreichii é bom candidato probiótico, uma vez que ela sobrevive ao longo do trato gastrointestinal, exerce uma atividade imunomoduladora, produz substâncias antibacterianas e entra em competição com outras bactérias patogênicas por nutrientes e sítios de ligação (Jan, Rouault and Maubois 2000; Leverrier et al. 2003a; Lan et al. 2007).

Visando uma melhor compreensão da sobrevivência desta bactéria, o presente trabalho de tese aborda o estudo e a caracterização de estratégias de sobrevivência em *P. freudenreichii*.

O manuscrito é constítuido em três capítulos: (I) revisão da literatura, (II) resultados e (III) discussão, conclusão e perspectivas. A primeira seção do trabalho é composta por uma revisão da literatura sobre o gênero *Propionibacterium* e sua espécie *P. freudenreichii*, auxiliando a nossa compreensão a respeito da biologia desta bactéria.

Ao longo dos últimos anos, o STLO já abordou o estudo da adaptação do *P. freudenreichii* a diferentes estresses ou ambientes, como em cólon de hospedeiro (modelo de porco) (Saraoui *et al.* 2013a), ou em processos de maturação de queijos PPC (Dalmasso *et al.* 2012a). Desse modo, os estudos da equipe STLO objetivaram a diferenciação das espécies de *P. freudenreichii* em subespécies, testando a capacidade para fermentação de lactose e redução de nitrato, além de analisarem a produção de compostos aromáticos. Isto permite selecionar uma linhagem de interesse na fabricação de queijos, além de ampliar conhecimentos gerais a cerca da fisiologia e metabolismo de *P. freudenreichii* (Thierry *et al.* 2011).

Assim, seguindo uma linha de pesquisa que visa melhor compreensão biológica deste microrganismo, o segundo capítulo deste manuscrito apresenta os resultados obtidos ao longo destes 4 anos de tese, sendo subdivido em duas seções: (I) caracterização fenotípica em diferentes fases do crescimento bacteriano durante um período de escassez de nutrientes de 11 dias; e (II) estudo da genômica funcional de P. freudenreichii para investigação ao nível molecular do fenômeno de adaptação à fase estacionária. Para isso, a metodologia utilizada foi o RNA-Seq, a qual permitiu analisar a expressão gênica global em fase exponencial e em fase estacionária de P. freudenreichii. Esta abordagem permitiu identificar os genes envolvidos na entrada da fase estacionária e previamente ao estado de dormência, em uma linhagem anteriormente selecionada por sua capacidade de sobrevivência à longo prazo. Como complemento a este estudo, os dados transcriptômicos foram associados aos dados bioquímicos (in vitro) gerados analisando a cinética do conteúdo das culturas em aminoácidos, ácidos orgânicos e açucares, resultado do metabolismo desta espécie. Estas seções são destinadas a identificar as estratégias que permitem à P. freudenreichii de sobreviver à longo-prazo em condições de carência nutricional.

Como último capítulo, teremos a discussão geral deste trabalho, seguido de conclusões e perspectivas.

#### Préambule

À la fin du XVIe siècle, les bactéries ont d'abord été décrites pour la première fois par Antoine van Leeuwenhoek. Certaines bactéries ont été explorées à partir d'expériences menées par Robert Koch, qui les décrit d'abord comme des pathogènes d'animaux et de l'homme, plus tard, ces bactéries recevront le nom de Mycobacterium pseudotuberculosis (bacille de Koch). Dans le même domaine, les travaux de Louis Pasteur contribuèrent à la compréhension des bactéries qui sont des êtres procaryotes et ubiquistes, capables de croître à différentes températures, dans différents environnements, à des intensités de lumière variables et en présence ou absence d'oxygène. Cela démontre la capacité d'adaptation de ces organismes et leur aptitude à affronter des conditions favorables ou défavorables. Une composante clé à cette aptitude est la prolifération et la survie bactérienne pendant de longues périodes (Long-Term Survival). Il existe différents degrés de survie dès la phase de latence d'une bactérie jusqu'à la période appelée « la survie à long terme ». Bien que cette expression soit très utilisée en littérature, ce concept de « survie à long terme » est associé à la persistance bactérienne à diverses échelles de temps, comme : l'heure, le jour, le mois et même l'année. Cette survie à long terme s'applique aux bactéries isolées d'environnements très variés , tels que: le sol, le fromage, l'eau, les fossiles et les êtres humains (Greenblatt et al. 2004a; Lemunier et al. 2005; Finkel 2006a).

Certaines des bactéries particulièrement reconnues pour leur capacité de survie à long terme appartiennent au *phylum* des *Actinobacteria*. Les études de survie à long terme ont été réalisées sur différentes espèces, telles que : *Micrococcus luteus* (Greenblatt *et al.* 2004a), *Mycobacterium bovis (Fine et al.* 2011), *M. tuberculosis (Primm et al.* 2000), *M. leprae (Wheat et al.* 2014), *Bifidobacterium lactis* (Olszewska, Staniewski and Łaniewska-Trokenheim 2012) et *Propionibacterium freudenreichii (Dalmasso et al.* 2012a). Dans le groupe bactérien mentionné ci-dessus, il y a aussi bien des bactéries pathogènes que non-pathogènes. Les bactéries pathogènes sont plus étudiées, car elles sont des agents de maladies et d'infections d'importance mondiale. Une recherche rapide sur PubMed révèle que des termes tels que « pathogens bacteria survival » présentent plus de 8400 entrées, tandis que « probiotics bacteria survival » en donnent 1074 (accès www.ncbi.nlm.nih.gov/PubMed en 15-08 -2015).

Pour que les bactéries pathogènes puissent survivre, il faut que celles-ci développent certaines capacités telles qu'adhérer, envahir, coloniser son hôte, échapper au système immunitaire et acquérir des nutriments à partir de l'hôte, leur permettant ainsi de s'adapter à différents environnements, souvent hostiles. Ce phénomène de survie pendant de longues périodes a déjà été décrit dans un modèle d'infection pour *M. leprae*, qui démontre que ces bactéries sont capables de maintenir leur virulence, *in vivo*, sur modèle murin et qu'elles restent viables jusqu'à huit mois dans des kystes chez les amibes. Cela renforce l'idée que ces bactéries peuvent persister dans plusieurs hôtes voire plusieurs environnements (Wheat *et al.* 2014).

Dans les bactéries non-pathogènes, l'étude de la survie présente un intérêt biotechnologique, puisque que les bactéries sont utilisées dans l'agriculture, l'alimentation, la médecine vétérinaire et humaine. Dans l'industrie alimentaire, en particulier des produits laitiers, cette survie est essentielle pour maintenir une activité bactérienne dans les produits, par exemple, pour les probiotiques qui expriment des fonctions bénéfiques pour l'hôte (Lan *et al.* 2007).

Dans ce contexte, il est intéressant de bien comprendre les stratégies de survie des bactéries non-pathogènes, surtout d'intérêt biotechnologique dans l'industrie alimentaire, comme par exemple les bactéries propioniques. Depuis la fin des années 1980, celles-ci sont la cible de recherche du laboratoire de Science & Technologie du Lait et de l'Oeuf (STLO), une unité mixte de recherche (UMR) Agrocampus Ouest et INRA, où s'est déroulé ce travail de thèse. Des études récentes effectuées sur P. freudenreichii ont montré que cette bactérie est résistante à des températures variables, comme dans les étapes importantes d'affinage des fromages à pâte pressée cuite (PPC). Ces bactéries restent vivantes, à des concentrations élevées, jusqu'à la fin de l'affinage et pendant le stockage de ces produits (Dalmasso et al. 2012a). Au cours des dernières années, de nombreuses recherches de biologie moléculaire ont été mises au point pour la caractérisation de P. freudenreichii, afin d'améliorer notre compréhension biologique de cette bactérie, ainsi que pour améliorer son utilisation dans le processus de fabrication fromagère ou d'autres applications biotechnologiques (Parayre et al. 2007; Falentin et al. 2010c; Dalmasso et al. 2011). Par ailleurs, P. freudenreichii est une bonne candidate probiotique, car elle survit dans le tractus gastro-intestinal, exerce une activité immunomodulatrice, produit des substances antibactériennes et entre en compétition avec d'autres bactéries pathogènes pour les nutriments et les sites de liaison (Jan, Rouault and Maubois 2000; Leverrier et al. 2003a; Lan et al. 2007).

Pour améliorer notre compréhension de la survie de cette bactérie, ce travail de thèse aborde l'étude et la caractérisation des stratégies de survie chez *P. freudenreichii*.

Le manuscrit est constitué de 3 chapitres : (I) révision de la littérature, (II) les résultats et (III) la discussion, la conclusion et les perspectives. La première partie est composé d'un introduction bibliographique sur le genre *Propionibacterium* et l'espèce *P. freudenreichii* nous aider à comprendre le phénomène de survie à long terme chez *P. freudenreichii*.

Au cours de ces dernières années, le STLO a déjà abordé l'étude de l'adaptation de *P. freudenreichii* à différents stress ou dans différents environnements, tels que le côlon de l'hôte (modèle du porc) (Saraoui *et al.* 2013a), ou dans le processus d'affinage des fromages PPC (Dalmasso *et al.* 2012a). De même, les recherches de l'équipe STLO se sont focalisées sur la différenciation des espèces de *P. freudenreichii* en sous-espèces, en testant la capacité de fermentation de lactose et la réduction de nitrate, en plus d'analyser la production des composants aromatiques. Ceci permet de sélectionner une souche d'intérêt dans la fabrication des fromages, en plus d'élargir les connaissances générales liées à la physiologie et au métabolisme de *P. freudenreichii* (Thierry *et al.* 2011).

Ainsi, suivant une ligne de recherche qui vise une meilleure compréhension biologique de ce microorganisme, le deuxième chapitre de ce manuscrit présente les résultats obtenus au cours de ces quatre années de thèse, qui sont subdivisés en deux sections: (i) la caractérisation phénotypique à différentes phases de croissance bactérienne pendant une période de 11 jours de carence nutritionnelle; et (ii) l'étude de la génomique fonctionnelle de P. freudenreichii pour aborder au niveau moléculaire le phénomène d'adaptation à la phase stationnaire. Pour cela, la méthode utilisée a été le RNA-Seq, qui a permis l'analyse globale de l'expression des gènes en phase exponentielle et stationnaire de P. freudenreichii. Cela à permis d'identifier les gènes impliqués dans l'entrée en phase stationnaire et précédant l'état de dormance, dans une souche précédemment sélectionnée pour sa capacité de survie à long terme. En plus de cette étude, les données transcriptomiques ont été associées à des données biochimiques (in vitro) générées en analysant l'évolution du contenu des cultures en acides aminés, acides organiques et sucres résultant du métabolisme de cette espèce. Ces sections étaient destinées à identifier des stratégies qui permettent à P. freudenreichii de survivre à long terme en condition de carence nutritionnelle.

Le dernier chapitre, comprend une discussion générale de ce travail, suivie des conclusions et perspectives.

**Objetivos / Objectifs** 

#### Objetivos

#### Geral

Esta tese visa a identificar os mecanismos de sobrevivência a longo prazo em *Propionibacterium freudenreichii* que permitem sua adaptação à períodos prolongados durante a carência nutricional.

#### Específicos

- Avaliar e caracterizar o fenótipo (morfologia, viabilidade e integridade de membrana) de 8 linhagens de *P. freudenreichii*, a fim de aferir a variabilidade entre as linhagens na capacidade de sobrevivência a longo prazo e de selecionar a linhagem que apresenta a melhor sobrevida;
- J Identificar a presença de células viáveis, mas não cultiváveis (VBNC) em *P. freudenreichii* através da associação de diferentes metodologias: UFC, microscopia de epifluorescência e qPCR;
- ) Identificar genes que são diferencialmente expressos em fases de crescimento exponencial e estacionária de *P. freudenreichii*;
- ) Analisar os genes possivelmente envolvidos no mecanismo de sobrevivência de *P. freudenreichii* de acordo com a técnica de RNA-Seq;
- ) Identificar, através de dados transcriptômicos e bioquímicos da linhagem selecionada (*P. freudenreichii* CIRM-BIA 138), quais são os genes presentes no organismo e quais são os fatores ambientais implicados para o desencadeamento de mecanismos moleculares que permitem a sobrevivência desta bactéria durante um período de carência nutricional;
- ) Correlacionar os resultados obtidos na análise do transcriptoma com os dados bioquímicos (dosagem de ácidos orgânicos, aminoácidos e açúcares) para validar a atividade metabólica de *P. freudenreichii*.

#### **Objectifs**

#### Généraux

Cette thèse vise à identifier les mécanismes de survie à long-terme chez *Propionibacterium freudenreichii* qui permettent l'adaptation à de longues périodes de carence nutritionnelle.

#### Spécifiques

• Évaluer et caractériser le phénotype (la morphologie, la viabilité et l'intégrité membranaire) de 8 souches de *P. freudenreichii*, afin d'évaluer la variabilité inter-souches dans la capacité de survie à long terme et de sélectionner la souche présentant la meilleure survie du panel ;

• Identifier la présence de cellules viables mais non cultivables (VBNC) en *P. freudenreichii* par l'association de différentes méthodologies : UFC, microscopie épifluorescence et PCRq ;

• Identifier des gènes qui sont différentiellement exprimés dans les phases de croissance exponentielle et stationnaire de *P. freudenreichii* ;

• Analyser les gènes possiblement impliqués dans le mécanisme de survie à long-terme de *P. freudenreichii* selon la technique de RNA-Seq ;

• Identifier, à travers les données transcriptomiques et biochimiques de la souche sélectionnée (*P. freudenreichii* CIRM-BIA 138), quels sont les gènes présents et les facteurs environnementaux impliqués dans le déclenchement des mécanismes moléculaires permettant la survie de cette bactérie pendant une période de carence nutritionnelle ;

• Corréler les résultats obtenus dans l'analyse du transcriptome avec les données biochimiques (dosage d'acides organiques et aminés et des sucres) pour valider l'activité métabolique de *P. freudenreichii*.
# CAPÍTULO 1: REVISÃO DA LITERATURA

CHAPITRE 1 : RÉVISION DE LA LITTÉRATURE Esta parte do manuscrito é dedicada a apresentação de dados gerais sobre o gênero *Propionibacterium* e a apresentação, mais detalhada, da espécie *P. freudenreichii*, relativo a sua utilização na indústria, suas propriedades metabólicas e probióticas, e sua capacidade de resistir aos diferentes estresses.

#### 1. Gênero Propionibacterium

#### 1.1. Ecologia, fisiologia e classificação

Em 1906, Von Freudenreichi e Orla-Jensen, relataram o primeiro isolamento de bactérias propiônicas a partir do queijo Emmental (von Freudenreich and Orla-Jensen 1906). Em 1909, o nome do gênero foi estabelecido por Orla-Jensen, sendo então denominado de *Propionibacterium* (Cummins and Johnson 1986), o qual foi proposto devido a capacidade desta bactéria de produzir ácido propiônico durante o processo de fermentação (Orla-Jensen 1909). Posteriormente, em 1946, Douglas e Gunter propuseram a inserção de bactérias pertencentes ao grupo de *Corynebacterium acnes* dentro do gênero de *Propionibacterium*. E, mais tarde em 1963, outros autores confirmaram esta reclassificação, sendo então *C. acnes* uma espécie incluida do genêro *Propionibacterium* (Moore and Cato 1963).

Bactérias do genêro Propionibacterium são bactérias Gram-positiva, pertencente à ordem Actinomycetales, dentro do filo Actinobacteria. Elas são imovéis e não formam esporos. Este gênero pode ser dividido em dois grandes grupos de acordo com o seu habitat: (i) lactícinios, os quais encontramos bactérias propiônicas lácticas; (ii) microbiota comensal da pele e/ou de membranas mucosas, os quais encontramos bactérias cutâneas ou comensais. No grupo de bactérias propiônicas lácticas, podemos encontrar diferentes espécies, as quais as mais conhecidas são: P. freudenreichii, P. thoenii, P. jensenii e P. acidipropionici. Estas espécies não são patogênicas para os seres humanos, e geralmente, são isoladas de produtos alimentares fermentados, como: silagem, fermentação de olivas, azeitonas e destilaria de rum. Em casos mais raros, podem ser isolados do meio-ambiente, por exemplo, em solos, e de animais, em especial no trato intestinal de ratos (Mantere-Alhonen 1982; Balows 1992). Outras espécies como: P. cyclohexanicum (Kusano et al. 1997) e P. microaerophilum (Koussémon et al. 2001) também foram recentemente descritas e classificadas pertencentes a este grupo de bactérias propiônicas lácticas não patogênicas, apesar de não serem isoladas de produtos lactéos, mas de alimentos variados ou de subprodutos. Exemplos destas ocorrências são:

*P. cyclohexanicum* encontrada em suco fermentado de laranja, e *P. microaerophilum* descrita em águas residuais de lagares de azeite. Entretanto, no grupo de *Propionibacterium* comensal ou cutânea, existem três espécies, tais como: *P. acnes*, *P. avidum* e *P. propionicum*, causando patologias em seres humanos. As espécies *P. lymphophilum* e *P. granulosum* pertencem também a classificação de comensais, porém representam um grupo distinto por estarem, filogeneticamente, mais distante de outros grupos (Meile *et al.* 1999).

### 1.2. Caracterização e importância do microrganismo em estudo: *Propionibacterium freudenreichii*

Em condições laboratoriais, o crescimento de *Propionibacterium freudenreichii* ocorre geralmente em condições ótimas no meio YEL (*Yeast Extract Lactate*) proposto por Malik e colaboradores (Malik, Reinbold and Vedamuthu 1968) e a uma temperatura de incubação de 30°C. Esta bactéria é anaeróbia facultativa que pode ser encontrada *in vitro* sob diversas formas (pleomórfica): cocóide, bífida ou ramificada (Figura 1).



Figura 1: Aspecto pleomórfico de *P. freudenreichii* durante o estresse ácido: (A) células não tratadas; (B) células submetidas a pH= 2,0; (C) células submetidas a pH= 5,0; (D) células submetidas a pH= 5,0, e posteriormente, submetidas ao pH=2,0 visualizadas por microscopia eletrônica de varredura (Jan *et al.*, 2001).

Antigamente, a espécie *P. freudenreichii* foi divida em duas sub-espécies: *P. freudenreichii* subsp. *freudenreichii*, que não é capaz de fermentar lactose, mas pode reduzir nitrato; e a *P. freudenreichii* subsp. *shermanii*, que fermenta lactose, porém não é capaz de reduzir o nitrato (Cummins and Johnson 1986). Porém, estudos recentes demonstram que existem linhagens de *P. freudenreichii* que podem conter os dois fenótipos: lactose e nitrato redutase positiva ou lactose e nitrato redutase negativa (de Carvalho, Gautier and Grimont 1994; Vorobjeva 1999; Dalmasso *et al.* 2011). Além disso, no genoma completo de *P. freudenreichii* CIRM-BIA 1, foi observado que o gene *lacZ*, responsável pela atividade beta-galactosidase, esta localizado numa região de transposase e que existem pseudogenes na região do genoma que codifica a informação necessária à redução de nitrato (Falentin *et al.* 2010a). Atualmente, não há uma classificação de subespécies ao nível fenotípico e outras investigações para explorar a genômica desta bactéria devem ser efetuadas. Isto é grande importância na indústria alimentícia, uma vez que estas propriedades bioquímicas podem alterar a qualidade final do produto, por exemplo, queijos PPC.

P. freudenreichii é um microrganismo seguro (saúde-sanitária) e possui dois importantes status para sua utilização: o Generally Recognized As Safe (GRAS) outorgado pelo órgão administrativo de alimentos e drogas (Food and Drug Administration - FDA, nos Estados Unidos) (Mogensen et al. 2002) e o Qualified Presumption of Safety (QPS) concedido pela autoridade européia de segurança dos alimentos (European Food Safety Authority - EFSA) (EFSA experts 2008). Estes status são aprovados por autoridades do setor que visam controlar a qualidade dos produtos, garantindo a segurança e a autorização destas bactérias na indústria de alimentos, como por exemplo, na fabricação de queijos Emmental. Estas bactérias são amplamente encontradas na natureza, incluindo o trato gastrointestinal, e estão presentes em muitos alimentos fermentados por possuírem características que as permitem de resistir ao pH ácido (Jan, Rouault and Maubois 2000; Huang and Adams 2004; Suomalainen et al. 2006) e a baixas temperaturas no processo de maturação de queijos (Dalmasso et al. 2012a). A concentração destas bactérias, geralmente, encontrada no Emmental são na ordem de 109 UFC/g. Estas bactérias possuem qualidades que são de grande importância para diversas aplicações na área industrial, as quais serão abordadas com mais detalhes na próxima seção.

#### 2. Aplicações industriais de Propionibacterium freudenreichii

Os alimentos fermentados, em particular os lactícinios, são consumidos pelo homem há milhões de anos. Desde o tempo da Idade Média, o processo de fabricação de queijos do tipo Emmental nos Alpes da região de Savoie é descrito em manuscritos. No século XIX, aparecem os primeiros relatos sobre "frutières", que são locais de exploração e transformação do leite em queijo, onde os agricultores, em conjunto, agrupam o "fruto" de suas produções de verão. Em seguida, no século XX, uma forte industrialização foi observada, conduzindo a uma deslocalização da produção destes queijos.

Assim como o Emmental, todos os laticínios que foram anteriormente iniciados empiricamente, começaram a se beneficiar de avanços no campo da microbiologia e da tecnologia de alimentos, o que permitiu um melhor controle, diversificação e segurança de suas produções.

As principais espécies bacterianas utilizadas hoje para a produção de alimentos fermentados são pertencentes aos gêneros: *Bifidobacterium*, *Lactobacillus*, *Lactococcus*, *Leuconostoc*, *Streptococcus* e *Propionibacterium* (Tabela 1 e 2). Estes gêneros exercem papéis importantes na indústria de laticínios, na síntese de biomoléculas, bem como no desenvolvimento de produtos probióticos.

Neste contexto, *P. freudenreichii* desempenha um papel importante na fabricação e, particularmente, na maturação de queijos PPC. Esta espécie é também utilizada por sua capacidade de produzir bacteriocinas, compostos do ácido linoléico, do ácido propiônico, propileno, trealose e vitaminas do complexo B, como por exemplo, a vitamina B12 (Piao *et al.* 2004; Poonam *et al.* 2012). Algumas linhagens possuem também propriedades probióticas e podem modular a composição da microbiota intestinal produzindo fatores bifidogênicos, ou modular ao nível de inflamação no contexto de doenças inflamatórias intestinais crônicas. Estas bactérias ainda são interessantes por serem capazes de resistir ao estresse do trato gastrointestinal.

Como descrito anteriormente, esta bactéria apresenta capacidade de adaptação à estresses de natureza variável, como: pH, temperatura e sais biliares. Esta robusticidade confere uma vantagem a esta espécie que a permite de permanecer viva e de manter suas atividades metabólicas em diversos ambientes. Estas características são importantes para suas aplicações em indústrias alimentícias, particularmente, na indústria de queijos PPC; e em tecnologia, para a síntese de biomoléculas e/ou para

alimento/saúde (probióticos), todas estas aplicações serão apresentadas com detalhes nesta seção.

Tabela	1:	Ação	microbiana	nos	alimentos	fermentados	(tabela	adaptada	de
http://mastervrv.free.fr/S3/BA/Cours/BA-FERMENTATION.pdf).									

Efeitos sob o produto	Exemplos	Microrganismos implicados				
	Coagulação do leite	Diversas bactérias lácticas				
Modificação da textura	Textura aveludada em laticínios fermentados	Bactérias lácticas (dextranes)				
	Formação de olhaduras em queijos	Bactérias propiônicas				
	Gosto amanteigado (diacetil)	Lactococcus lactis diacetylactis				
Aromatização	Gosto típico de iogurte (acetaldeído e ácido láctico)	Lactobacillus delbrueckii bulgaricus				
	Desenvolvimento do gosto de cacau	Diversas bactérias				
Coloração	Escurecimento da massa de queijos PPC* por propionato de cálcio	Bactérias propiônicas				
	Acidificação do repolho	Lactobacillus				
Alteração do gosto	Desacidificação do vinho	Bactérias malolácticos				
Estabilização microbiana	Acidificação	Bactérias lácticas e acéticas				

\*PPC: queijos de massa cozida e prensada.

Tabela	2:	Principais	fermentadores	para	a	indústria	de	laticínios	(tabela	adaptada	de
http://mastervrv.free.fr/S3/BA/Cours/BA-FERMENTATION.pdf).											

Fermentadores	Aplicações
Lactobacillus bulgaricus Streptococcus thermophilus Bifidobacterium sp.	Iogurtes
Lactobacillus acidophilus	Leite fermentado
Lactobacillus caucasius	Kéfir (bebida fermentada)
Streptococcus cremosnis Streptococcus lactis Streptococcus diacetylactis Leuconostoc cremonis Lactobacillus cremonis Lactobacillus lactis	Etapas iniciais (coalhamento) da maioria dos queijos Queijos não afinados (massa fresca) Manteiga
Propionibacterium freudenreichii ssp shermanii Propionibacterium freudenreichii ssp. freudenreichii	Maturação de queijos PPC*

\*PPC: queijos de massa cozida e prensada.

Nos paragráfos seguintes, iremos apresentar algumas informações sobre a importância de *P. freudenreichii* em aplicações industriais:

- J Queijos PPC → as bactérias propiônicas são necessárias para a maturação dos queijos PPC. Sua sobrevivência é necessária para garantir o sabor e a tipicidade destes queijos.
- ) Biomoléculas → bactérias propiônicas pemitem a produção de diversas biomoléculas de interesse industrial, como por exemplo, o ácido propiônico e a vitamina B12, produtos obtidos durante o metabolismo destas bactérias.
- ) Probióticos → a manutenção da atividade metabólica de *P. freudenreichii* nos produtos probióticos é de grande importância, uma vez que esta bacéria pode atuar no trato gastrointestinal, como por exemplo, através da imunomodulação. Para isto, é necessário que as bactérias sobrevivam por longos períodos passando por etapas de transformação e armanezamento, sem grandes alterações na viabilidade e atividade esperada para o produto final.

#### 2.1. Indústria de queijos PPC ("pasta cozida e prensada")

Os queijos são mundialmente produzidos e são classificados de acordo com suas características, tais como: consistência da pasta, teor de gordura, tipo de leite utilizado, tipo de coagulação, tipo de casca e tempo de cura. *P. freudenreichii* é utilizada em conjunto com outras culturas bacterianas, principalmente, na fabricação de queijos Emmental. Este grupo é o terceiro segmento mais importante na indústria de queijos, sendo chamado de "pasta cozida e prensada" (PPC, do francês: pâte pressée cuite), e composto por outros queijos, como o queijo Comté. A denominação PPC corresponde aos queijos, os quais a coagulação e desidratação são extensas e que, em seguida, são prensados no momento da moldagem para remover a quantidade máxima de soro do leite, nomeados também de "queijos duro". Após a etapa de coagulação e moldagem, a coalhada é "cozida" em uma etapa de alto aquecimento, acima de 50°C durante 30 à 60 minutos no soro, o que permite de obter, após a desidratação e prensagem, um elevado extrato seco e uma conservação destes queijos por mais tempo.

De acordo com o *Centre National Interprofessionnel de l'Economie Laitière*, em 2014, a França produziu mais de 260.000 toneladas de Emmental. A região do *Grand Ouest* da França, contemplando a Bretanha, País de Loire e Baixa Normandia, representa dois terços desta produção (CNIEL, 2014). Destaque para a região da Bretanha que é a primeira em produção de leite de vaca et de Emmental na França (Figura 2).



Figura 2: Produção de leite de vaca no território francês (*La géographie du lait em 2013 – Enquête Annuelle Latière*).

Estes produtos são muito importantes para a economia de lactícinios da França, particularmente o Emmental, que é um grande representante deste setor (Tabela 3).

Queijos PPC	2011	2012	2013	
Emmental	256.980	253.225	252.892	
Comté AOP	58.114	57.561	59.459	
Beaufort AOP	5.044	4.919	4.787	
Gruyère IGP	-	2.679	2.929	
Autres PPC	-	8.913	7.938	
Total PPC	333.258	327.298	328.006	

Tabela 3: Produção de queijos PPC em toneladas (L'Enquête Annuelle Latière).

Para compreender o processo de fabricação destes queijos PPC, a tabela 4 resume algumas etapas importantes e a figura 3, ilustra todo o processo.

#### Tabela 4: Etapas para a fabricação de queijos.

Etapas	Descrição
Pasteurização	Aquecimento do leite a temperaturas elevadas, esta etapa visa a eliminação de microorganismos indesejáveis. Porém, esta etapa não ocorre para queijos feito com leite cru ou pasteurizado.
Coagulação	Esta etapa visa a concentração da proteína do leite, podendo ser divida em duas etapas: (I) ácida e (II) enzimática. A primeira ocorre pela atividade da flora microbiana do leite, ou pela adição do extrato de renina (coalho) ou femento. Já a coagulação ácida ocorre a partir da produção de ácido láctico por bactérias, ou pela adição de ácidos orgânicos ao leite.
Moldagem	A coalhada obtida no processo anterior é colocada em diferentes moldes, conforme o queijo desejado.
Desidratação	O objetivo desta etapa é separar o soro de leite da coalhada, o que prolonga sua conservação. Alguns queijos podem ser consumidos após este processo.
Salga	Os queijos frescos resultantes são desenformados e salgados, com um sal fino ou no banho de salmoura saturada. Isto garante uma ação anti-séptica, melhoria na conservação e também adiciona o sabor aos queijos.
Maturação	Esta final pode demorar dias ou até mêses de acordo com a variedade do queijo. Estes serão armazenados em "caves", local o qual terão a temperatura e umidade controlada.



Figura 1: Processos de transformação de queijos PPC (http://www.produits-laitiers.com/). O processo de fabricação dos queijos PPC, inicia-se pela coleta do leite por meio de máquinas automatizadas em fazendas de gado leiteiro. Em seguida, este leite é refrigerado e analisado, de acordo com as normas de segurança do consumidor, visando garantir um produto final de boa qualidade. Este leite é transportado em caminhões isotérmicos até a indústria de laticínios, onde começam todas as etapas de tratamento e fabricação de queijos.

O Emmental é um queijo de peso, cujo as unidades pesam entre 60 e 120 kg (grande em volume de peso). É necessário 12 litros de leite para obter 1kg deste queijo (CNIEL,2015). Durante o processo de maturação, muitos parâmetros variam, tais como: temperatura, pH e atividade de água. *P. freudenreichii* exerce um papel importante nesta maturação (Figura 4), produzindo acetato e propionato através da fermentação, que estão envolvidos no sabor do Emmental, e CO<sub>2</sub> responsável pela "abertura do queijo" por meio da formação de olhaduras na massa (Langsrud and Reinbold 1973; Meurice *et al.* 2004).



Figura 4: P. freudenreichii em um queijo Emmental (©INRA).

*P. freudenreichii* desempenha também um papel importante na composição do aroma e sabor em alguns lactícinios, dentre eles o Emmental. Isto porque durante o processo de maturação prolongada deste queijo, *P. freudenreichii* garante a ocorrerrência das reações de lipólise. A lipólise contribui para a formação de sabores e é realizada através de esterases, as quais são capazes de efetuar à hidrólise dos triacilglicerídeos do leite liberando ácidos graxos de cadeia curta, de cadeia média e de cadeia longa (Figura 5). Estes últimos é que conferem aos queijos suas diferentes sensações gustatórias, como por exemplo, sabor forte, picante e/ou salgado (Berdagué *et al.* 1990).



Figura 5: Esquema da reação de hidrólise de triacilglicerídeos.

Estudos como o de Dherbécourt e colaboradores, identificaram 23 esterases (*putative esterases*) em *P. freudenreichii*, potencialmente envolvidas neste processo de maturação. Esta identificação foi realizada a partir da combinação de métodos clássicos e do estudo da genômica desta bactéria (Dherbécourt *et al.* 2008). Através da abordagem genômica, as ferramentas bioinformáticas e os métodos utilizados foram: (i) busca automática de esterases por meio de homologia, (ii) busca por motivos protéicos no InterProScan, (iii) busca manual por homologia e (iv) busca manual por motivos específicos através do Prosite. Estes métodos foram essenciais para uma primeira triagem de enzimas de interesse. Assim, 12 genes candidatos que codificam as potenciais esterases apresentando os motivos GXSXG característicos de / hidrolase foram selecionados e clonados em *E. coli*. Testes zimográficos também foram realizados no intuito de caracterizar a atividade destas esterases. Assim, foi constatado que as 12 enzimas, em *P. freudenreichii*, desempenham papéis impotantes na lipólise no queijo Emmental.

Sabemos que *P. freudenreichii* sobrevive a diversos ambientes e estresses de maneira linhagem-dependente. A importância deste fenômeno atrae estudos sobre as estratégias que permitem a sobrevivência e manutenção da atividade metabólica por longos períodos. Assim, no intuito de explorar estes mecanismos de sobrevivência à longo prazo, estudos que associam experimentos microbiológicos (*in vitro*) relacionado com aplicações industriais e biotecnológicas são de uma grande importância. Estes estudos serão descritos nas próximas seções.

#### 2.2. Otimização da produção de vitamina B12

Em 1911, o termo "vitamina" foi proposto por Casimur Funk para designar as aminas que são essencias para a vida (vita-amina). Posteriormente, esta nomenclatura começou a ser utilizada também para substâncias orgânicas essenciais, sem valor energético mas ativas em pequenas quantidades, contribuindo para a manutenção do metabolismo de um organismo (Florent 1986; Vandamme 1992).

A vitamina B12, também denominada de cobalamina ou cianocobalamina, foi isolada com primazia em 1948, tendo sido sintetizada somente em 1973. Isto ocorreu devida a dificuldade de obtenção do produto, e sua síntese necessita de diversas etapas de processamento até o rendimento final da molécula (Vandamme 1989). Como esta vitamina está implicada em processos essenciais para a regulação do metabolismo de animais e seres humanos, como a formação e maturação de hemácias e a manutenção do sistema nervoso, estratégias alternativas que visam a otimização de sua produção são de grande interesse.

Neste contexto, a obtenção de vitamina B12 utilizando microorganismos tem se tornado uma solução promissora para o mercado industrial. Dentre as bactérias produtoras, destaca-se a *P. freudenreichii*, devido ao seu bom rendimento de vitamina B12. Assim, uma concentração de 5,12 mg/L pode ser obtida no queijo após 168h de fabricação (Marwaha, Sethi and Kennedy 1983). Estudos visando aprimorar a produção de vitamina B12 em *P. freudenreichii* mostraram que é possível alcançar um rendimento de até 6,35 mg/L após 192h de fermentação (García Hernández and Pérez Mendoza 1983).

Novas abordagens tem sido propostas para a biossíntese de vitamina B12 ao decorrer das últimas décadas. A obtenção de vitamina B12 por meio da fermentação de *P. freudenreichii* foi analisada, no trabalho de Paio e colaboradores, de acordo com o seu desempenho sob a expressão de genes, como: *cobA*, *cbiL*F e *cbi*EGH, envolvidos na síntese de vitamina B12, poderia aumentar a produção de vitamina B12 (Piao *et al.* 2004). Outros estudos com o mesmo objetivo, como o trabalho de Murooka e colaboradores, expressaram e amplificaram os genes do operon *cob* e *cbi*, para o aumento da biossíntese de cobalaminas durante o processo de fermentação (Murooka *et al.* 2005).

Para otimizar a produção de vitamina B12, um outro estudo por Wang e colaboradores com enfoque no efeito do ácido propiónico e do 5,6dimetilbenzimidazole durante a biossíntese desta vitamina no processo de cultivo de *P*. *freudenreichii* (Wang *et al.* 2015). Foi visto que a concentração de ácido propiônico varia da fase inicial (10-20 g/L) à fase tardia (20-30 g/L). Isto conduz a um feed-back negativo para a biossíntese de vitamina B12, mas que pode ser alterado com a inclusão de 5,6-dimetilbenzimidazole (Wang *et al.* 2015). Assim, um controle e uma adaptação detalhada das condições de cultivo bacteriano são essenciais para aobtenção de grandes quantidades de vitamina B12.

#### 2.3. Obtenção do ácido propiônico

O ácido propiônico é um ácido graxo de cadeia curta, saturado, de massa molecular 74,08 g/mol, também conhecido como ácido propílico, ácido propanóico ou  $C_3H_6O_2$  (GESTIS Substance Database – disponível em http://gestis-en.itrust.de/nxt/gateway.dll/gestis\_en/012590.xml?f=templates\$fn=default.htm\$3.0) (Figura 6).



#### Figura 6: Representação da fórmula química do ácido propiônico.

Este ácido é amplamente utilizado em setores industriais, como: petroquímicos, cosméticos, plásticos, herbicidas e conservantes. Neste contexto, a biologia sintética é um setor inovador que envolve a biologia e a engenharia, com o intuito de desenvolver novas funções e sistemas, por exemplo, a utilização de uma determinada via bioquímica para a obtenção de um dado material por meio de uma fonte renovável (Parizzi *et al.* 2012).

A fermentação é um dos processos mais utilizados na indústria para a obtenção de diversos produtos e subprodutos, entre eles, o ácido propiônico. Atualmente, estratégias alternativas visando a obtenção deste produto foram desenvolvidas utilizando diferentes fontes de carbono, tais como a fermentação da cana-de-açúcar (sacarose), estas estratégias buscam uma redução no custo e uma melhoria na produção deste ácido. As bactérias propiônicas são capazes de sintetizar o ácido propiônico e elas representam uma fonte interessante para tal utilidade. Assim, estudos que visam a fisiologia e metabolismo de Propionibacterias foram realizados, dentre outros, no intuito de identificar as melhores espécies e as melhores linhagens para uma utilização industrial (Thierry *et al.* 2011).

A fermentação de lactato, através do metabolismo inicial de *P. freudenreichii*, gera produtos como acetato, propionato e  $CO_2$ , que desempenham papéis variados e importantes para as indústrias alimentares e biotecnológicas (Meurice *et al.* 2004). Por exemplo, o propionato de cálcio é um produto oriundo do ácido propiônico, tendo a capacidade de atuar como conservante, sendo capaz de combater bactérias desfavoráveis e fungos em alimentos.

A partir destas observações sobre o ácido propiônico que Stowers e colaboradores avaliaram 17 linhagens do gênero *Propionibacterium* e definiram parâmetros optimais de produção e de rendimento de ácido propiônico em fermentador. Com base nestes parâmetros, linhagens das espécies de *P. acidipropionici* e *P. freudenreichii* produziram altas quantidades de ácido propiônico (Stowers, Cox and Rodriguez 2014).

Porém, ainda é difícil a obtenção de um bom rendimento de ácido propiônico a partir de Propionibacteria, uma vez que a geração de subprodutos do seu metabolismo, como ácido succínico, ácido láctico, e ácido acético diminui o rendimento do ácido propiônico (Stowers, Cox and Rodriguez 2014). Neste intuito, é importante a realização de estudos para aumentar a eficácia da fabricação deste produto. Esta produção lança oportunidades em setores como o de química ambiental, através da produção de propileno ( $C_3H_6$ ), precursor de ácido propiônico, um dos hidrocarbonetos utilizados na indústria petroquímica. A polimeração do propileno forma o polipropileno, um composto muito utilizado em diversas indústrias para gerar combustíveis, embalagens descartáveis, produtos têxteis, eletrodomésticos e mercado automobilístico (Ammar *et al.* 2014). Assim, bactérias que sintetizam o ácido propiônico a um alto rendimento são objetos de interesse crescente, devido ao desenvolvimento da química ambiental e de um mercado econômico favorável para produtos orgânicos.

#### 2.4. Probiótico

Como falado anteriormente, diversas bactérias, em especial bactérias lácticas e propiônicas, tem sido alvo de interesse devido as suas propriedades probióticas. Um exemplo disso é *Propionibacterium freudenreichii* que é amplamente estudada por sua capacidade de inibir microrganismos indesejáveis, modular a microbiota do cólon através da produção de 1,4-di-hidroxi-2-naftóico (DHNA), um fator bifidogênico (Pérez Chaia, Zarate and Oliver 1999), e à sobreviver a diferentes estresses (Isawa *et al.* 2002; Okada *et al.* 2013), em particular aos estresses encontrados durante a passagem pelo trato gastrointestinal (Jan *et al.* 2001; Leverrier *et al.* 2003a; Saraoui *et al.* 2013a). *P. freudenreichii* é também investigada por desencadear a apoptose em células cancerígenas do cólon, aderir à células epiteliais do intestino e muco, estimulando assim atividades imunomodulatórias e a proliferação celular no hospedeiro (Ouwehand *et al.* 2002; Zarate, Chaia and Oliver 2002). Nesta seção, iremos apresentar uma pequena síntese de estudos que abordam as diversas propriedades probióticas atribuídas à *P. freudenreichii*.

#### 2.4.1. Histórico e definição do termo probiótico

Embora os benefícios de alguns alimentos fermentados para a saúde sejam estudados há muitos anos (trabalhos de Metchnikoff no início do século XX), o termo probiótico teve origem em 1965, quando Lilly e Stillwell, relacionaram a sobrevivência de determinados microrgansimos com o efeito benéfico de seus produtos (Lilly and Stillwell 1965). Atualmente, os probióticos são definidos como microrganismos vivos que se administrados de maneira adequada, promovem benefícios à saúde do hospedeiro (Bron, van Baarlen and Kleerebezem 2012). Devida a sua importância, publicações no PubMed vem ganhando destaque ao longo dos útimos anos (Figura 7).



#### Publicações de probióticos (Pubmed)

Figura 7: Publicações no PubMed com as palavras-chave "probiotics-bacteria" ao decorrer dos últimos anos.

Além disso, para a utilização de probióticos em humanos e para que as reivindicações de saúde sejam aceitas, são necessários rigorosos testes *in vitro* e *in vivo*, os quais visam garantir a qualidade e segurança destes microrganismos antes de suas possíveis aplicações tanto na prevenção ou no tratamento de enfermidades quanto em suplementos alimentares. Algumas espécies bacterianas como *Lactobacillus paracasei*, *L. johnsonii*, *L. fermentum*, *L. plantarum*, *L. rhamnosus*, *Bifidobacterium longum* e *P. freudenreichii* são objetos de estudo por apresentarem capacidades que promovem a melhoria da saúde humana, incluindo o controle e a restauração do equilíbrio da microbiota intestinal (Parvez et al. 2006; Cousin et al. 2012; Santos Rocha et al. 2012; Luerce et al. 2014).

#### 2.4.2. Controle da microbiota

Bactérias propiônicas podem exercer efeitos na microbiota dos animais e/ou de seres humanos, seja inibindo microrganismos não-desejáveis e/ou aumento a proliferação de populações de *Bifidobacterium* sp., bactérias que exercem funções importantes no trato digestivo (Picard *et al.* 2005). O seu aumento apresenta uma série de vantagens fisiológicas e clinicas para a saúde humana e/ou animal, tais como: (i) acelera o tempo de passagem pelo trato gastrointestinal; (ii) exerce o efeito de barreira, já que produz substâncias que bloqueiam os sítios de ligação de patógenos; (iii) exerce efeito sobre o sistema imunológico e em doenças do trato-gastrointestinal; (iv) reduz o

risco de diarréia por infecção por rotavírus; (v) produz nitrosaminas, nitritos e aminas heterocíclicas; (vi) exerce efeitos sobre a proliferação de células intestinais e (vii) atua na carcinogênese do cólon (Picard *et al.* 2005). Além de Bifidobactérias, existem outras espécies bacterianas dominantes que colonizam diversas porções do trato gastrointestinal e que são de grande importância para o controle da microbiota intestinal. Estas espécies são mostradas na Figura 8.



Figura 8: Espécies microbianas dominantes em porções do trato gastrointestinal. Figura adaptada do artigo "Microbial Influences in Inflammatory Bowel Diseases" (Sartor 2008).

A microbiota intestinal exerce um papel importante no sistema imunológico dos seres humanos e/ou animais e esta é, geralmente, adquirida durante o primeiro ano de vida logo após ao nascimento. A microbiota pode ser influenciada por fatores ambientais externos, como por exemplo, a ingestão de produtos lácteos na dieta (Bernstein 2014; Geuking *et al.* 2014).

A composição da microbiota intestinal apresenta um interesse fundamental quando associada a patologias do trato gastrointestinal. Diversos estudos buscam também um melhor entendimento destas comunidades microbianas e seus efeitos de modulação do sistema imune e, em geral, da saúde do hospedeiro (Sartor 2008; Hooper and Macpherson 2010; de Vos and de Vos 2012; Vangay *et al.* 2015; Velasquez-Manoff 2015).

#### 2.4.3. Imunomodulação

A imunomodulação é uma propriedade importante de bactérias probióticas, permitindo a indução e regulação de respostas imunes, sendo útil na prevenção e no tratamento de câncer, de doenças auto-imunes e no controle de microrganismos indesejáveis (Ouwehand *et al.* 2009; Sathish *et al.* 2013).

A associação de bactérias propiônicas à bactérias lácticas et bifidobactérias podem exercer efeitos benéficos em patologias do trato gastrointestinal em crianças e idosos. Neste contexto, bactérias propiônicas agem indiretamente favorecendo o crescimento e a manutenção de concentrações elevadas de bifidobactérias durante um período de tratamento (Mantere-Alhonen 1995). As bactérias propiônicas parecem somente promover um efeito probiótico através da imunomodulação (Cousin et al. 2010; Le Marechal et al. 2015). O efeito imunomodulador de linhagens probióticas é comumente associado a sua capacidade de adesão ao epitélio intestinal. Esta adesão permite a permanência destas bactérias por mais tempo no trato digetivo, durante o trânsito intestinal. Esta capacidade de adesão pode ser um critério de seleção de probióticos e algumas linhagens de P. freudenreichii desencadearam uma série de estudos neste campo (Ouwehand et al. 2002). A implicação de proteínas de superfície no papel imunomodulatório destas bactérias foi testado através da caracterização proteômica "da superfície" de P. freudenreichii. Foi demonstrado também que algumas proteínas de superfície, extraídas de linhagens de P. freudenreichii e purificadas, foram capazes de reproduzir, in vitro, uma resposta anti-inflamatória observada com uma linhagem completa (Le Maréchal et al. 2015).

#### 2.4.4. Apoptose induzida por ácidos graxos de cadeia curta

As bactérias propiônicas são conhecidas por sua capacidade de produzir ácidos graxos de cadeia curta (SCFA – short chain fatty acids), entre eles, o propionato e o acetato que são os principais produtos da fermentação de *P. freudenreichii*. Foi observado que estes SCFA apresentam efeitos interessantes sob a fisiologia das células epiteliais intestinais (Jan *et al.* 2002; Pérez Chaia and Zarate 2005).

Os SCFA, e principalmente o butirato, constituem uma fonte de energia privilegiada, um verdadeiro "combustível" para as células epitaliais intestinais. Em 1995, Hague e colaboradores demonstraram que estes mesmos SCFA, por exemplo, o butirato de sódio, são também capazes de induzir a apoptose em cancerígenas do cólon (Hague *et al.* 1995). Posteriormente, o trabalho de Jan e colaboradores em 2002, demonstra que *in vitro* as linhagens de *P. acidipropionici* e *P. freudenreichii* são também capazes de induzir a apoptose em linhagens de células humanas de câncer colorretal, como: HT29 e Caco2. Este efeito também foi observado em linhagens celulares humanas de câncer de cólo de útero (HeLa). Os SCFA induzem sinais característicos de apoptose, tais que: (i) perda do potential transmembranar das mitocôndrias, (ii) geração da cromatina ao nível nuclear. O mecanismo geral de apoptose através de SCFA é esquematizada na Figura 9. Além disso, esta atividade pode ser exercida *in vivo*, já que as linhagens se mantém vivas e metabolicamente ativas dentro do intestino humano (Jan *et al.* 2002).



Figura 9: Indução da apoptose a partir de SCFAs produzidos por bactérias propiônicas no cólon.

Além disso, foi proposto que as bactérias propiônicas poderiam complementar o arsenal de opções terapêuticas disponíveis na prevenção e no tratamento de câncer de cólon (Lan *et al.* 2008). Recentemente, esta mesma equipe de pesquisa propôs uma nova formulação de um leite fermentado com algumas linhagens de *P. freudenreichii*, visando uma dieta preventiva ou um suplemento na alimentação de pessoas que apresentam pré-disposição a este tipo de câncer (Cousin *et al.* 2012).

#### 2.4.5. Molécula ácido 1,4-di-hidroxi-2-naftóico (DHNA)

A molécula ácido 1,4-di-hidroxi-2-naftóico (DHNA) apresenta um acentuado efeito bifidogênico, uma vez que favorece o crescimento de bifidobactérias, propriedade importante para um prebiótico. Os prebióticos são hidratos de carbono de cadeia curta não-digeríveis por enzimas digestivas em seres humanos. Assim, o crescimento de *P. freudenreichii*, que produz DHNA, influencia diretamente no crescimento de bifidobactérias, fazendo de *P. freudenreichii* uma bactéria que possui atividades pro e pre-bióticos. Esta bactéria contribui também para o equilíbrio da flora intestinal, favorecendo o crescimento de bactérias benéficas. De outra maneira, o DHNA permite reduzir a inflamação no cólon (Quigley, Hudson and Englyst 1999; Okada *et al.* 2006). *In vivo*, a administração de DHNA provoca alterações do sistema imune intestinal de camundongos, modificando a resposta imune inata em células intestinais (Okada *et al.* 2013).

Assim, DHNA é uma molécula promissora, uma vez que pode ser utilizada no tratamento de colite. Em um modelo de colite em camundongos, sua administração provoca uma melhoria na histologia do epitélio intestinal, uma supressão da migração de linfócitos e da síntese de interleucinas pró-inflamatórias (Okada *et al.* 2013). Neste contexto, pesquisas suplementares devem ser realizadas objetivando uma otimização de DHNA para aplicações terapêuticas e para demonstrar que as quantidades produzidas *in situ* por *P. freudenreichii* poderiam conduzir aos mesmos efeitos.

#### 2.4.6. Capacidade de adaptação

Esta subseção é destinada a capacidade de sobrevivência de *P. freudenreichii* e sua capacidade de adaptação a diferentes estresses. Esta será dividida em duas partes: tolerância ao estresse térmico e ao estresse digestivo. A tolerância a estes estresses é uma propriedade muito importante em bactéria propiônica para sua manutenção e sua viabilidade, otimizando assim sua utilização como probióticos (Leverrier *et al.* 2003a).

#### **N** Tolerância ao estresse térmico:

As bactérias propiônicas sobrevivem a uma ampla gama de temperaturas. No caso das etapas da maturação de queijos PPC, estas suportam temperaturas baixas (4°C), médias (30°C) e altas (52-53°). Estas temperaturas podem variar de acordo com as etapas do processamento e o tipo de queijo a ser produzido. Para o queijo Gruyère tradicional, há uma etapa de cozimento que submete as bactérias a uma temperatura bem elevada, entre 52-53°C durante 30 minutos. A termotolerância à 55°C de diferentes linhagens utilizadas na fabricação Gruyère foi estudada através da abordagem proteômica e os perfis obtidos por eletroforese 2D revelaram diferentes respostas de acordo com as linhagens termotolerantes e termossensíveis (Anastasiou et al. 2006). Em espécies termotolerantes, proteínas associadas ao processo de fermentação propiônico, metabolismo de aminoácidos, nucleotídeos e fosforilação oxidativa foram superexpressas. Em particular, as chaperonas moleculares e os compostos antioxidantes de tiol são mais induzidas pelo calor. Entretanto, observou-se que para a adaptação térmica as vias de transdução de sinal, biossíntese e manutenção da parede celular contribuíram para a manutenção do processo de fermentação (Anastasiou et al. 2006). Outros estudos abordando bactérias propiônicas relataram também que chaperonas moleculares e proteínas de choque térmico são induzidas em muitas situações de estresse (Jan et al. 2001; Leverrier et al. 2003a). Esta diferença na expressão de proteínas de diversas linhagens de bactérias propiônicas, ressaltam o argumento de que existem fenótipos diferentes dentro da mesma espécie (Anastasiou et al. 2006). Esta forma de tolerância é muito importante em P. freudenreichii para manter a sobrevivência bacteriana e garantir a fluidez do processo de fabricação de queijos do tipo PPC. Ela consititue também uma vantagem incontestável para sua utilização como probiótico, se associamos sua resistência aos estresses digestivos (Leverrier et al. 2003a).

#### **N** Tolerância ao estresse digestivo:

Para que um probiótico exerça sua função é necessário a manter sua atividade metabólica e sobrevivência após a passagem pelo trato gastrointestinal. O trabalho de Lan e colaboradores demonstraram esta capacidade em *P freudenreichii*, demonstrando a viabilidade destas bactérias no trato digestivo e no cólon de camundongos (Lan *et al.* 2007). Este estudo confirmou também a tolerância ao pH ácido de algumas linhagens específicas de *P. freudenreichii*, como já relatado em estudos anteriores sobre a resistência ao estresse ácido (Jan, Rouault and Maubois 2000) e também a tolerância aos sais biliares (Warminska-Radyko, Laniewska-Moroz and Babuchowski 2002). Experimentos de proteômica, através de eletroforese 2D, revelaram 24 proteínas induzidas durante a adaptação de *P. freudenreichii* em meio contendo sais biliares. Entre elas, estavam proteínas de transdução de sinal e fatores sigma alternativos (Leverrier *et al.* 2003a).

Para avaliar a atividade metabólica de *P. freudenreichii*, uma RT-PCR quantitativa utilizando uma sonda *TaqMan* e selecionando o RNAm codificante para a enzima transcarboxilase foi desenvolvido. Este método não invasivo e eficiente foi utilizado para provar a sobrevivência e a atividade metabólica de *P. freudenreichii* no trato intestinal de camundongos (Hervé *et al.* 2007). Outros métodos buscando avaliar a atividade metabólica de *P. freudenreichii* no trato intestinal de camundongos (Hervé *et al.* 2007). Outros métodos buscando avaliar a atividade metabólica de *P. freudenreichii* no queijo foram desenvolvidos no intuito de caracterizar as linhagens quanto à sua sobrevivência. Neste contexto, um método de RT-PCR quantitativa utilizando *SyberGreen* foi optimizado para a quantificação de células e de marcação de RNAm (gènes 16S, *tuf, groL*) de *P. freudenreichii* e de *Lactobacillus paracasei*, durante o processo de fabricação de queijos PPC (Falentin *et al.* 2010c). Este método permitiu quantificar a viabilidade celular, bem como instiga estudos para investigar como ocorre a sobrevivência de *P. freudenreichii*.

#### 3. Genoma de Propionibacterium freudenreichii

Através das tecnologias de sequenciamento de próxima geração (NGS), foi possível a obtenção do sequenciamento de vários genomas de bactérias propiônicas, o que permitiu um melhor conhecimento biológico da espécie. Estes genomas apresentam, em geral, um alto conteúdo GC, em torno de 67% (Falentin *et al.* 2010a) e algumas linhagens podem apresentar plasmídeos em sua composição (Luijk *et al.* 2002). Dados provenientes do sequenciamento podem ser utilizados em estudos comparativos e/ou evolutivos visando compreender a genômica, ao nível estrutural e ao nível funcional deste microorganismo.

Em 2010, o primeiro genoma completo de *P. freudenreichii* foi o da linhagem CIRM-BIA1, o qual foi sequenciado utilizando a metodologia tradicional de Sanger (Falentin *et al.* 2010a). A montagem inicial deste genoma foi realizada sem sequências repetitivas, usando o pacote do software Phred/Phrap/Consed (www.phrap.org) (Vallenet *et al.* 2008; Falentin *et al.* 2010a). Posteriormente, para o fechamento de *gaps*, foi necessário a utilização do *kit GC sequence finishing (GEhealthcare)* e o *kit transposition bombs*, técnicas que permitiram a obtenção do primeiro genoma completo de *P. freudenreichii* pela equipe do STLO - INRA de Rennes, France (Falentin *et al.* 2010a). Este genoma possui um cromossomo circular de 2,7 Mb, alto conteúdo de GC (67%) e encontra-se disponível no banco de dados *Genbank*, com o número de acesso FN806773 (Falentin *et al.* 2010a).

Ao longo dos últimos anos, outros 21 genomas de P. freudenreichii foram sequenciados por esta equipe. O sequenciamento destes genomas foi realizado através da plataforma Illumina, utilizando bibliotecas paired-ends, com exceção da linhagem CIRM-BIA 9 que utilizou single reads. A montagem destes 21 genomas foi obtida através da abordagem de novo, e em seguida, os scaffolds foram ordenados por comparação ao genoma de referência completo CIRM-BIA1 (Falentin et al. 2010a; Loux et al. 2015a). Os detalhes da montagem destes 21 genomas são apresentados na Tabela 5. Todos os genomas foram anotados na plataforma AGMIAL, que é uma interface gráfica para anotação de genomas completos e incompletos de procariotos. Os 21 genomas foram depositados no banco de dados do EMBL-EBI e estes dados econtram-se disponíveis em http://www.ebi.ac.uk/ena/data/view/Taxon:Propio nibacterium % 20freudenreichii) (Loux et al. 2015a). Dentre estes 21 genomas, inclui P. freudenreichii CIRM-BIA138, uma das linhagens utilizadas durante este trabalho de tese para a análise do RNA-Seq.

Porém, a maioria destes novos genomas de *P. freudenreichii* são ainda incompletos, ou seja, em estado *draft*, isto se deve a dificuldade na etapa de montagem de genomas associada a presença de um elevado número de sequências repetitivas (transpóson).

Análises bioinformáticas foram realizadas em diferentes genomas de *P. freudenreichii*, com o intuito de identificar os genes envolvidos: (i) na resistência a diferentes pH e a presença de sais biliares; (ii) na síntese de vitaminas e aminoácidos e (iii) no armazenamento de carbono. Estas análises demonstram que ela é uma bactéria versátil, adotada de um amplo arsenal de genes para sobreviver em ambientes desfavoráveis (Falentin *et al.* 2010a; Loux *et al.* 2015a). Estes dados contribuíram para a explicação da capacidade de *P. freudenreichii* manter uma atividade metabólica em condições de estresse e de sobreviver a longo prazo. Isto explica em parte sua viabilidade em ambientes que simulam o trato gastrointestinal em humanos e animais. Estas análises genômicas foram também utilizadas para identificar genes potencialmente associados a sua atividade imunomodulatória.

Outros estudos revelaram a presença de mutações pontuais e ilhas genômicas, variando de uma linhagem a outra. Isto demonstra a plasticidade cromossômica desta espécie, que resulta, provavelmente, de transferências horizontais, duplicações, transposições e mutações acumuladas. A análise destas variações entre linhagens, acoplada a uma caracterização fenotípica completa de sua capacidade a metabolizar açucares permitiu estabelecer uma ligação entre esta plasticidade e o efeito de que determinadas linhagens de *P. freudenreichii* utilizam, preferencialmente, determinadas fontes de carbono. Estas análises abrem também novas perspectivas para o melhoramento de seleção e utilização destas bactérias para a indústria de queijos e de probióticos (Loux *et al.* 2015a).

O genoma completo de *P. freudenreichii* CIRM-BIA1 foi também analisado e comparado ao de *P. acnes*, no intuito de identificaros fatores que diferenciam estas duas espécies próximas. Esta análise também foi realizada para identificar o que estava subjacente as capacidades probióticas de determinadas linhagens de *P. freudenreichii* (Falentin *et al.* 2010a).

As análises genômicas estruturais baseadas no DNA são completadas à análises funcionais através de estudos sobre o RNA, proteínas e produtos do metabolismo desta bactéria, respectivamente, abordagens de transcriptômica, proteômica e metabolômica. Isto será ilustrado no segundo capítulo da seção de resultados deste manuscrito de tese: utilização da genômica funcional para a compreensão do fenômeno de sobrevivência à longo prazo em *P. freudenreichii*.

Number in CIRM-BIA	Locus_tag	Isolated from (date)	Bioproject	Raw reads accession	Read	Read	Sequencin	Number of	Cumulativ	N50	Number of	Number of
collection (Rennes,				number	quantity	length	g depth	scaffolds	e contig		N	complete
France)								>1000 pp	length		segments	genes
9	PFCIRM9	Emmental cheese (1989)	PRJEB6427	ERS638416	26 013 151	37	370	204	2 533 825	21 887	0	2 308
118	PFCIRM118	Gruyère cheese (1973)	PRJEB6428	ERS638410	51 926 392	37	739	41	2 583 496	193 767	59	2 325
119	PFCIRM119	Gruyère cheese (1973)	PRJEB6430	ERS638409	58 084 132	37	827	46	2 644 402	128 788	91	2 422
121	PFCIRM121	Swiss cheese (1937)	PRJEB6431	ERS638414	52 432 380	37	746	59	2 561 530	86 244	68	2 304
122	PFCIRM 122	NCIB (1992)	PRJEB6432	ERS638421	54 950 090	37	782	78	2 614 705	73 741	70	2 348
123	PFCIRM123	Morbier cheese (1992)	PRJEB6438	ERS638423	54 523 668	37	776	47	2 604 827	114 519	78	2 377
125	PFCIRM125	Emmental cheese (1992)	PRJEB6434	ERS638418	54 868 488	37	781	43	2 511 448	238 430	59	2 272
127	PFCIRM127	Emmental cheese (1992)	PRJEB6435	ERS638428	55 178 920	37	785	57	2 567 344	81 280	53	2 330
129	PFCIRM 129	Emmental cheese (1992)	PRJEB4826	ERS638425	52 269 870	37	744	59	2 591 314	123 180	59	2 338
134	PFCIRM134	NA	PRJEB6441	ERS638419	50 523 582	37	719	47	2 635 396	153 798	152	2 318
135	PFCIRM135	ewe raw milk (1994)	PRJEB6442	ERS638422	50 649 408	37	721	47	2 620 043	114 923	75	2 343
138	PFCIRM138	Emmental cheese (1992)	PRJEB6433	ERS638424	58 100 372	37	827	72	2 573 580	80 515	110	2 304
139	PFCIRM 139	Emmental cheese (1992)	PRJEB6436	ERS638426	60 700 312	37	864	45	2 589 117	106 963	53	2 362
456	PFCIRM456	raw milk Raclette cheese (1992)	PRJEB6445	ERS638427	49 094 398	37	699	38	2 502 068	131 281	61	2 253
508	PFCIRM 508	Gruyère (1973)	PRJEB6429	ERS638411	50 259 834	37	715	47	2 578 004	141 315	93	2 302
512	PFCIRM 512	raw milk Morbier cheese (1994)	PRJEB6439	ERS638413	63 808 750	37	908	45	2 580 419	153 592	57	2 349
513	PFCIRM 513	egyptian ras cheese (1995)	PRJEB6440	ERS638415	62 230 134	37	886	48	2 659 761	179 407	54	2 416
514	PFCIRM 514	hay (1994)	PRJEB6443	ERS638417	51 481 488	37	733	51	2 521 626	121 592	66	2 285
516	PFCIRM516	nepalian yack cheese (2000)	PRJEB6444	ERS638420	61 823 266	37	880	62	2 630 828	125 877	205	2 342
527	PFCIRM 527	Fribourg cheese (1992)	PRJEB6437	ERS638412	69 806 362	37	993	63	2 576 369	104 864	57	2 344
1025	PFCIRM 1025	Emmental cheese (1992)	PRJEB6446	ERR738371	61 323 626	81	1 910	104	2 699 685	62 523	19	2 308

Tabela 5: Genomas depositados de *P. freudenreichii* pela equipe do STLO.

#### 4. Transcriptômica de Propionibacterium freudenreichii

Nos últimos 10 anos, vários estudos abordaram a expressão gênica em *Propionibacterium*, no intuito de melhor compreender e de explicar como ocorre a regulação destes genes neste grupo bacteriano. Estes dados transcriptômicos são depositados em bancos de dados, como o EMBL-EBI (http://www.ebi.ac.uk/genomes/), criados para permitir o acesso público, fornecendo dados de genoma e transcriptoma de diferentes bactérias. Estes trabalhos nos auxiliam a compreender como as bactérias propiônicas se adaptam e regulam a expressão de genes em resposta a diferentes condições ambientais.

Espécies como *P. acnes* e P. *freudenreichii* são de grande importância neste gênero, uma vez que *P. acnes* é comensal e faz parte da microbiota da pele; enquanto que *P. freudenreichii*, como falado anteriormente, é de interesse industrial de laticínios no setor de queijos PPC e de interesse biotecnológico para a síntese de biomoléculas e por suas propriedades probióticas. Os primeiros estudos de *P. freudenreichii* foram realizados com as técnicas de microarranjo e RT-qPCR. Estes estudos demonstraram a sobrevivência destas bactérias à temperaturas variadas durante o processo de maturação de queijos PPC, assim como sua adaptação metabólica (*in situ*) no cólon de porcos (Dalmasso *et al.* 2012a; Saraoui *et al.* 2013a). A fim de ampliar os conhecimentos a cerca da sobrevivência desta bactéria, utilizamos o RNA-Seq, uma abordagem transcriptômica mais compleeta, associada à uma análise metabolômica direcionada. Nós apresentaremos na seção II de resultados deste manuscrito, um artigo a respeito da análise de *P. freudenreichii* CIRM-BIA 138, integrando seus dados transcriptômicos e metabolômicos obtidos *in vitro*.

A associação destes dados a níveis de RNA e bioquímica, aos dados proteômicos já obtidos de *P. freudenreichii*, permitirá de abordar o estudo funcional desta bactéria e nos esclarecerá estas capacidades de adaptação e de sobrevivência (Figura 10).



Correlação dos dados genômicos, transcriptômicos e proteômicos de P. freudenreichii

Figura 10: Esquema de correlação dos dados de genoma, transcriptoma e proteoma para a compreensão do metaboloma desta bactéria.

#### 5. Principais vias metabólicas de Propionibacterium freudenreichii

As bactérias propiônicas obtém sua energia através de diversos substratos de carbono através da fermentação propiônica. Em *P. freudenreichii*, a associação dos ciclos de Krebs e de Wood-Werkman são características da espécie.

O ciclo de Krebs, denominado também de ciclo do ácido cítrico ou de ciclo dos ácidos tricarboxílicos (TCA - termo em inglês), corresponde a reações metabólicas que ocorrem no citoplasma da célula bacteriana. Este ciclo é uma via anfipática apresentando etapas de (i) degradação dos metabólitos para a geração de energia –ATP – (via catabólica), e (ii) produção de precursores para a biossíntese (via anabólica) (Nelson *et al.* 2008).

Em *P. freudenreichii*, o ciclo de Wood-Werkman, sendo conhecido também como via do carbono central, é um componente central do metabolismo. Este ciclo permite a formação de propionato à partir do piruvato (Wood 1981). Durante a sua fermentação, as bactérias propiônicas são capazes de utilizar o lactato e de produzir, como citado anteriormente, determinados subprodutos de interesse para a composição do Emmental. A figura 11 ilustra o metabolismo de *P. freudenreichii*, baseado no ciclo de Wood-Werkman e sua associação ao ciclo de Krebs.



Figura 11 : Via metabólica de Wood-Werkman em *P. freudenreichii* acoplada ao ciclo de Krebs. Figura adaptada do artigo: A Temporal -omic Study of *Propionibacterium freudenreichii* CIRM-BIA1<sup>T</sup> Adaptation Strategies in Conditions Mimicking Cheese Ripening in the Cold (disponível em http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0029083).

A reação global descrita nestas bactérias segue o seguinte equilíbrio: (Crow 1986)

```
3 lactato 2 propionato + 1 acetato + 1 CO_2 + 1 H_2O (via de Fitz)
```

*P. freudenreichii* tem um papel fundamental na maturação de queijos PPC e no desenvolvimento de seus sabores (Figura 12). Também as diversas vias metabólicas resuultam na formação de compostos aromáticos, os quais foram pesquisados, especialmente, combinando estudos genômicos e metabolômicos. Diversas vias metabólicas, como o metabolismo da glicólise, da lipólise, dos aminoácidos e do armazenamento de energia foram reconstruídos integralmente para determinadas linhagens. Estes trabalhos foram alvos de muitos artigos e revisões detalhadas (Thierry *et al.* 2011).

#### Maturação de queijos



Figura 12: Esquema de processos metabólicos envolvidos na maturação dos queijos PPC visando a obtenção de compostos aromáticos.

## INTRODUCTION BIBLIOGRAPHIQUE

Cette partie du manuscrit est consacrée à la présentation de données générales sur le genre *Propionibacterium* et à celle, plus détaillée, de l'espèce *P. freudenreichii*, concernant son application dans l'industrie, ses propriétés métaboliques et probiotiques, et sa capacité à résister aux différents stress.

#### 1. Genre Propionibacterium

#### 1.1. Classification, écologie et physiologie

En 1906, Von Freudenreichi et Orla-Jensen ont rapporté le premier isolement de bactéries propioniques à partir de fromage Emmental (von Freudenreich and Orla-Jensen 1906). En 1909, le nom du genre *Propionibacterium* a été proposé par Orla-Jensen (Cummins and Johnson, 1986), en raison de la capacité de cette bactérie à produire de l'acide propionique pendant le processus de fermentation (Orla-Jensen 1909). Plus tard, en 1946, Douglas et Gunter ont proposé l'inclusion de bactéries appartenant à l'espèce *Corynebacterium acnes* au sein du genre *Propionibacterium*. Et, par la suite, en 1963, d'autres auteurs ont confirmé cette reclassification, faisant de *C. acnes* une espèce du genre *Propionibacterium* (Moore and Cato 1963).

Les bactéries du genre Propionibacterium sont des bactéries à Gram positif, appartenant à l'ordre des Actinomycétales, dans le phylum Actinomycètes. Elles sont immobiles et ne forment pas de spores. Ce genre peut être divisé en deux grands groupes selon son habitat : (i) les produits laitiers, dans lesquels on trouve les bactéries propioniques laitières ; (ii) la peau et/ou les muqueuses, sur laquelle on trouve les bactéries cutanées ou commensales. Le groupe des bactéries propioniques laitières, comprend différentes espèces, dont les plus connues sont P. freudenreichii, P. thoenii, P. jensenii et P. acidipropionici. Ces espèces ne sont pas pathogènes pour l'homme et sont généralement isolées des produits fermentés alimentaires, comme l'ensilage, la fermentation d'olives, les olives, la distillerie de rhum. Quelles études rapportent leur présence dans des échantillons de l'environnement, par exemple, des sols, et des animaux notamment dans le tractus intestinal des rats (Cummins and Johnson 1986; Mantere-Alhonen 1982; Balows 1992). D'autres espèces telles que P. cyclohexanicum (Kusano et al. 1997) et P. microaerophilum (Koussémon et al. 2001) ont été plus récemment décrites et classées dans des groupe des bactéries propioniques laitières nonpathogènes, bien qu'ayant été isolées non pas de produits laitiers, mais d'aliments
avariés ou de sous-produits. Des exemples de ces occurrences sont P. cyclohexanicum, trouvé dans le jus d'orange fermenté et P. microaerophilum, décrit dans les eaux usées provenant de pressoirs à olives. Toutefois, dans le groupe de Propionibacterium commensal ou cutané, il y a trois espèces, dont certaines comme P. acnes, P. avidum et P. propionicum, provoquent des pathologies chez l'homme. Les espèces P. *lymphophilum* et *P. granulosum* appartiennent aussi à la classification des commensales, néanmoins elles représentent un groupe distinct, étant phylogénétiquement loin des autres groupes (Meile et al. 1999).

### 1.2. La caractérisation et l'importance du micro-organisme étudié : *Propionibacterium freudenreichii*

Les conditions optimales de laboratoire pour la croissance de *P. freudenreichii* sont le milieu YEL (bouillon lactate et extrait de levure), proposé par Malik et collaborateurs (Malik, Reinbold and Vedamuthu 1968), et à une température d'incubation de 30°C. Il s'agit d'une bactérie anaérobie facultative qui peut être trouvée *in vitro* sous diverses formes (pléomorphe) : cocciforme, bifide ou ramifiée (Figure 1).



Figure 1 : Aspect pléomorphe de *P. freudenreichii* pendant le stress acide : (A) cellules non traitées; (B) cellules soumises à un pH = 2,0; (C) cellules soumises à un pH = 5,0; (D) cellules soumises à un pH = 5,0 et soumis ensuite à un pH = 2,0, visualisées par le microscopie électronique à balayage (Jan *et al.*, 2001).

Auparavant, l'espèce P. freudenreichii était divisée en deux sous-espèces : P. freudenreichii subsp. freudenreichii, qui est incapable de fermenter le lactose, mais peut réduire le nitrate ; et P. freudenreichii subsp. shermanii, qui fermente le lactose sans réduire le nitrate (Cummins and Johnson, 1986). Cependant, des études récentes montrent qu'il existe des souches de P. freudenreichii qui présentent les deux phénotypes : le lactose et le nitrate réductase positif ou le lactose et le nitrate réductase négative (de Carvalho, Gautier and Grimont 1994; Vorobjeva 1999; Dalmasso et al. 2011). En outre, dans le génome complet de P. freudenreichii CIRM-BIA 1, il a été observé que le gène lacZ, responsable de l'activité beta-galactosidase, est situé dans une région de transposase et qu'il y a des pseudogènes dans la région du génome qui code l'information nécessaire à la réduction du nitrate (Falentin et al. 2010a). Actuellement, il n'y a pas de classification des sous-espèces de P. freudenreichii au niveau phénotypique et d'autres recherches pour explorer le génome de cette bactérie doivent encore être effectuées. Ceci est d'une grande importance dans l'industrie alimentaire, puisque que ces propriétés biochimiques peuvent modifier la qualité finale d'un produit, en l'occurrence les fromages PPC.

P. freudenreichii présente une parfaite innocuité sanitaire et possède deux statuts importants pour son utilisation : le Generally Recognized As Safe (GRAS), délivré par l'Agence américaine des produits alimentaires et médicaux (Food and Drug Administration - FDA) (Mogensen et al. 2002); et la Qualified Presumption of Safety (QPS), délivré par l'Autorité européenne de sécurité des aliments (European Food Safety Authority - EFSA) (EFSA EXPERTS, 2008). Ces statuts sont approuvés par les autorités du secteur visant à contrôler la qualité des produits, ce qui garantit la sécurité et autorise l'utilisation de ces bactéries dans l'industrie alimentaire, par exemple, dans la fabrication du fromage Emmental. Ces bactéries sont très largement répandues dans la nature, on les trouve dans le tractus gastro-intestinal si elles ont été ingérées, et elles sont aussi présentes dans de nombreux aliments fermentés, du fait de leurs caractéristiques qui les rendent résistantes au pH acide (Jan, Rouault and Maubois 2000; Huang and Adams 2004; Suomalainen et al. 2006), et aux basses températures dans le processus d'affinage des fromages PPC (Dalmasso et al. 2012a). La concentration généralement rencontrée dans l'Emmental pour ces bactéries est de l'ordre de 10<sup>9</sup> UFC/g. Ces bactéries possèdent des qualités très importantes pour diverses applications dans le domaine industriel, qui seront présentées dans la section suivante.

#### 2. Applications industrielles de Propionibacterium freudenreichii

Les aliments fermentés, en particulier les produits laitiers fermentés, sont consommés par l'homme depuis plusieurs milliers d'années. Depuis le Moyen Age, la fabrication des fromages de type Emmental dans les alpages de Savoie est décrite dans des manuscrits. Au XIX<sup>e</sup> siècle, apparaissent les premiers rapports sur les "fruitières" qui sont des lieux d'exploitation et de transformation du lait en fromage, regroupant des producteurs mettant en commun le « fruit » de leur production d'été. Puis, au XX<sup>e</sup> siècle, une forte industrialisation a été observée, conduisant à une délocalisation de la production de ces fromages.

A l'instar de l'Emmental, tous les produits laitiers qui auparavant étaient produits de manière plutôt empirique ont bénéficié des progrès de la microbiologie et de la technologie alimentaire, ce qui a permis de mieux maîtriser, diversifier et sécuriser leur production.

Les principales espèces de bactéries utilisées aujourd'hui pour la production de produits laitiers fermentés appartiennent aux genres : *Bifidobacterium*, *Lactobacillus*, *Lactococcus*, *Leuconostoc*, *Streptococcus* et *Propionibacterium* (Tableaux 1 et 2). Ces genres jouent des rôles importants dans l'industrie laitière, aussi bien comme dans la synthèse de biomolécules, que comme dans le développement de produits probiotiques.

Dans ce contexte, *P. freudenreichii* joue un rôle important dans la fabrication et particulièrement dans l'affinage des fromages PPC. Cette espèce est aussi utilisée pour sa capacité à produire des bactériocines, des composés de l'acide linoléique, de l'acide propionique, du propène, du tréhalose et des vitamines du groupe B, comme par exemple, la vitamine B12 (Piao *et al.* 2004; Poonam *et al.* 2012). Certaines souches possèdent également des propriétés probiotiques et peuvent moduler la composition du microbiote intestinal en produisant des facteurs bifidogènes, ou moduler le niveau d'inflammation dans le contexte de maladies inflammatoires chroniques de l'intestin (MICI). Elles sont d'autant plus intéressantes qu'elles sont capables de résister au stress du tractus gastro-intestinal.

Comme décrit précédemment, cette bactérie présente la capacité de s'adapter à des stress de nature variable, tels que le pH, la température et les sels biliaires. Cette robustesse confère un avantage à cette espèce qui lui permet de rester viable et de maintenir des activités métaboliques dans divers environnements. Ces caractéristiques sont importantes pour ses applications dans l'industrie alimentaire, tout particulièrement dans l'industrie fromagère des PPC ; en technologie, pour la synthèse de

biomolécules et/ou pour l'aliment-santé (probiotiques), toutes ces applications seront présentées en détail dans cette section.

Effets sur le produit	Exemples	Microrganismes impliqués				
	Coagulation du lait	Plusieurs bactéries lactiques				
Modification de la texture	Texture filante ou veloutée de produits laitiers frais fermentés	Bactéries lactiques (dextranes)				
	Formation des «trous» dans le fromage	Bactéries propioniques				
Aromatisation	Goût typique du beurre (diacétyle)	Lactococcus lactis diacetylactis				
	Goût typique du yaourt (acétaldéhyde et acide lactique)	Lactobacillus delbrueckii bulgaricus				
	Développement du goût du cacao	Plusieurs bactéries				
Coloration	Brunissement de la pâte des fromages PPC* par le propionate de calcium	Bactéries propioniques				
	Acidification du chou (choucroute)	Lactobacillus				
Modification du goût	Désacidification du vin	Bactéries malolactiques				
Stabilisation microbienne	Acidification	Bactéries lactiques et acétiques				

Tableau	1:	L'action	microbienne	dans	les	aliments	laitiers	fermentés	(tableau	adapté
http://mastervrv.free.fr/S3/BA/Cours/BA-FERMENTATION.pdf).										_

\*PPC = pâte pressée cuite

 Tableau
 2 :
 Principaux
 ferments
 pour
 l'industrie
 laitière
 et
 fromagère
 (tableau
 adapté

 http://mastervrv.free.fr/S3/BA/Cours/BA-FERMENTATION.pdf).

Ferments	Applications
Lactobacillus bulgaricus Streptococcus thermophilus Bifidobacterium sp.	Yaourts
Lactobacillus acidophilus	Laits fermentés
Lactobacillus caucasius	Kéfir (boisson fermentée)
Streptococcus cremosnis Streptococcus lactis Streptococcus diacetylactis Leuconostoc cremonis Lactobacillus cremonis Lactobacillus lactis	Premières étapes (caillage) de la plupart des fromages Fromages non affinés à pâte fraîche Beurre
Propionibacterium shermanii Propionibacterium freudenreichii	L'affinage des fromages PPC*

\*PPC = pâte pressée cuite

Dans les paragraphes à suivre, nous présentons quelques données sur l'importance de *P*. *freudenreichii* dans les applications industrielles :

• Fromages PPC  $\rightarrow$  les bactéries propioniques sont nécessaires pour l'affinage des fromages PPC. Leur survie est nécessaire pour assurer la saveur et la typicité de ces fromages.

Biomolécules → bactéries propioniques permettent la production de diverses biomolécules d'intérêt industriel, comme par exemple, l'acide propionique et la vitamine B12, produits du métabolisme de ces bactéries.

• **Probiotiques**  $\rightarrow$  le maintien de l'activité métabolique de *P. freudenreichii* dans les produits probiotiques est d'une grande importance, car cette bactérie peut agir sur le tractus gastro-intestinal via, par exemple, l'immunomodulation. Pour cela, il est nécessaire que les bactéries survivent pendant de longues périodes en passant par des étapes de transformation et de stockage, sans altérations majeures sur la viabilité et l'activité visée.

#### 2.1. Industrie des fromages PPC (pâte pressée cuite)

Les fromages sont produits partout dans le monde et sont classés en fonction de leurs caractéristiques, tels que la consistance de la pâte, la teneur en matière grasse, le type de lait utilisé, le type de coagulation, le type de croûte et le temps d'affinage. *P. freudenreichii* est utilisé conjointement avec d'autres cultures de bactéries, principalement dans la fabrication du fromage Emmental. Ce groupe est le troisième segment le plus important dans l'industrie du fromage, et est appelé « pâte pressée cuite» (PPC), il comporte d'autres fromages, comme le Comté. Le terme PPC correspond aux fromages dont la coagulation et la déshydratation sont prolongées et qui sont ensuite pressés au moment du moulage pour éliminer la quantité maximum de lactosérum, on les appelle également "fromages à pâte dure". Après l'étape de coagulation et de tranchage, le caillé est « cuit » par une étape de chauffage au-dessus de 50°C pendant 30 à 60 minutes dans le sérum, ce qui permet d'obtenir, après égouttage et pressage, un extrait sec élevé et une plus longue conservation.

Selon le Centre National Interprofessionnel de l'Economie Laitière en 2014, la France a produit plus de 260.000 tonnes d'Emmental. La région Grand Ouest de la France, comprenant la Bretagne, le Pays de Loire et la Basse-Normandie, représente les deux tiers de cette production (CNIEL, 2014). La Bretagne est la première région productrice de lait de vache et d'Emmental en France (Figure 2).



Figure 1 : La production du lait de vache sur le territoire français (La géographie du lait en 2013 - Enquête Annuelle Latière).

Ces produits sont très importants pour l'économie laitière de la France, particulièrement l'Emmental, qui est le grand représentant de ce secteur (Tableau 3).

Fromages PPC	2011	2012	2013
Emmental	256.980	253.225	252.892
Comté AOP	58.114	57.561	59.459
Beaufort AOP	5.044	4.919	4.787
Gruyère IGP	-	2.679	2.929
Autres PPC	-	8.913	7.938
Total PPC	333.258	327.298	328.006

Tableau 3: La production des fromages PPC en tonnes (L'Enquête Annuelle Latière).

Pour comprendre le processus de fabrication de ces fromages PPC, le tableau 4 résume quelques étapes importantes, et la figure 3 illustre l'ensemble du processus.

Tableau 4: Les étapes de la fabrication des fromages.

Étapes	Description
La pasteurisation	Le chauffage du lait à des températures élevées. Cette étape vise à éliminer les microorganismes indésirables. Cependant, cette étape n'est pas nécessaire pour les fromages fabriqués avec du lait cru ou pasteurisé.
La coagulation	Cette étape vise à la concentration de la protéine du lait et peut être divisée en deux sous-étapes: (i) acide et (ii) présure. La première est obtenue à partir de la production d'acide lactique par les bactéries ou par l'addition d'acides organiques dans le lait. La deuxième, l'étape enzymatique, se produit à partir de l'activité de la flore microbienne du lait ou par l'ajout de l'extrait de présure.
Le moulage	Les caillés obtenus à l'étape précédente sont placés dans différents moules, en fonction du fromage désiré.
L'égouttage	Le but de cette étape est de séparer le lactosérum du lait caillé, ce qui prolonge la conservation du fromage. Certains fromages peuvent être consommés après ce processus.
Le salage	Les fromage frais sont démoulés et salés avec du sel ou de la saumure saturée en sel. Cela assure une action antiseptique et une meilleure conservation et renforce la saveur au fromage.
L'affinage	Cette étape peut prendre des jours ou des mois, selon la variété de fromage. Ceux-ci seront stockés et recevront des soins en cave, où la température et l'humidité sont contrôlées.



Figure 3 : Processus de transformation des fromages PPC (http://www.produits-laitiers.com/). Le processus de fabrication des fromages PPC commence par la collecte du lait dans les machines automatiques des exploitations laitières. Ensuite, ce lait est refroidi et analysé conformément à la réglementation pour la sécurité des consommateurs, afin d'assurer un produit final de bonne qualité. Ce lait est transporté dans des camions citernes isothermes à l'industrie laitière où commencent toutes les étapes de traitement et de fabrication du fromage.

L'Emmental est un fromage de poids dont les meules pèsent entre 60 et 120 kg. Il faut 12 litres de lait pour obtenir 1 kg de ce fromage (CNIEL, 2015). Pendant son affinage de nombreux paramètres varient : la température, le pH et l'activité de l'eau. *P. freudenreichii* joue un rôle important dans son affinage (Figure 4), produisant par fermentation de l'acétate et du propionate, qui participent la flaveur de l'Emmental, et du  $CO_2$ , responsable de l'ouverture du fromage par la formation de trous dans la pâte (les « yeux ») (Langsrud and Reinbold 1973; Meurice *et al.* 2004).



Figure 4 : P. freudenreichii dans le fromage Emmental (© INRA).

*P. freudenreichii* joue ainsi un rôle important dans la composition de l'arôme et de la saveur de certains produits laitiers, parmi lesquels l'Emmental. En effet, pendant l'affinage prolongé de ce fromage, *P. freudenreichii* assure des réactions de lipolyse. La lipolyse contribue à la formation de saveurs et est réalisée par des esterases qui sont capables d'effectuer l'hydrolyse des triacylglycérides du lait en libérant des acides gras à chaîne courte, à chaîne moyenne et à chaîne longue (Figure 5). Ceux sont eux qui donnent aux fromages leurs différentes sensations gustatives, comme par exemple, une saveur forte, épicée et/ou salée (Berdagué *et al.* 1990).



#### Figure 5 : Schéma de réaction d'hydrolyse de triacylglycérides.

Les études de Dherbécourt et collaborateurs ont identifié 23 estérases (estérases putatives) chez *P. freudenreichii*, potentiellement impliquées dans ce processus d'affinage.

Cette identification s'est faite à partir de la combinaison de méthodes classiques et de l'étude génomique de cette bactérie (Dherbécourt *et al.* 2008). Pour l'approche génomique, les outils bioinformatiques et les méthodes utilisés ont été : (i) la recherche automatique d'estérases par homologie, (ii) la recherche des motifs protéiques dans l'InterProScan, (iii) la recherche manuelle par homologie et (iv) la recherche manuelle faite par motifs spécifiques grâce à Prosite. Ces méthodes ont été indispensables pour une première sélection d'enzymes d'intérêt. Ainsi 12 gènes candidats codant pour des estérases potentielles présentant des motifs GXSXG caractéristiques des / hydrolases ont été sélectionnés et clonés dans *E. coli*. Des zymographes ont également été effectués afin de caractériser l'activité de ces estérases. L'inactivation et la complémentation des deux gènes d'estérases codant pour des protéines ancrées en surface de *P. freudenreichii* ont révélé que le gène pf279 était responsable de la lipolyse. De plus, l'analyse du génome de 21 souches de *P. freudenreichii* a montré que ce gène pouvait présenter des mutations non-sens dans quelques souches qui ne présentent pas d'activité lipolytique (Abeijon Mukdsi *et al.* 2014).

Nous avons déjà vu que *P. freudenreichii* survivait dans divers environnements et contraintes de façon souche dépendante. L'importance de ce phénomène donne de l'attrait à l'étude des stratégies qui permettent la survie et le maintien d'une activité métabolique pendant de longues périodes. Afin d'exploiter ces mécanismes de survie à long terme, les études qui associent des expériences microbiologiques (*in vitro*) en lien avec des applications industrielles et biotechnologiques sont d'une grande importance. Ces études seront décrites dans les sections suivantes.

#### 2.2. Optimisation de la production de vitamine B12

En 1911, le terme « vitamine » a été proposé par Casimur Funk pour désigner les amines qui sont essentiels à la vie (vita-amine). Plus tard, cette nomenclature a commencé à être utilisée également pour les substances organiques essentielles, sans valeur énergétique mais actives en petites quantités, contribuant à l'entretien du métabolisme d'un organisme (Florent 1986; Vandamme 1992).

La vitamine B12, aussi appelée cobalamine ou cyanocobalamine, a été isolée pour la première fois en 1948 mais elle n'a été synthétisée qu'en 1973. Elle est en effet difficile à obtenir et sa synthèse comporte plusieurs étapes pour obtenir la molécule finale (Vandamme 1989). Comme cette vitamine est impliquée dans des processus essentiels pour la régulation du métabolisme des animaux et des humains comme la formation et maturation des érythrocytes et la maintenance du système nerveux, les stratégies alternatives visant à l'optimisation de sa production sont d'un grand intérêt.

Dans ce contexte, l'obtention de la vitamine B12 en utilisant des micro-organismes est devenue une solution prometteuse pour le marché industriel. Parmi les bactéries qui en produisent, *P. freudenreichii* se distingue en raison de son bon rendement. Ainsi, une concentration de 5,12 mg/L peut être obtenue dans le fromage après 168h de fabrication (Marwaha, Sethi and Kennedy 1983). Des études d'amélioration de la production de vitamine B12 chez *P. freudenreichii* ont montré qu'il est possible de parvenir à un rendement allant jusqu'à 6,35 mg/L après 192h de fermentation (García Hernández and Pérez Mendoza 1983).

De nouvelles approches ont donc été proposées pour la biosynthèse de la vitamine B12 au cours des décennies. L'obtention de la vitamine B12 par fermentation de *P. freudenreichii* a été analysée dans le travail de Piao et collaborateurs, qui ont montré qu'en jouant sur l'expression des gènes *cobA*, *cbi*LF et *cbi*EGH, impliqués dans la synthèse de la vitamine B12, on pouvait augmenter la production de vitamine B12 (Piao *et al.* 2004). D'autres études ayant le même but, comme le travail de Murooka et collaborateurs, ont exprimé et amplifié les gènes de l'opéron *cob* et *cbi*, pour augmenter la biosynthèse des cobalamines pendant le processus de fermentation (Murooka *et al.* 2005).

Pour optimiser la production de vitamine B12, une autre étude a été réalisée par Wang et collaborateurs se concentrant sur l'effet de l'acide propionique et du 5,6diméthylbenzimidazole au cours de la biosynthèse de cette vitamine au cours de la culture de *P. freudenreichii (Wang et al.* 2015). Il a été montré que la concentration d'acide propionique varie de la phase initiale (10 à 20 g/L) à la phase tardive (20 à 30 g/L). Ceci conduit à une rétroaction négative sur la biosynthèse de la vitamine B12, mais peut être modifié par l'addition de 5,6-dimetilbenzimidazole (Wang *et al.* 2015). Ainsi, un contrôle et une adaptation fine des conditions de culture sont essentiels pour l'obtention de grandes quantités de vitamine B12.

#### 2.3. L'obtention d'acide propionique

L'acide propionique est un acide gras àchaîne courte, saturé, de masse moléculaire 74,08 g/mol, également connu sous le nom d'acide propanoïque ou  $C_3H_6O_2$  (GESTIS Substance Database – disponible en http://gestisen.itrust.de/nxt/gateway.dll/gestis\_en /012590.xml?f=templates\$fn=default.htm\$3.0) (Figure 6).

Figure 6 : Représentation de la formule chimique de l'acide propionique.

Cet acide est largement utilisé dans les industries suivantes: la pétrochimie, les produits cosmétiques, les plastiques, les herbicides et les agents de conservation. Dans ce contexte, la biologie synthétique est un secteur innovant, impliquant la biologie et l'ingénierie, qui permet de développer de nouvelles fonctions et de nouveaux systèmes, par exemple, l'utilisation d'une voie biochimique spécifique pour obtenir un matériau à travers une source d'énergie renouvelable (Parizzi *et al.* 2012).

La fermentation est l'un des procédés les plus utilisés dans l'industrie pour l'obtention de divers produits et sous-produits, parmi lesquels l'acide propionique. Actuellement, plusieurs stratégies visant l'obtention de ce produit ont été développées en utilisant différentes sources de carbone, telles que la fermentation du sucre de canne (saccharose), ces stratégies recherchent une réduction des coûts et une amélioration de la production de cet acide. Les bactéries propioniques sont capables de synthétiser l'acide propionique et elles représentent une source d'intérêt pour cet usage. Aussi, des études visant la physiologie et le métabolisme des Propionibacteries ont été réalisées afin, entre autres, d'identifier les meilleures espèces et les meilleures souches pour une utilisation industrielle (Thierry *et al.* 2011).

La fermentation de lactate, par le métabolisme initial de *P. freudenreichii*, génère des produits, tels que l'acétate, le propionate et le  $CO_2$ , qui jouent des rôles variés et importants pour les industries alimentaires et de biotechnologie (Meurice *et al.* 2004). Par exemple, le propionate de calcium est un produit dérivé de l'acide propionique ayant la capacité d'agir comme agent de conservation, capable de lutter contre les bactéries défavorables et les champignons dans les aliments.

C'est à partir de ces observations sur l'acide propionique que Stowers et collaborateurs ont évalué 17 souches de *Propionibacterium* et ont défini des paramètres

optimaux de production et de rendement pour l'acide propionique en fermenteur. En suivant ces paramètres, des souches des espèces *P. acidipropionici* et *P. freudenreichii* produisent des quantités élevées d'acide propionique (Stowers, Cox and Rodriguez 2014).

Cependant, il est toujours difficile d'obtenir un bon rendement d'acide propionique à partir de Propionibacteria, car la génération de sous-produits dérivés de son métabolisme, tels que l'acide succinique, l'acide lactique, l'acide acétique réduisent le rendement de l'acide propionique (Stowers, Cox and Rodriguez 2014). Dans cette optique, il est important de mener des études pour augmenter l'efficacité de la production de ce produit. Cette production trouve aussi des débouchés en chimie verte à travers de la production de propylène ( $C_3H_6$ ), précurseur de l'acide propionique, un des hydrocarbures utilisés dans l'industrie pétrochimique. La polymérisation du propylène conduit au polypropylène, un composé largement utilisé dans diverses industries pour générer des carburants, des emballages jetables, des textiles, utilisés dans les appareils électro-ménagers et l'industrie automobile (Ammar *et al.* 2014). Ainsi, les bactéries qui synthétisent l'acide propionique à fort rendement font l'objet d'un intérêt croissant, en raison de l'essor de la chimie verte et d'un marché économique porteur pour les produits bios-sourcés.

#### 2.4. Probiotique

Nous avons déjà évoqué que plusieurs bactéries, notamment des bactéries lactiques et propioniques sont l'objet d'intérêt en raison de leurs propriétés probiotiques. A ce titre, *P. freudenreichii* est largement étudié pour sa capacité à inhiber certains microorganismes indésirables, à moduler la microflore du côlon par la production de 1,4-dihydroxy-2-naphtoïque (DHNA), un facteur bifidogène, (Pérez Chaia, Zarate and Oliver 1999) et à survivre à différents stress (Isawa *et al.* 2002; Okada *et al.* 2013), notamment les stress rencontrés pendant le transit au sein du tube digestif (Jan *et al.* 2001; Leverrier *et al.* 2003a; Saraoui *et al.* 2013b). *P. freudenreichii* est également investigué à déclencher l'apoptose dans les cellules cancéreuses du côlon, à adhérer aux cellules épithéliales intestinales et au mucus, stimulant ainsi des activités immunomodulatrices et la prolifération cellulaire chez l'hôte (Ouwehand *et al.* 2002; Zárate *et al.* 2002). Dans cette section, nous présenterons une courte synthèse des études portant sur les diverses propriétés probiotiques attribuées à *P. freudenreichii*.

#### 2.4.1 L'histoire et la définition du terme probiotique

Bien que les bénéfices de certains aliments fermentés pour la santé soient étudiés depuis longtemps (travaux de Metchnikoff au début du XX<sup>e</sup> siècle), l'origine du terme probiotique date de 1965, lorsque Lilly et Stillwell ont associé la survie de certains microorganismes à l'effet bénéfique de leurs produits (Lilly and Stillwell 1965). Actuellement, les probiotiques sont définis comme des microorganismes vivants qui, lorsqu'ils sont administrés d'une manière appropriée, apportent des effets positifs sur la santé de l'hôte (Bron, van Baarlen and Kleerebezem 2012). En raison de leur importance, les publications sur PubMed les concernant ont pris de l'ampleur au cours des dernières années (Figure 7).



### Les publications sur les probiotiques

Figure 7 : Publications dans PubMed avec les mots clés « probiotics bacteria » au cours des dernières années.

Cependant, pour une utilisation des probiotiques chez l'homme et pour que des allégations santé soient acceptées, il est nécessaire d'effectuer des essais rigoureux *in vitro* et *in vivo*, qui visent à démontrer la qualité et la sécurité de ces organismes avant leurs applications dans la prévention ou le traitement de maladies et la supplémentation alimentaire. Quelques espèces bactériennes comme *Lactobacillus paracasei*, *L. johnsonii*, *L. fermentum*, *L. plantarum*, *L. rhamnosus*, *Bifidobacterium longum* et *P. freudenreichii* font l'objet d'étude sur leurs capacités à améliorer la santé humaine, notamment par le contrôle et la restauration de l'équilibre du microbiote intestinal (Parvez et al. 2006; Cousin et al. 2012; Santos Rocha et al. 2012; Luerce et al. 2014).

#### 2.4.2 Contrôle du microbiote

Les bactéries propioniques peuvent avoir un effet sur la microflore des animaux et/ou des humains en inhibant des microorganismes indésirables et/ou en augmentant la prolifération des populations de *Bifidobacterium* sp., des bactéries qui jouent un rôle positif sur le tube digestif (Picard *et al.* 2005). Leur augmentation présente un certain nombre d'avantages physiologiques et cliniques pour la santé humaine et/ou animale, tels que : (i) accélérer le temps de passage à travers le tractus gastro-intestinal ; (ii) exercer un effet de barrière, car elles produisent des substances qui bloquent les sites de liaison de pathogènes ; (iii) exercer des effets sur le système immunitaire et dans les maladies du tractus gastro-intestinale ; (iv) réduire le risque de diarrhée lors d'infection à rotavirus ; (v) exercer des

effets sur la prolifération des cellules intestinales et (vi) empêcher la carcinogenèse du côlon (Picard *et al.* 2005). En plus des Bifidobactéries, il existe d'autres espèces bactériennes dominantes qui colonisent différentes parties du tractus gastro-intestinal et qui sont d'une grande importance pour le contrôle du microbiote intestinal. Ces espèces sont présentées dans la Figure 8.



Figure 8 : Espèces microbiennes dominantes dans les différentes parties du tractus gastro-intestinal. Figure adaptée de l'article "Influences microbiennes dans les maladies inflammatoires chroniques de l'intestin" (Sartor 2008).

Le microbiote intestinal joue un rôle important dans le système immunitaire chez les humains et/ ou les animaux et il est généralement acquis au cours de la première année qui suit la naissance. Il peut être influencé par des facteurs environnementaux externes, tels que la consommation de produits laitiers dans l'alimentation (Bernstein 2014; Geuking *et al.* 2014).

La composition du microbiote intestinal présente un intérêt fondamental de par ses liens avec certaines pathologies gastro-intestinales. De nombreuses études cherchent ainsi à améliorer notre compréhension de ces communautés microbiennes et de leur effet de modulation du système immunitaire et la santé de l'hôte en général (Sartor 2008; Hooper and Macpherson 2010; de Vos and de Vos 2012; Vangay *et al.* 2015; Velasquez-Manoff 2015).

#### 2.4.3. Immunomodulation

L'immunomodulation est une propriété importante des bactéries probiotiques, permettant l'induction et la régulation de la réponse immunitaire. Elle est aussi utile pour la prévention ou le traitement du cancer, des maladies auto-immunes et dans le contrôle des microorganismes indésirables (Ouwehand et al. 1999; Sathish et al. 2013).

L'association de bactéries propioniques à des bactéries lactiques et des bifidobactéries peut avoir des effet bénéfiques sur des pathologies gastro-intestinales chez les enfants et les personnes âgées. Dans ce cadre, les bactéries propioniques agiraient indirectement en favorisant la croissance et le maintien à des concentrations élevées des bifidobactéries au cours de la période du traitement (Mantere-Alhonen 1995). Les bactéries propioniques seules semblent aussi pouvoir exercer un effet probiotique à travers une immunomodulation (Cousin et al. 2010; Le Maréchal et al. 2015). L'effet immunomodulateur des souches probiotiques est souvent associé à leur capacité d'adhésion à l'épithélium intestinal. Cette adhésion leur permet de se maintenir plus longtemps dans le tractus digestif, lors du transit. Cette capacité d'adhésion peut être un critère de sélection de probiotiques et certaines souches de P. freudenreichii ont fait l'objet d'études dans ce domaine (Ouwehand et al. 2002). L'implication des protéines de surface dans le rôle immunomodulateur de ces bactéries a été testée par la caractérisation protéomique du « surfaçome » de P. freudenreichii. Il a ainsi été montré que certaines protéines de surface, extraites de souches de P. freudenreichii et purifiées, étaient capables de reproduire, in vitro, une réponse anti-inflammatoire observée avec la souche complète (Le Maréchal et al. 2015).

#### 2.4.4. Apoptose induite par les acides gras à chaîne courte

Les bactéries propioniques sont connues pour leur capacité à produire des acides gras à chaîne courte (SCFA pour short chain fatty acids), tels que le propionate et l'acétate qui sont les principaux produits de fermentation de *P. freudenreichii*. Il a été observé que ces SCFA ont des effets intéressants sur la physiologie des cellules épithéliales intestinales (Jan *et al.* 2002; Pérez Chaia and Zarate 2005).

Les SCFA, et principalement le butyrate, constituent une source d'énergie privilégiée, un véritable carburant, pour les cellules épithéliales intestinales. En 1995, Hague et colaborateurs ont montré que ces mêmes SCFA, par exemple, le butyrate de sodium, sont également capables au contraire d'induire l'apoptose dans les cellules cancéreuses du colon (Hague *et al.* 1995). Par la suite, les travaux de Jan et collaborateurs en 2002, ont montré *in vitro* que des souches de *P. acidipropionici* et *P. freudenreichii* sont elles aussi capables d'induire l'apoptose dans des lignées de cellules humaines de cancer colorectal, telles que HT29 et Caco2. Cet effet a aussi été observé sur des lignées cellulaires humaines de cancer du col de l'utérus (HeLa). Les SCFA induisent des signes caractéristiques de l'apoptose, tels que : (i) la perte du potentiel transmembranaire des mitochondries, (ii) la génération d'espèces réactives de l'oxygène, (iii) d'induction de la cascade caspase-3 et (iv) la condensation de la chromatine au niveau nucléaire. Le mécanisme général de l'apoptose via les SCFA est schématisé sur la Figure 9. Par ailleurs, cette activité pourrait s'exercer in vivo car ces souches restent vivantes et métaboliquement actives dans l'intestin humain (Jan *et al.* 2002).



Figure 9 : Induction de l'apoptose à partir de SCFAs produites par les bactéries propioniques dans le côlon.

Il a ainsi été proposé que les bactéries propioniques pourraient venir compléter l'arsenal d'options thérapeutiques disponibles dans la prévention ou le traitement du cancer du colon (Lan *et al.* 2008). Plus récemment, la même équipe a proposé une nouvelle formulation d'un lait fermenté avec certaines souches de *P. freudenreichii*, visant une alimentation préventive ou un supplément dans l'alimentation de personnes qui sont prédisposées à ce type de cancer (Cousin *et al.* 2012).

#### 2.4.5. Molécule acide 1,4-dihydroxy-2-naphtoïque (DHNA)

Le composant 1,4-dihydroxy-2-naphtoïque (DHNA) présente un effet bifidogène marqué (i.e., il favorise la croissance des Bifidobactéries) et constitue de ce fait un prébiotique. Les prébiotiques sont généralement des oligosaccharides non digérés par les enzymes digestives chez l'homme. Ainsi, la croissance de *P. freudenreichii*, qui produit du DHNA, influe directement sur la croissance des bifidobactéries, faisant de *P. freudenreichii* une bactérie combinant des activités pro- et pré-biotiques. Elle contribue ainsi à l'équilibre de la flore intestinale, en favorisant le développement de bactéries bénéfiques. Le DHNA permettrait par ailleurs de réduire l'inflammation du colon (Quigley, Hudson and Englyst 1999; Okada *et al.* 2006). *In vivo*, l'administration de DHNA entraîne aussi des modifications du système immunitaire intestinal chez la souris, en modifiant la réponse immunitaire innée chez les cellules intestinales (Okada *et al.* 2013).

De même, le DHNA est une molécule prometteuse qui pourrait être utilisée dans le traitement de la colite. Dans un modèle de colite induite chez la souris, son administration entraîne une amélioration dans l'histologie de l'épithélium instestinal, une suppression de la migration des lymphocytes et de la synthèse d'interleukines pro-inflammatoires (Okada *et al.* 2013). Dans ce contexte, des recherches supplémentaires doivent être effectuées visant à une optimisation du DHNA pour des applications thérapeutiques et pour montrer que les quantités produites in situ par *P. freudenreichii* pourraient aboutir aux mêmes effets.

#### 2.4.6. Capacité d'adaptation

Cette sous-section fait le point sur la capacité de survie de *P. freudenreichii* et sa capacité d'adaptation à différents stress. Elle sera divisée en deux parties: la tolérance au stress thermique et au stress digestif. La tolérance à ces stress est une propriété très importante chez les bactéries propioniques pour leur maintien et leur viabilité, ce qui optimise leur utilisation comme probiotiques (Leverrier *et al.* 2003a).

#### **Ñ** Tolérance au stress termique:

Les bactéries propioniques survivent à une large gamme de températures. Dans les étapes d'affinage des fromages PPC, elles résistent autant à basse température (4°C), qu'à moyenne (30°C) et haute température (52-53°C). Ces températures varient en fonction des différentes étapes du processus de production et du type de fromage. Pour le fromage Gruyère traditionnel, il y a une étape de cuisson soumettant les bactéries à une température très élevée, entre 52-53°C, pendant 30 minutes. La thermotolérance à 55°C de différents souches utilisées dans la fabrication de Gruyère a été étudiée par une approche protéomique et les profiles obtenus en électrophorèse 2D ont révélé différentes réponses selon que les souches sont thermotolérantes ou thermosensibles (Anastasiou et al. 2006). Dans les espèces thermotolérantes, les protéines associées au processus de la fermentation propionique, au métabolisme des acides aminés, aux nucléotides et à la phosphorylation oxydative étaient surexprimés. En particulier, les chaperons moléculaires et les composés antioxydants de type thiol sont induits par la chaleur. Toutefois, il a aussi été observé que, lors de l'adaptation thermique, les voies de la transduction du signal, la biosynthèse et le maintien de la paroi cellulaire contribuent à maintenir le processus de fermentation (Anastasiou et al. 2006). D'autres études portant sur les bactéries propioniques ont également rapporté que les chaperons moléculaires et des protéines de choc thermique sont induites dans de nombreuses situations de stress (Jan et al. 2001; Leverrier et al. 2003a). Cette différence dans l'expression des protéines des diverses souches de bactéries propioniques renforce l'argument selon lequel il y a différents phénotypes au sein de la même espèce (Anastasiou et al. 2006). Cette forme de tolérance est importante chez P. freudenreichii pour maintenir la survie bactérienne et assurer le bon déroulement du processus de fabrication des fromages de type PPC. Elle constitue en outre un avantage indéniable pour son utilisation comme probiotique, si on y associe sa résistance aux stress digestifs (Leverrier et al. 2003b).

#### **N** Tolérance au stress digestif:

Pour qu'un probiotique exerce sa fonction, il faut maintenir son activité métabolique et sa survie après son passage à travers le tractus gastro-intestinal. Le travail de Lan et collaborateurs a démontré cette capacité chez *P. freudenreichii* en démontrant la viabilité des bactéries dans le tractus digestif et dans le colon de la souris (Lan *et al.* 2007). Cette étude a également confirmé la tolérance au pH acide de certaines souches spécifiques de *P. freudenreichii*, comme cela avait déjà été rapporté dans les études précédentes portant sur la résistance au stress acide (Jan, Rouault and Maubois 2000), mais aussi sur la tolérance aux sels biliaires (Warminska-Radyko, Laniewska-Moroz and Babuchowski 2002). Des expériences de protéomique, utilisant l'électrophorèse 2D, ont montré 24 protéines induites pendant l'adaptation de *P. freudenreichii* dans un milieu contenant des sels biliaires. On compte parmi elles des protéines de transduction du signal et des facteurs sigma alternatifs.

Pour évaluer l'activité métabolique de *P. freudenreichii*, une RT-PCR quantitative utilisant une sonde *TaqMan* et ciblant l'ARNm codant pour l'enzyme transcarboxylase a été développée. Cette méthode non-invasive et efficace a été utilisée pour prouver la survie et l'activité métabolique de *P. freudenreichii* dans le tractus intestinal des souris (Hervé *et al.* 2007). D'autres méthodes cherchant à évaluer l'activité métabolique de *P.freudenreichii* dans le fromage ont aussi été développées dans le but de caractériser ces souches pour leur survie. Dans ce contexte, une méthode de RT-PCR quantitative utilisant du *SyberGreen* a été optimisée pour la quantification de cellules et d'ARNm marqueurs (gènes 16S, tuf, groL) de *P. freudenreichii* et de *Lactobacillus paracasei*, pendant le processus de production des fromages PPC (Falentin *et al.* 2010). Cette méthode a permis de quantifier la viabilité cellulaire, et d'aller plus loin dans les études concernant la survie de *P. freudenreichii*.

#### 3. Génomique de Propionibacterium freudenreichii

Grâce aux technologies de séquençage de nouvelle génération (NGS), il a été possible d'obtenir le séquençage de plusieurs génomes de bactéries propioniques, ce qui a conduit à une meilleure connaissance biologique de l'espèce. Ces génomes présentent, en général, une teneur élevée en GC, environ 67% (Falentin *et al.* 2010a) et certaines souches peuvent héberger en outre des plasmides (Luijk *et al.* 2002). Les données provenant du séquençage peuvent être utilisées dans les études comparatives et/ou évolutives visant à comprendre la génomique, au niveau structurel et au niveau fonctionnel de ce microorganisme.

En 2010, le STLO a publié le premier génome complet de *P. freudenreichii* (souche CIRM-BIA1) qui a été séquencée en utilisant la méthode traditionnelle de Sanger (Falentin *et al.* 2010a). L'assemblage initial de ce génome a été effectué sans séquences répétitives, en utilisant le logiciel Phred/Phrap/CONSED (www.phrap.org) (Vallenet *et al.* 2008; Falentin *et al.* 2010a). Ensuite, pour la clôture des *gaps*, il a été nécessaire d'utiliser le *kit sequence GC finishing* (GEhealthcare) et le kit *transposition bombs* (Falentin *et al.* 2010a). Ce génome présente un chromosome circulaire de 2,7 Mb, de haute teneur en GC (67%) et est disponible dans la base de données *Genbank*, avec le numéro d'accès FN806773 (Falentin *et al.* 2010a).

Depuis, 21 autres génomes de *P. freudenreichii* ont été séquencés par l'équipe. Leur séquençage a été réalisé par la méthode Illumina, en utilisant la bibliothèque *paired-ends*, sauf pour la souche CIRM-BIA 9 qui a utilisé la bibliothèque *single reads*. L'assemblage de ces 21 génomes a été mené par une approche *de novo*, puis les *scaffolds* ont été ordonnés par comparaison avec le génome de référence complet CIRM-BIA1 (Falentin *et al.* 2010a; Loux *et al.* 2015a). Les détails de l'assemblage de ces 21 génomes sont présentés dans le Tableau 5. Tous les génomes ont été annotés sur la plate-forme AGMIAL, qui est une interface graphique pour l'annotation de génomes complets et incomplets des procaryotes. Les 21 génomes ont été déposés dans la base de données de EBI-EMBL et ces données sont disponibles en suivant le lien : http://www.ebi.ac.uk/ena/data/view/Taxon:Propionibacte rium%20freudenreichii (Loux *et al.* 2015b). Ces 21 génomes comprennent notamment celui *P. freudenreichii* CIRM-BIA138, une des souches utilisées au cours de ces travaux de thèse pour l'analyse en RNA-Seq.

Cependant, la plupart de ces nouveaux génomes de *P. freudenreichii* sont encore incomplets, à l'état de *drafts*, cela est dû à la difficulté de l'étape d'assemblage liée à la présence de nombreuses séquences répétées (transposons).

Des analyses bioinformatiques ont été réalisées sur ces différents génomes de *P. freudenreichii*, afin d'identifier les gènes impliqués dans (i) la résistance aux différents pH et à la présence des sels biliaires; (ii) la synthèse des vitamines et des acides aminés et (iii) le stockage du carbone. Ces analyses montrent qu'il s'agit d'une bactérie versatile, dotée d'un large arsenal de gènes pour survivre dans des environnements défavorables (Falentin *et al.* 2010a; Loux *et al.* 2015b). Ces données contribuent à expliquer la capacité de *P. freudenreichii* à maintenir une activité métabolique en conditions de stress et de survie à long terme. Ceci explique pour partie sa viabilité dans des environnements qui simulent le tractus gastro-intestinal chez les humains et les animaux. Ces analyses génomiques ont aussi été largement utilisées pour identifier des gènes potentiellement liés à son activité immunomodulatrice.

D'autres études ont révélé la présence de mutations ponctuelles et d'îlots génomiques, variables d'une souche à l'autre. Ceci montre la plasticité chromosomique de cette espèce, et résulte probablement de transferts horizontaux, duplications, transpositions et mutations accumulées. L'analyse de ces variations inter-souches, couplée à une caractérisation phénotypique complète de leur capacité à métaboliser les sucres a permis d''établir un lien entre cette plasticité et le fait que certaines souches de *P. freudenreichii* utilisent, préférentiellement, certaines sources de carbone. Ces analyses ouvrent aussi de nouvelles perspectives pour améliorer leur sélection et leur utilisation pour l'industrie fromagère et des probiotiques (Loux *et al.* 2015b).

Le génome complet de *P. freudenreichii* CIRM-BIA 1 a aussi été analysé et comparé à celui de *P. acnes* afin d'identifier les facteurs différenciant ces deux espèces proches. Cette analyse a aussi été menée pour cerner ce qui sous-tend les capacités probiotiques de certaines souches de *P. freudenreichii* (Falentin *et al.* 2010a).

Ces analyses génomiques structurelles basées sur l'ADN sont complétées d'analyses fonctionnelles à travers des études portant sur l'ARN, les protéines et les produits de métabolisme de cette bactérie (respectivement, des approches transcriptomique, protéomique et métabolomique). Cela est illustré dans le deuxième chapitre de la section de résultats de ce manuscrit thèse : l'utilisation de la génomique fonctionnelle pour comprendre le phénomène de la survie à long terme chez *P. freudenreichii*.

Tableau 5 : Génomes	de <i>P</i> .	freudenreichii	déposés	par le	STLO.
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Number in CIRM-BIA collection (Rennes,	Locus_tag	Isolated from (date)	Bioproject	Raw reads accession	Read quantity	Read length	Sequencin g depth	Number of scaffolds	Cumulativ e contig	N50	Number of N	Number of complete
France)					quantity		g aspin	>1000 bp	length		segments	genes
9	PFCIRM9	Emmental cheese (1989)	PRJEB6427	ERS638416	26 013 151	37	370	204	2 533 825	21 887	0	2 308
118	PFCIRM118	Gruyère cheese (1973)	PRJEB6428	ERS638410	51 926 392	37	739	41	2 583 496	193 767	59	2 325
119	PFCIRM119	Gruyère cheese (1973)	PRJEB6430	ERS638409	58 084 132	37	827	46	2 644 402	128 788	91	2 422
121	PFCIRM 121	Swiss cheese (1937)	PRJEB6431	ERS638414	52 432 380	37	746	59	2 561 530	86 244	68	2 304
122	PFCIRM 122	NCIB (1992)	PRJEB6432	ERS638421	54 950 090	37	782	78	2 614 705	73 741	70	2 348
123	PFCIRM123	Morbier cheese (1992)	PRJEB6438	ERS638423	54 523 668	37	776	47	2 604 827	114 519	78	2 377
125	PFCIRM125	Emmental cheese (1992)	PRJEB6434	ERS638418	54 868 488	37	781	43	2 511 448	238 430	59	2 272
127	PFCIRM127	Emmental cheese (1992)	PRJEB6435	ERS638428	55 178 920	37	785	57	2 567 344	81 280	53	2 330
129	PFCIRM 129	Emmental cheese (1992)	PRJEB4826	ERS638425	52 269 870	37	744	59	2 591 314	123 180	59	2 338
134	PFCIRM134	NA	PRJEB6441	ERS638419	50 523 582	37	719	47	2 635 396	153 798	152	2 318
135	PFCIRM135	ewe raw milk (1994)	PRJEB6442	ERS638422	50 649 408	37	721	47	2 620 043	114 923	75	2 343
138	PFCIRM138	Emmental cheese (1992)	PRJEB6433	ERS638424	58 100 372	37	827	72	2 573 580	80 515	110	2 304
139	PFCIRM 139	Emmental cheese (1992)	PRJEB6436	ERS638426	60 700 312	37	864	45	2 589 117	106 963	53	2 362
456	PFCIRM456	raw milk Raclette cheese (1992)	PRJEB6445	ERS638427	49 094 398	37	699	38	2 502 068	131 281	61	2 253
508	PFCIRM 508	Gruyère (1973)	PRJEB6429	ERS638411	50 259 834	37	715	47	2 578 004	141 315	93	2 302
512	PFCIRM512	raw milk Morbier cheese (1994)	PRJEB6439	ERS638413	63 808 750	37	908	45	2 580 419	153 592	57	2 349
513	PFCIRM513	egyptian ras cheese (1995)	PRJEB6440	ERS638415	62 230 134	37	886	48	2 659 761	179 407	54	2 416
514	PFCIRM514	hay (1994)	PRJEB6443	ERS638417	51 481 488	37	733	51	2 521 626	121 592	66	2 285
516	PFCIRM516	nepalian yack cheese (2000)	PRJEB6444	ERS638420	61 823 266	37	880	62	2 630 828	125 877	205	2 342
527	PFCIRM 527	Fribourg cheese (1992)	PRJEB6437	ERS638412	69 806 362	37	993	63	2 576 369	104 864	57	2 344
1025	PFCIRM 1025	Emmental cheese (1992)	PRJEB6446	ERR738371	61 323 626	81	1 910	104	2 699 685	62 523	19	2 308

#### 4. Transcriptomique de Propionibacterium freudenreichii

Au cours des 10 dernières années, plusieurs études ont porté sur l'expression des gènes chez *Propionibacterium* afin de mieux comprendre et d'expliquer comment se produit leur régulation dans ce groupe bactérien. Ces données transcriptomiques sont déposées dans des bases, comme l'EMBL-EBI (http://www.ebi.ac.uk/genomes/), créées pour permettre l'accès public, qui fournissent des données du génome et du transcriptome de différentes bactéries. Ces travaux nous aident à comprendre comment les bactéries propioniques s'adaptent et régulent l'expression des gènes en réponse à différentes conditions environnementales.

Des espèces comme *P. acnes* et *P. freudenreichii* sont d'une grande importance dans ce genre, car *P. acnes* est commensale et fait partie de la flore de la peau ; quant à *P. freudenreichii*, nous avons déjà mentionné plus haut qu'elle est d'intérêt pour l'industrie laitière dans le secteur des fromages PPC et d'intérêt biotechnologique pour la synthèse de biomolécules et pour ses propriétés probiotiques. Les premières études de transcriptomiques sur *P. freudenreichii* ont été réalisées avec les techniques de puce d'ADN et RT-qPCR. Ces études ont montré la survie et l'adaptation de ces bactéries à des températures variées pendant le processus d'affinage des fromages PPC, ainsi que leur adaptation métabolique (*in situ*) dans le côlon de porcs (Dalmasso *et al.* 2012b; Saraoui *et al.* 2013b). Afin d'accroître les connaissances sur la survie de cette bactérie, nous avons utilisé le RNA-Seq, une approche transcriptomique plus exhaustive, couplé à une analyse métabolomique ciblée. Nous présenterons, dans la section II des résultats de ce manuscrit, un article concernant une analyse de la souche *P. freudenreichii* CIRM-BIA138, intégrant ses données transcriptomiques et métabolomiques obtenues *in vitro*.

L'association de ces données aux niveaux ARN et biochimique, aux données protéomiques déjà obtenues concernant *P. freudenreichii* permettra d'aborder l'étude fonctionnelle de cette bactérie et nous éclairera sur ses capacités d'adaptation et de survie (Figure 10).



Corrélation des données génomiques, transcriptomiques et protéomiques de P. freudenreichii

Figure 10 : Combinaison des données des données génomiques, transcriptomiques et protéomiques.

#### 5. Les principales voies métaboliques de Propionibacterium freudenreichii

Les bactéries propioniques tirent leur énergie de divers substrats carbonés par la fermentation propionique. Chez *P. freudenreichii*, l'association des cycles de Krebs et de Wood-Werkman sont caractéristiques de l'espèce.

Le cycle de Krebs, appelé également cycle de l'acide citrique ou cycle de l'acide tricarboxylique (TCA, en anglais), correspond à des réactions métaboliques qui se produisent dans le cytoplasme des cellules bactériennes. Ce cycle est une voie amphibolique présentant des étapes de (i) dégradation des métabolites pour générer de l'énergie – ATP – (voie catabolique) et (ii) production de précurseurs pour la biosynthèse (voie anabolique) (Nelson *et al.* 2008).

Chez *P. freudenreichii*, le cycle Wood-Werkman, également connu sous le nom de voie de carbone central, est une composante centrale du métabolisme. Ce cycle permet la formation de propionate à partir du pyruvate (Wood 1981). Au cours de la fermentation, les bactéries propioniques sont capables d'utiliser le lactate et de produire, comme cité précédemment, certains sous-produits d'intérêt pour la composition de l'Emmental. La Figure 11 illustre le métabolisme de *P. freudenreichii*, basé sur le cycle Wood-Werkman et son association au cycle de Krebs.



Figure 11 : Voie métabolique de Wood-Werkman chez *P. freudenreichii* liée au cycle de Krebs. Figure adaptée de l'article: A Temporal -omic Study of *Propionibacterium freudenreichii* CIRM-BIA1<sup>T</sup> Adaptation Strategies in Conditions Mimicking Cheese Ripening in the Cold (http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0029083).

La réaction globale décrite dans ces bactéries suit l'équilibre suivant: (Crow 1986)

3 lactate 2 propionate + 1 acetate + 1  $CO_2$  + 1  $H_2O$  (voie de *Fitz*)

*P. freudenreichii* jouant un rôle majeur dans la maturation des fromages PPC et le développement de leur flaveur (Figure 12). Aussi les diverses voies métaboliques aboutissant à la formation de composés d'arôme ont-elles été recherchées, notamment en combinant des études génomiques et métabolomiques. Diverses voies métaboliques (métabolisme de la glycolyse, de la lipolyse, des acides aminés, de stockage d'énergie) ont ainsi été reconstruites intégralement, pour certaines souches. Ces travaux ont fait l'objet de nombreuses articles et de revues détaillées (Thierry *et al.* 2011).



L'affinage des fromages

Figure 12 : Schéma des processus métaboliques impliqués dans l'affinage des fromages PPC visant l'obtention des composés aromatiques.

## **CAPÍTULO 2 : RESULTADOS**

# **CHAPITRE 2 : RÉSULTATS**

## SEÇÃO I: Caracterização do fenótipo de linhagens de *Propionibacterium* freudenreichii

SECTION I : Caractérisation duphénotype des souches dePropionibacterium freudenreichii

#### 1. Introdução

A primeira seção de resultados será apresentada na forma de artigo científico. O objetivo deste estudo foi avaliar a variabilidade entre as linhagens e de caracterizar os fenótipos das células de *P. freudenreichii* durante a sobrevivência a longo prazo.

Este manuscrito descreve, inicialmente, a caracterização fenotípica de 8 linhagens de *P. freudenreichii*, em meio de cultura sem a adição de nutriente durante um período de 11 dias, sendo os 8 últimos correspondentes à uma carência nutricional. Durante este período, cada linhagem foi monitorada por densidade óptica, enumeração de colônias (ágar) e observação morfológica das colônias. A viabilidade e integridade da membrana também foram avaliadas. Para isso, foi utilizado o *kit live/dead* acompanhado de uma observação em microscopia de epifluorescência, e nós verificamos a ausência de lise celular qPCR com primers específicos para o gene alvo *tuf*.

Após 2 dias de cultura a população atingiu 9 log<sub>10</sub> UFC/mL. Após 11 dias as 8 linhagens de *P. freudenreichii*, mantiveram uma população elevada, de aproximadamente 7.8 a 8.2 log<sub>10</sub> UFC/mL. O lactato, principal composto de carbono foi totalmente consumido entre 2 e 3 dias após a inoculação, e as bactérias tiveram que lidar com um período de estresse nutricional. A integração dos métodos utilizados permitiu comparar os níveis de sobrevivência e serviu como base para a triagem das 8 linhagens. A linhagem CIRM-BIA 138 é aquela que tem mostrado uma melhor sobrevivência à 11 dias após a inoculação. As pesquisas desta tese foram então focadas nesta linhagem.

Existem três principais mecanismos que permitem a sobrevivência bacteriana, quando as bactérias estão expostas à condições ambientais desfavoráveis: (i) a esporulação, (ii) a morte celular programada e (iii) dormência. Já está estabelecido que *P. freudenreichii* não esporula. Quanto ao mecanismo de morte celular programada, os restos celulares de parte da população lisada fornecem nutrientes para a subpopulação não lisada, permitindo assim que determinadas espécies bacterianas enfrentem as condições de carência nutricional. Os resultados de qPCR mostram uma estabilidade da população bacteriana à 9 log<sub>10</sub> e nos levam a concluir que *P. freudenreichii* CIRM-138 não usa a morte celular como mecanismo para a sobrevivência a longo prazo para manter a sua viabilidade, uma vez que a lise celular não é detectável nestas culturas.

Portanto, nossa hipótese é que esta bactéria permanece viável por causa da sua entrada em estado de dormência. Os resultados da microscopia de epifluorescência mostraram que metade das células bacterianas de todas as linhagens apresentaram membranas permeabilizadas após 11 dias de cultura. Durante o período de sobrevivência, uma perda de 1  $\log_{10}$  UFC/mL foi observada em comparação aos resultados de quantificação da população por qPCR. A comparação dos resultados de contagem de colônias e de qPCR, assim como a ausência de lise sugerem que a 11 dias de cultura, 90% da população bacteriana perderam a capacidade de formar colónias em ágar, mas que uma boa parte das células são, no entanto, viáveis. A integração dos resultados da caracterização fenotípica sugerem que esta bactéria atinge um estado VBNC no momento o qual começa o período de sobrevivência a longo prazo. Este estado é descrito em muitas bactérias Gram-positivas e Gram-negativas, sendo uma estratégia para a sobrevivência nos períodos de estresse, tais como a temperatura, o pH e a disponibilidade de nutrientes variados.

Este trabalho mostra pela primeira vez um mecanismo de dormência abordado em *P. freudenreichii*, cujas células entram em estado VBNC para garantirem sua sobrevivência a longo prazo em condições de carência nutricional. Isso aumenta o nosso conhecimento nesta espécie de lactéos e abre perspectivas para novos estudos. Novas abordagens integradas, baseadas em técnicas *omics* devem ser realizadas para compreender o mecanismo de dormência e a regulação metabólica bacteriana durante este período de sobrevivência a longo prazo em *P. freudenreichii* (ver secção - Secção II deste manuscrito de tese).

#### 1. Introduction

La première section des résultats sera présentée sous forme d'article scientifique. L'objectif de cette étude a été d'évaluer la variabilité inter-souches et de caractériser les phénotypes des cellules de *P. freudenreichii* pendant la survie à long terme.

Ce manuscrit décrit d'abord la caractérisation phénotypique de 8 souches de *P. freudenreichii*, en milieu de culture sans ajout sur une période de 11 jours dont les 8 derniers correspondent à une carence nutritionnelle. Pendant cette période, chaque souche a fait l'objet d'un suivi par densité optique, par dénombrement sur gélose et en observant la morphologie des colonies. La viabilité et l'intégrité membranaire ont aussi été évaluées. Pour cela, nous avons utilisé le *kit live/dead* suivi d'une observation en microscopie à épifluorescence et nous avons vérifié l'absence de lyse cellulaire par PCR quantitative en ciblant le gène *tuf*.

Après 2 jours de culture la population atteignait 9  $\log_{10}$  UFC/mL. Après 11 jours les 8 souches de *P. freudenreichii*, maintenaient une population élevée, d'environ 7.8 à 8.2  $\log_{10}$  UFC/mL. Le lactate, principal composé carboné étant épuisé entre 2 et 3 jours après inoculation, et les bactéries ont dû faire face à une période de stress nutritionnel. L'intégration des méthodes utilisées a permis de comparer les niveaux de survie et a servi de base au le criblage des 8 souches. La souche CIRM-BIA 138 est celle qui a montré la meilleure survie à 11 jours après inoculation. Les recherches de la thèse se sont ensuite focalisées sur cette souche.

Il y a trois mécanismes majeurs qui permettent la survie bactérienne lorsque les bactéries sont exposés à des conditions environnementales défavorables : (i) la sporulation, (ii) la mort cellulaire programmée et (iii) la dormance. Il est déjà établi que *P. freudenreichii* ne sporule pas. Lors du mécanisme de mort programmée, les débris cellulaires d'une partie de la population lysée fournissent des nutriments pour la souspopulation non-lysée, et permettent à certaines espèces bactériennes de faire face à des conditions de carence nutritionnelle. Les résultats de PCRq montrant une stabilité de la population bactérienne à 9 log<sub>10</sub> nous amènent à conclure que *P. freudenreichii* CIRM-138 n'utilise pas la mort cellulaire programmée comme mécanisme de survie à long terme pour maintenir sa viabilité, car aucune lyse cellulaire n'est détectable dans les cultures.
Nous émettons donc l'hypothèse que cette bactérie reste viable en raison de son entrée en état de dormance. Les résultats de la microscopie à épifluorescence ont démontré que la moitié des cellules bactériennes de toutes les souches présentait des membranes perméabilisées après 11 jours de culture. Pendant la période de survie, une perte de 1 log<sub>10</sub> UFC/mL a été observée par rapport aux résultats de quantification de la population par qPCR. La comparaison des résultats de dénombrement et de qPCR, ainsi que l'absence de lyse suggèrent qu'à 11 jours de culture, 90 % de la population bactérienne a perdu sa capacité à former des colonies sur gélose mais qu'une bonne partie des cellules sont néanmoins viables. L'intégration des résultats de la caractérisation phénotypique suggère que cette bactérie atteint un état VBNC au moment où commence la période de survie à long terme. Cet état est décrit chez de nombreuses bactéries à Gram positif et à Gram négatif, comme étant une stratégie pour la survie dans des périodes de stress tels que la température, le pH et la disponibilité de nutriments variés.

Ce travail montre pour la première fois qu'un mécanisme de dormance se met en place chez *P. freudenreichii*, dont les cellules entrent en état en VBNC pour assurer leur survie à long terme en condition de carence nutritionnelle. Cela accroît nos connaissances sur cette espèce laitière et ouvre des perspectives pour de nouvelles études. De nouvelles approches intégrées, basées sur des techniques *omiques* devraient être menées pour comprendre le mécanisme de dormance et la régulation métabolique bactérienne au cours de cette période de survie à long terme chez *P. freudenreichii* (voir article - section II de ce manuscrit de thèse).

# 2. Artigo 1: The long-term survival of *Propionibacterium freudenreichii* in a context of nutrient shortage.

Short running head: P. freudenreichii survival

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#### Abstract

**Aims**: *Propionibacterium freudenreichii* is an actinobacterium widely used in dairy industry during the ripening process of Swiss-type cheeses and which presents probiotic properties. PF is reportedly a hardy bacterium, able to survive during the cheese-making process and when subjected to digestive stresses. During this study the long-term survival of *P. freudenreichii* was investigated for 11 days by means of phenotypic characterization in a culture medium without the addition of any nutrients.

**Methods and Results**: For 11 days, in a non-nutrient supplemented culture medium, eight strains were monitored by measuring their optical density, counting colony-forming units (CFU) and using LIVE/DEAD staining and microscopy observation. Under these conditions, all strains displayed high survival rates in the culture medium, their culturability reaching more the 9 log<sub>10</sub> CFU/mL after two days. After 11 days, this value ranged from 7.8 to 8.2 log<sub>10</sub> CFU/mL depending on the strain, and at least 50% of the *P. freudenreichii* population displayed an intact envelope. Since lysis of part of a bacterial population may be a microbial strategy to recover nutrients, in CIRM-BIA 138 (the strain with the highest population at day 11), cell lysis was assessed by quantifying intact bacterial cells using qPCR targeting the housekeeping gene *tuf*. No lysis was observed.

**Conclusion**: Taken together, our results suggest that *P. freudenreichii* strains use a viable but nonculturable (VBNC) state to adapt to the long term survival phase.

**Significance and Impact of the study**: Assessing the viability of *P. freudenreichii* and understanding their mechanisms for survival should be of great interest regarding their potential probiotic applications.

**Keywords:** adaptation; bacteria; growth phases; culturability; long-term survival; viable but nonculturable; *Propionibacterium freudenreichii*.

#### Introduction

Propionibacterium freudenreichii is an Actinobacterium that is responsible for the opening and aroma of Swiss-type cheeses. PF is also used as a source of probiotics because PF has bifidogenic properties (Isawa et al., 2002), PF is resistant to digestive stress (Jan et al., 2000), and some strains are endowed with anti-inflammatory capabilities that could be used to prevent inflammatory bowel diseases (Okada et al., 2013; Le Maréchal *et al.*, 2015). Its metabolic activity is characterized by its ability to ferment various carbohydrates, the main product being propionic acid (Houwen et al., 1991; Loux et al., 2015). PF is reportedly a hardy bacterial species, which can stay alive and metabolically active for long periods under stressful conditions (i.e. cave-aged Swiss-type cheeses at cool temperatures) (Dalmasso et al., 2012). The genome sequence of the type-strain CIRM-BIA1 revealed the genetic basis for its hardiness (Falentin et al., 2010a). In the laboratory, PF can be stored in a spent medium at room temperature for more than 6 months and remains culturable if inoculated in a fresh medium. Actinobacteria are known to be hardy and extremely resistant to nutrient shortage (Greenblatt et al., 2004). However, except for the Mycobacterium genus which has been extensively explored (Ghodbane 2014), little is known about the mechanisms underlying the long term survival of this class.

During their growth and survival, Gram-positive and Gram-negative bacteria pass through different physiological phases (Kolter *et al.*, 1993; Hicks, 2005; Finkel, 2006; Bruno & Freitag, 2011; Wen *et al.*, 2013). In some bacterial species, conventional

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exponential and stationary phases are followed successively by the death phase and a long-term survival phase (Finkel, 2006; Helmus et al., 2012). Long-term survival (LTS) can be defined as a phase during which the bacteria develop mechanisms to adapt and remain viable, even after a long period of nutritional shortage (Finkel, 2006; Meena & Rajni, 2010; Bharati et al., 2012; Justice et al., 2014): (i) sporulation (Fimlaid and Shen, 2015), (ii) quiescence/dormancy or (iii) programmed cell death (Finkel, 2006; Rittershaus et al., 2013). During sporulation, a vegetative bacterial cell differentiates into a metabolically dormant cell-type known as a spore, which can reawaken when favourable conditions return (Fimlaid and Shen, 2015). Quiescence is characterized by decelerated or arrested growth in a viable non-replicating state (Rittershaus et al., 2013), which can evolve to a viable but nonculturable (VBNC) state (Ramamurthy et al., 2014). In programmed cell death (bacterial apoptosis), the death and lysis of part of a bacterial population may be profitable to surviving cells, which can catabolize the detritus. Cell death and lysis can be observed by monitoring the fall in the viable count using standard plating assays associated with a leakage of intracellular content that can be assessed biochemically (lactate dehydrogenase quantification) (Wilkinson et al., 1994) or using qPCR methods (Falentin et al., 2010b).

PF is a non-sporulating species, but PF is as yet unknown whether it survives by adopting a strategy of quiescence/dormancy or lysis of part of the bacterial population. During this study, we investigated the LTS (OD, culturability, membrane permeability and lysis) of eight *P. freudenreichii* strains throughout an 11-day period of culture without the addition of any nutrients, in order to investigate inter-strain variability in long-term survival capacities and to better understand the strategies used to ensure survival.

#### **Materials and Methods**

#### **Bacterial strains**

Eight *P. freudenreichii* strains (*Table 1*) were used during this work. The strains were supplied by the International Centre for Microbial Resources - Bacteria of Food Interest – (Centre International de Ressources Microbiennes - Bactéries d'Intérêt Alimentaire; INRA Rennes).

**Table 1.** Propionibacterium freudenreichii strains used during this work.

Strains of P. freudenreichil*	Source
CIRM-BIA 1	Reference strain
CIRM-BIA 9	Emmental cheese 1989 Van Niel
CIRM-BIA 119	Gruyère cheese 1973
CIRM-BIA 121	Swiss cheese 1937
CIRM-BIA 127	Emmental cheese 1992
CIRM -BIA 129 alias ITG-P20 b	Emmental cheese 1992
CIRM-BIA 138 alias ITG-P9	Emmental cheese 1992
CIRM-BIA 139 alias ITG-P23 <sup>b</sup>	Emmental cheese 1992
a Bacterial strains obtained from the Centre Inter Food Interest (Bactéries d'Intérêt Alimentaire, BIA)	national de Ressources Microbiennes (CIRM) dedicated to Bacteria of (CIRM-BIA, UMR1253, INRA Rennes, France).
b Strain CIRM-BIA (alias ITG-P20), strain CIRM-BIA supplied by Actalia (Rennes, France).	138 (alias ITG-P9) and strain CIRM-BIA 139 (alias ITG-P23) were kindly

#### **Conditions for bacterial growth**

All the strains were routinely cultured in YEL medium (pH=7.0) (Malik *et al.*, 1968) with *no agitation* and at 30°C. Each strain was grown in a different test tube for each manipulation, thus preventing the interference of oxygen in the environment and maintaining the *P. freudenreichii* strains under microaerophilic conditions. At T0, an inoculum of  $7 \log_{10}$  CFU/mL was added to all the tubes.

Growth kinetics were followed by Optical Density measurements using a Model DU 640 spectrophotometer (Beckman Coulter, Fullerton, California, USA) at 650 nm (OD<sub>650</sub>) and by counting the number of CFU (CFU/mL) using the micromethod as described previously (Baron *et al.*, 2006). For enumerations, all the *P. freudenreichii* strains were plated in YEL agar culture medium at 30°C, under microaerophilic conditions, until the visualization of colonies (6 days). In all cases, two technical replicates and two biological replicates were performed.

#### **Epifluorescence microscopy**

The eight strains were monitored for 11 days using the LIVE/DEAD® BacLight<sup>TM</sup> Bacterial Viability Kit – Molecular Probes® (*Life Technologies, USA*), as recommended by the supplier. Microscopic observations were performed at each of the time points used for the OD<sub>650</sub> and CFU/mL measurements. Briefly, SYTO9 (which stains all cells green, regardless of their membrane integrity) and propidium iodide (which stains permeabilized cells red) were mixed at a ratio of 1: 1 (5µL SYTO 9 and 5 µL propidium iodide). An 0.5µL aliquot of the mix was added to 100µL of the bacterial culture, vortexed, and incubated in the dark for 15 minutes. The bacteria were then analysed using epifluorescence microscopy with a phase contrast Nikon G11 microscope (Nikon D300) and Camera Control Pro 2 software (Nikon, Japan). For each sample, six photos (three photos with a x40 objective and three with a x100 objective) were analysed so as to visually estimate the percentage of permeabilized cells (red / red + green cells) at each time point.

#### **DNA extraction of non-lysed cells**

The DNA extraction method only enables the recovery of DNA from non-lysed cells. Samples were collected at three time points (79h, 144h and 168h post-inoculation) during the growth of *P. freudenreichii* CIRM-BIA 138, as described previously (Parayre *et al.*, 2007). Briefly, cells were harvested from 1 mL of the culture sample by centrifugation (5000g, 10 min.). The cell pellets were re-suspended in lysis buffer (Qiagen, Courtaboeuf, France) supplemented with lysozyme (Promega, France; 8mg/sample) and mutanolysin (Sigma, France; 20U/sample). These samples were incubated for 30 min. at 37°C for 1 h, before being treated with proteinase K (0.5mg/sample) prior to purification using the DNeasy Tissue Kit, as recommended by the supplier (Qiagen, Courtaboeuf, France). Three biological replicates and two technical replicates were used for each time point.

#### Quantification of bacterial cells using quantitative PCR

To determine any cell lysis of CIRM-BIA 138 after entry into stationary phase and just before entering the LTS phase, quantitative PCR (qPCR) was used to quantify the non-lysed bacterial cells throughout growth, according to the method described by Falentin et al. (2010b). Briefly, the qPCR targeted *tuf*, a gene present in a single copy in the *P. freudenreichii* chromosome, using primers designed on the *tuf* sequence (F: CGAACGAGTTCCACTGCGGGGT; R: GCAACATCGGCACCATCGGAC). Amplifications were performed exactly as described previously. Cycle thresholds (Ct) were determined after manual adjustment of the threshold within the linear part of qPCR logarithmic amplification curves. CIRM-BIA 138 samples were collected at 79, 144, and 168 hours for qPCR determination. Each experiment was carried out in triplicate (independent biological replicates). We checked by a spiking experiment that

lysed cells could not contaminate qPCR results. A sample of entire cells (8  $log_{10}$  CFU) was spiked with (9  $log_{10}$  CFU) lyzed cells (the lysis was performed as described below). Extraction and qPCR on entire cells and spiked sample gave the same Ct. The qPCR results will give a quantification of entire cells only.

#### Results

# Growth and survival rate of *Propionibacterium freudenreichii* under laboratory conditions.

The growth of the eight P. freudenreichii strains was monitored spectrophotometrically by measuring the optical density at 650 nm ( $OD_{650}$ ), and by plate counting (CFU/mL) throughout an 11 day period. Strains were inoculated at 7  $\log_{10}$  CFU/mL in fresh YEL medium. Both of the monitoring methods showed that P. freudenreichii growth curves had roughly similar profiles despite variations in terms of their final culturability at 11 days. The curves established over a prolonged incubation revealed several phases, as shown for CIRM-BIA129 and CIRM-BIA138 (Figure 1. See all growth curves in the supplemental data, Figure S1). Under the conditions applied, the cultures entered the exponential growth phase to reach a maximum  $OD_{650}$  of 3 to 4 (for CIRM-BIA 138) and a population of 9 to 9.6 log<sub>10</sub> CFU/mL between 50 h and 72 h. The OD<sub>650</sub> curves then decreased steadily for about 100h, which corresponded to entry into the stationary phase, and reached a plateau of culturability at around 168 h, for a population of 8 to 8.7 log<sub>10</sub> CFU/mL, depending on the strain (Figure 1) but always with a lower  $OD_{650}$ . The latter phase was designated the long term survival (LTS) phase. The other six P. freudenreichii growth curves were similar (Supplementary data Figure S1). In terms of culturability, all the strains survived well with a population of around 8  $\log_{10}$ CFU/mL after 11 days in the spent medium, i.e. without the addition of any nutrients (Figure 2, Table S1). Interestingly, two strains, CIRM-BIA121 and CIRM-BIA138, survived better than the six others (Figure 2) and maintained an average population of 8.2  $\log_{10}$  CFU/ mL at day 11, suggesting a strain-dependency in ability of P. freudenreichii for long-term survival. On Petri plates, colonies of P. freudenreichii maintained the same size and aspect whereas photon microscopy images revealed bacteria of the same size (not shown) but single cells in the exponential phase and groups of two to five cells in the stationary phase.



**Figure 1:** Growth curves of *P. freudenreichii* strains CIRM-BIA129 and CIRM-BIA138 determined at OD650 (upper panels), and plate counting (CFU ml<sup>-1</sup>, lower panels).



**Figure 2:** Viability of the eight *P. freudenreichii* strains (in CFU  $ml^{-1}$ ) after 11 days in the spent culture medium (YEL) based on a mean of two technical replicates and two biological replicates. Upper bars are standard deviations.

#### **Evaluation of cell envelope permeability**

LIVE/DEAD staining assays showed that the numbers of intact cells (stained green by SYTO9) remained stable (100% index) for each of the eight strains throughout the exponential and early stationary phases (Table S2). The numbers of intact cells then started decreasing at 50 or 150h post-inoculation and dropped to 80% and 58.6% for CIRM-BIA 129 and CIRM-BIA 138, respectively, 11 days post-inoculation. The numbers of cells with a non-permeabilized membrane (stained in green) decreased from 50 h to 150 h onwards, depending on the strains, to reach a minimum proportion of 50% at day 11 (Figure 3), suggesting that the number of dying or dead (but non-lysed) cells increased during the stationary and LTS phases.



**Figure 3:** (**A**) Proportion of non-permeabilized cells during the growth and survival of *P. freudenreichii* strains CIRM-BIA129 (**a**) and 138 (**b**) measured by LIVE/DEAD assay followed by epifluroscence microscopy (mean values of three epifluorescence microscopy fields at x40 magnification, and three fields at x100 magnification). Representative epifluorescence microscopy views of *P. freudenreichii* (CIRM-BIA 138)

cultures after LIVE/DEAD staining during exponential growth (c) and after 11 days of growth (d).

## Assessment of lysis by quantifying bacterial cells using qPCR

Because both plate counting assays and epifluorescence microscopy showed a reduction in culturability and an increase in membrane permeabilization, quantitative PCR was used to assess bacterial lysis. CIRM-BIA 138 (the strain with the highest population at day 11) cells were pelleted at three different time points before and after the observed loss of culturability. Quantitative PCR targeting the *tuf* gene was carried out on the DNA extracted from the pelleted cells. Under these conditions, lysed cells cannot normally be recovered (Falentin *et al.*, 2012). The copy number of the *tuf* gene did not vary significantly in the CIRM-BIA138 culture, demonstrating that the bacterial cells did not undergo lysis when entering the LTS phase (Figure 4).



**Figure 4:** Quantification of bacterial cells during the growth of *P. freudenreichii* CIRM-BIA138. Each point gives the copy number of *tuf* gene determined by qPCR on DNA extracted from *P. freudenreichii* CIRM-BIA138 (three biological replicates).

#### Discussion

#### P. freudenreichii has a high survival rate at day 11

P. freudenreichii belongs to the Actinobacteria, a group of Gram positive bacteria with high GC content genomes. This group is known for its considerable versatility (Gao & Gupta, 2012), and includes members that can survive under extreme conditions (Greenblatt et al., 2004). P. freudenreichii is reported to be a hardy species widely used in the cheese-making industry and is also well-documented for its probiotic potential (Jan et al., 2000). Genome sequence analysis has revealed the genetic basis for its hardiness: (i) its ability to store polyphosphates as an energy reserve and to utilize them instead of ATP, (ii) its ability to store glycogen as a carbon source, and (iii) several copies of chaperones to cope with different stresses (Falentin et al. 2010a). The eight P. freudenreichii strains studied here survived well during a period of 11 days under nutritional shortage, according to the optical density and plate counting assays performed (around 8 log<sub>10</sub> CFU/mL). In YEL medium, lactate - the preferential carbon source for PF- was exhausted two days after inoculation (Dalmasso et al. 2012). LIVE/DEAD staining followed by epifluorescence microscopy ascertained that more than 50% of the *P. freudenreichii* populations had an intact (non-permeabilized) membrane at this date. The growth curves for CIRM-BIA 129 and 138 (Fig. 1) displayed some fluctuations during long-term incubation. We suggest that intermittent cryptic regrowth might be responsible for this variability. These cryptic growths may result from some specific metabolisms induced to adapt to starvation or the growth of a sub-population of mutants derived from the inoculated strain and with a growth advantage that was explained in the review by Finkel et al. (2006).

#### The best surviving strain did not use partial lysis to sustain viability

Previous studies had shown that some lactic acid bacteria present in dairy products can lyse under difficult conditions and that the cell debris released into the culture medium may serve as a nutrient source to ensure survival of the non-lysed sub-population (Lortal *et al.*, 1997; Deutsch *et al.*, 2002; Lortal & Chapot-Chartier, 2005). A study of the autolysis of *Lactobacillus* and dairy propionic bacteria showed that this autolysis is associated with several factors, including among others the type of strain, autolysins, pH, temperature and ionic strength (Lortal *et al.*, 1997). For CIRM-BIA 138, the quantification of intact cells using quantitative PCR showed that under the study conditions applied here, lysis could be ruled out. Whether this is a general feature of *P. freudenreichii* still needs to be determined. The absence of lysis (i) could be confirmed by quantitative PCR on cells exposed to propidium monoazide, so as to prevent any contamination by DNA from dead cells, (ii) needs to be confirmed in other *P. freudenreichii* strains and under different conditions before it is generally accepted.

#### P. freudenreichii probably enters a viable but nonculturable (VBNC) state

The experimental data reported here offer a range of presumptions suggesting that *P. freudenreichii* enters into a VBNC state. VBNC is a survival strategy adopted by numerous bacteria in response to adverse conditions such as starvation, extreme temperature, pH, oxygen stress, antibiotic pressure, etc. Once in a VBNC state, the bacteria no longer grow in routine laboratory medium and conditions, but they nevertheless remain viable and can be resuscitated when more favourable conditions are provided (Ramamurthy *et al.*, 2014). The methods generally used to detect VBNC bacteria include bright field microscopy with nalidixic acid; fluorescent microscopy,

and especially use of the BacLight LIVE/DEAD kit; and molecular techniques (DNA or RNA) (Oliver, 2009; Fakruddin *et al.*, 2013). All of the eight *P. freudenreichii* strains studied here displayed a loss of culturability of ~1  $\log_{10}$  CFU/mL (Table S1 and Figure 5), when this parameter was compared between 50-72 hours and 11 days post-inoculation. LIVE/DEAD staining showed that about half of the bacterial cells had a permeabilized membrane. Using quantitative PCR, an absence of lysis was shown on strain CIRM-BIA138. These findings suggest that at least a part of the *P. freudenreichii* population entered a VBNC state and that the entry into LTS is concomitant with entry into a VBNC state, at least for 1  $\log_{10}$  (i.e. 90%) CFU/mL. Since the total bacterial population was stained either green (50%) or red (50%), we suggest that VBNC cells (90%) can take up both colours and that LIVE/DEAD staining cannot be used to quantify VBNC cells.



**Figure 5**: Fate of CIRM-BIA138 bacterial cells between 50-72 h post-inoculation and the long-term survival phase 11 days post-inoculation assessed by (a) culturability on

Petri plates, (b) LIVE/DEAD staining followed by epifluorescence and (c) quantification of intact cells by quantitative PCR.

The VBNC state has been described in various Gram positive bacteria, such as *Enterococcus faecalis, Enterococcus hirae, Listeria monocytogenes,* and *Mycobacterium tuberculosis* (Oliver et al., 2009), and in Gram negative bacteria such as *Vibrio cholerae* (Gonzales-Escalona et al. 2006), *Escherichia coli* (Darcan *et al.*, 2009) and *Helicobacter pylori* (Casasola-Rodríguez *et al.*, 2013). Many of the studies on the VBNC state have focused on pathogenic bacteria (Ramamurthy et al. 2014). This state has also recently been reported for bacteria of food interest and particularly in ripened cheese, suggesting a role for these bacteria in the organoleptic qualities of cheeses (Martin-Platero *et al.* 2008, Desfossés-Foucault *et al.* 2012, Ruggirello *et al.* 2014).

In conclusion, our findings suggest that one strategy for bacterial survival is dormancy, rather than partial bacterial lysis, in order to sustain the remaining cells. During the past decade, many studies have investigated the bacterial mechanisms underlying LTS in various fields: environmental, industrial, medical and biotechnological microbiology. This study represents the first work involving the phenotypic characterization of *P. freudenreichii* over an 11-day period without the addition of nutrients, and revealed different phases of growth, membrane permeabilization and entry into dormancy and a VBNC state to ensure LTS. A VBNC state may however have an important impact on the technological uses of these bacteria, as either starters in cheese-making or probiotics, since viability and metabolic activity are important traits in their wide range of potential uses. This therefore opens the way for further investigations on a molecular basis of the LTS of *P. freudenreichii*. The availability of genome sequences (Loux *et al.*, 2015) will greatly assist such investigations using comparative genomics and transcriptomics approaches in order to

understand the organization, genes and metabolic pathways required for LTS. Assessing viability and understanding the mechanisms underlying the survival of *P. freudenreichii* should be of considerable interest regarding the development of effective bacterial probiotics.

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### **Conflict of Interest section**

The authors have no conflict of interests to declare.

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## **Supporting Information**

Additional supporting information can be found in the online version of this article.

**Figure S1.** Kinetics of growth and survival measured by optical density in eight strains of *Propionibacterium freudenreichii*.



**Table S1**. Kinetics of the growth and survival of eight *P. freudenreichii* strainsmeasured by culturability in Petri dishes.

	Exp 1	Exp 1	Exp 1	Exp 1
Time (h)	Cirm 1	Cirm 9	Cirm 121	Cirm 138
0,00	1,33E+07	1,86E+07	1,28E+07	9,65E+06
7,00	1,39E+07	1,45E+07	1,19E+07	1,11E+07
24,30	1,49E+08	1,58E+08	1,42E+08	5,60E+07
31,30	1,47E+08	1,42E+08	3,86E+08	3,18E+08
48,30	3,51E+08	1,33E+09	5,76E+08	1,20E+09
55,30	1,43E+09	2,83E+09	1,95E+09	1,55E+09
72,30	1,08E+09	1,18E+09	3,55E+09	2,60E+09
97,00	5,51E+08	1,35E+09	8,26E+08	1,20E+09
168,00	2,45E+08	7,22E+08	2,95E+08	2,59E+08
192,30	1,59E+08	6,39E+08	1,35E+08	4,73E+08
216,30	1,56E+08	4,08E+08	2,62E+08	1,47E+09
240,30	1,34E+08	1,41E+08	1,01E+08	1,40E+08
264,30	1,01E+08	1,14E+08	2,00E+08	1,89E+08
	8,0	8,1	8,3	8,3

Table S1

-average day 11	1,14E+008	8,058105595
-standard error day 11	4,64E+007	7,666202446

# <u>Exp 1</u>

max day 11	2,00E+008	Cirm 121	
min day 11	1,01E+008	Cimr1	

	Exp 2	Exp 2	Exp 2	Exp 2
Time (h)	Cimr 119	Cirm 127	Cirm 129	Cirm 139
0,00	1,50E+07	2,55E+07	2,05E+07	6,45E+07
24,00	2,80E+08	6,10E+08	1,65E+08	4,43E+08
47,30	7,68E+08	1,09E+09	7,03E+08	1,51E+09
54,30	1,07E+09	1,49E+09	7,00E+08	2,13E+09
71,30	8,11E+08	3,27E+09	1,07E+09	4,35E+09
95,00	8,50E+08	4,25E+09	4,75E+08	2,03E+09
167,30	1,24E+08	6,40E+08	5,60E+08	6,03E+08
191,30	1,08E+08	4,58E+08	1,13E+08	5,78E+08
215,30	1,65E+08	3,00E+08	1,03E+08	2,93E+08
239,00	1,05E+08	1,57E+08	1,04E+08	1,45E+08
263,00	1,15E+08	1,43E+08	8,28E+07	8,53E+07
	8,1	8,2	7,9	7,9

#### <u>Exp 2</u>

max day 11	1,43E+008	Cirm 127
min day 11	8,28E+007	Cirm 129

	Exp 3	Exp 3	Exp 3	Exp 3
Time (h)	Cimr 9	Cirm 121	Cirm 127	Cirm 138
0,00	1,23E+07	2,35E+07	1,63E+07	1,14E+07
24,00	3,88E+08	4,73E+08	3,33E+08	4,83E+08
31,30	8,38E+08	1,87E+09	2,43E+08	3,53E+08
47,00	1,31E+09	2,78E+09	1,09E+09	1,30E+09
53,00	1,27E+09	2,03E+09	7,03E+08	7,05E+08
71,00	1,36E+09	1,03E+09	1,25E+09	2,98E+09
95,00	1,46E+09	1,32E+09	1,75E+09	1,52E+09
168,00	5,75E+08	2,45E+08	1,70E+08	2,33E+08
192,00	4,60E+08	3,35E+08	1,65E+08	1,43E+08
216,00	4,58E+08	4,13E+08	1,50E+08	1,77E+08
240,00	3,95E+08	3,28E+08	1,20E+08	1,16E+08
263,30	1,39E+08	9,23E+07	1,20E+08	1,29E+08
	8,1	8,0	8,1	8,1

<u>Exp 3</u>

max day 11	1,39E+008	Cimr 9
min day 11	9,23E+007	Cirm 121

| Exp 4    |
|----------|----------|----------|----------|----------|
| Time (h) | Cimr 1   | Cirm 119 | Cirm 129 | Cirm 139 |
| 0,00     | 1,24E+07 | 1,48E+07 | 1,80E+07 | 1,27E+07 |
|          |          |          |          |          |
| 23,00    | 1,31E+08 | 1,58E+08 | 3,13E+07 | 1,42E+08 |
| 29,00    | 7,00E+08 | 3,08E+08 | 1,98E+08 | 4,50E+08 |
| 47,00    | 1,35E+09 | 8,05E+08 | 5,95E+08 | 1,47E+09 |
| 53,00    | 1,48E+09 | 1,05E+09 | 6,68E+08 | 1,26E+09 |
| 71,00    | 1,03E+09 | 9,55E+08 | 9,13E+08 | 2,80E+09 |
| 94,30    | 4,55E+08 | 8,70E+08 | 9,15E+08 | 9,75E+08 |
| 166,00   | 2,58E+08 | 7,28E+08 | 1,73E+08 | 5,85E+08 |
| 189,30   | 1,13E+08 | 1,50E+08 | 7,25E+07 | 4,18E+08 |
| 212,00   | 6,43E+07 | 1,19E+08 | 4,50E+07 | 4,38E+08 |
| 235,30   | 7,33E+07 | 1,00E+08 | 8,83E+07 | 3,80E+08 |
| 259,00   | 6,15E+07 | 1,45E+08 | 7,78E+06 | 1,05E+08 |
|          | 7,8      | 8,2      | 6,9      | 8,0      |

# <u>Exp 4</u>

max day 11	1,45E+008	Cirm 119
min day 11	7,78E+006	Cirm 129

**Table S2.** Proportion of non-permeabilized cells during the growth and survival of eight

 *P. freudenreichii* strains measured by LIVE/DEAD staining followed by epifluroscence

 microscopy.

	Exp 1	Exp 1	Exp 1	Exp 1
Time (h)	Cimr1	Cirm 9	Cirm 121	Cirm 138
0,00	100,0%	100,0%	100,0%	100,0%
7,00	100,0%	100,0%	100,0%	100,0%
24,30	98,00%	90,00%	98,00%	98,00%
31,30	100,00%	100,00%	100,00%	100,00%
48,30	100,00%	100,00%	100,00%	100,00%
55,30	100,00%	100,00%	100,00%	100,00%
72,30	100,00%	99,00%	98,00%	100,00%
97,00	99,00%	100,00%	100,00%	99,00%
168,00	85,00%	95,00%	90,00%	95,00%
192,30	40,00%	64,00%	57,50%	54,00%
216,30	54,40%	74,25%	46,00%	67,33%
240,30	37,50%	64,50%	45,66%	63,50%
264,30	51,33%	56,83%	32,40%	58,55%

	Exp 2	Exp 2	Exp 2	Exp 2
Time (h)	Cimr 119	Cirm 127	Cirm 129	Cirm 139
0,00	90,6%	96,3%	98,0%	94,0%
24,00	96,5%	98,6%	95,4%	100,0%
47,30	96,90%	98,80%	100,00%	100,00%
54,30	99,83%	100,00%	99,50%	100,00%
71,30	97,00%	100,00%	97,75%	99,83%
95,00	95,00%	99,00%	67,85%	100,00%
167,30	71,05%	95,33%	75,50%	90,22%
191,30	69,70%	87,71%	78,62%	82,33%
215,30	68,00%	79,16%	68,88%	85,33%
239,00	55,03%	55,42%	77,00%	84,16%
263,00	59,37%	49,50%	77,86%	68,25%

	Exp 3	Exp 3	Exp 3	Exp 3
Time (h)	Cirm 9	Cirm 121	Cirm 127	Cirm 138
0,00	98,0%	100,0%	100,0%	100,0%
24,00	100,0%	92,0%	100,0%	89,5%
30,00	98,76%	99,23%	88,07%	99,08%
47,00	97,27%	100,00%	100,00%	99,61%
53,00	100,00%	99,25%	98,58%	99,90%
71,00	100,00%	100,00%	100,00%	99,00%
95,00	93,27%	78,50%	95,80%	87,16%
168,00	95,20%	72,60%	76,33%	88,16%
192,00	85,33%	87,00%	83,66%	78,83%
216,00	84,57%	81,58%	77,58%	78,60%
240,00	71,50%	84,33%	51,85%	49,00%
263,30	61,60%	77,00%	57,83%	61,50%

	Exp 4	Exp 4	Exp 4	Exp 4
Time (h)	Cirm 1	Cirm 119	<b>Cirm 129</b>	Cirm 139
0,00	97,7%	97,0%	98,5%	96,0%
23,00	100,0%	98,5%	100,0%	100,0%
29,00	100,00%	98,80%	95,00%	99,00%
47,00	100,00%	99,00%	94,71%	98,50%
53,00	100,00%	99,66%	98,83%	100,00%
71,00	98,42%	98,00%	100,00%	100,00%
94,30	98,33%	87,50%	95,60%	97,83%
166,00	97,83%	61,50%	69,58%	66,33%
189,30	69,00%	59,50%	63,00%	90,66%
212,00	67,00%	71,75%	74,70%	90,66%
235,30	39,88%	68,33%	78,87%	91,08%
259,00	62,00%	68,16%	65,85%	77,16%

SEÇÃO II: Utilização da genômica funcional para a compreensão do fenômeno de sobrevivência à longo prazo em *Propionibacterium freudenreichii* 

SECTION II : L'utilisation de lagénomiquefonctionnellepourcomprendre le phénomène de la survie àlongtermechezPropionibacteriumfreudenreichii

#### 1. Introdução

Esta segunda parte dos resultados é também apresentada sob forma de artigo científico. Este artigo segue os resultados obtidos no estudo da caracterização fenotípica apresentada na seção I de resultados deste manuscrito de tese.

O objetivo deste estudo foi conhecer o metabolismo e o transcriptoma de *P. freudenreichii* antes e depois da entrada em fase estacionária para compreender sua adaptação fisiológica à sobrevivência a longo prazo. Durante a cinética de crescimento e a sobrevivência durante 11 dias (Secção I - Resultados) da linhagem CIRM-BIA138, o sobrenadante da cultura foi amostrado para dosar as quantidades residuais de lactato, lactose, glicose, frutose, manose do sobrenadante de cultura bem como de produtos da fermentação: piruvato, propionato, acetato e succinato. Em paralelo, o RNAm destas células foram extraídos e sequenciados através da técnica de RNA-Seq à 1, 3 e 9 dias após a inoculação.

A análise dos dados de sequenciamento por mapeamento foi possível apenas nos pontos de 1 dia e 3 dias após a inoculação, correspondentes à fase exponencial e a entrada na fase estacionária, respectivamente. Isto ocorreu porque por motivos de qualidade e quantidade de ARN, não foi possível obter resultados das amostras de 9 dias. A análise dos genes diferencialmente expressos por EdgeR mostrou que 912 genes foram expressos diferencialmente entre a fase exponencial e o início da fase estacionária do crescimento bacteriano, entre eles, 458 genes são induzidos e reprimidos 454.

Para a obtenção do perfil metabolômico desta linhagem, os dados de expressão gênica global de *P. freudenreichii* CIRM-BIA138 foram combinados aos dados obtidos pela dosagem de metabólitos durante todo este período de 11 dias.

Os resultados bioquímicos demonstram que *P. freuden*reichii CIRM-BIA138 utiliza toda a glicose, manose e frutose no início da fase exponencial de crescimento até a exaustão destes compostos. Esta consumação é seguida de uma libertação e recaptação de piruvato, metabolito intermediário destes açúcares. Durante esta mesma fase de crescimento, observa-se um consumo de lactato até a exaustão à entrada na fase estacionária e uma produção de propionato e de acetato em quantidade equimolar. Os dados de RNA-Seq demostraram uma redução da expressão nos processos de transcrição, tradução e secreção de proteínas à 3 dias após a inoculação (entrada em fase estacionária) em comparação com a fase exponencial. Isto pode acontecer devido a uma carência de lactato no início da fase estacionária. Além disso, a análise de vias metabólicas diferencialmente expressas revelou que a glicólise, a fosforilação oxidativa, o ciclo de Wood-Werkman, a síntese de peptidoglicano e de ácidos graxos foram reprimidos à 3 dias de cultura. Em contraste, à 3 dias após a inoculação, o gene de síntese da cápsula de polissacárideo é induzida, assim como as vias de degradação de inositol, arabinose, asparagina, glicina, alanina, polifosfatos e, provavelmente, a via de degradação de putrescina ao GABA através do glutamato.

Assim, integrando neste estudo os dados transcriptômicos e metabolômicos, foi possível encontrar pistas sobre os mecanismos envolvidos na adaptação à carência nutricional que permitir a sobrevivência a longo prazo em *P. freudenreichii* CIRM-BIA138. As vias reprimidas indicam, provavelmente, uma redução da atividade metabólica desta bactéria para a sua entrada em dormência. Estes resultados abrem caminhos para outros estudos mais direcionados para compreender melhor os mecanismos envolvidos na estratégia de adaptação para a sobrevivência a longo prazo em *P. freudenreichii*.

#### 1. Introduction

Cette deuxième partie des résultats est aussi présentée sous forme d'article scientifique. Cet article fait suite aux résultats obtenus dans l'étude de la caractérisation phénotypique présentée dans la section I de résultats de ce manuscrit de thèse.

L'objectif de cette étude était d'appréhender le métabolisme et le transcriptome de *P. freudenreichii* avant et après à entrée en phase stationnaire pour comprendre son adaptation physiologique à la survie à long terme. Au cours de la cinétique de croissance et de survie pendant 11 jours (section I - Résultats) de la souche CIRM-BIA138, le surnageant de culture a été échantillonné pour doser les quantités résiduelles de lactate, lactose, glucose, fructose, mannose du milieu de culture ainsi que les produits de la fermentation : pyruvate, propionate, acétate et succinate. En parallèle, l'ARNm des cellules a été extrait et séquencé (RNA-Seq) à 1, 3 et 9 jours après inoculation.

L'analyse des données de séquencage par mapping n'a été possible, pour des raisons de qualité et de quantité d'ARN que sur les points 1j et 3j après inoculation correspondant à la phase exponentielle et à l'entrée en phase stationnaire respectivement. L'analyse des gènes différentiellement exprimés par EdgeR a montré que 912 gènes étaient différentiellement exprimés entre la phase exponentielle et le début de la phase stationnaire de la croissance bactérienne, parmi eux, 458 gènes sont induits et 454 réprimés.

Pour obtenir le profil métabolomique de la souche, les données de l'expression génique globale de *P. freudenreichii* CIRM-BIA138 ont été combinées aux données obtenues par dosage de métabolites au cours de toute cette période de 11 jours.

Les résultats biochimiques montrent que *P. freudenreichii* CIRM-BIA138 utilise tout le glucose, le mannose et le fructose au tout début de la phase exponentielle de croissance jusqu'à épuisement de ces composés. Cette consommation est suivie d'un relargage puis d'une reconsommation de pyruvate, métabolite intermédiaire de ces sucres. Pendant cette même phase de croissance, on observe une consommation du lactate jusqu'à épuisement à l'entrée en phase stationnaire et une production de propionate et d'acétate en quantité équimolaire. Les données biochimiques de la phase exponentielle sont en accord avec celles décrites dans la bibliographie. Pendant la phase stationnaire et la survie à long-terme le pyruvate présent dans le milieu est progressivement consommé jusqu'à épuisement à 11 jours après inoculation. Les données RNA-Seq ont montré une réduction d'expression dans les processus de transcription, traduction et sécrétion de protéines à 3 jours après inoculation (entrée en phase stationnaire) en comparaison avec la phase exponentielle. Cela peut se produire en raison de la carence en lactate au début de la phase stationnaire. De plus, l'analyse des voies métaboliques différentiellement exprimées a montré que la glycolyse, la phosphorylation oxydative, le cycle de Wood-Werkman, la synthèse de peptidoglycane et d'acides gras étaient réprimés à 3 jours de culture. En revanche, à 3 jours après inoculation, le gène de synthèse de capsule polysaccharidique est induit ainsi que les voies de dégradation de l'inositol, de l'arabinose, de l'asparagine, de la glycine, de l'alanine, des polyphosphates et probablement la voie de dégradation de la putrescine en GABA via le glutamate.

Ainsi, en intégrant dans cette étude les données transcriptomiques et métabolomiques, il a été possible de trouver des pistes concernant les mécanismes impliqués dans l'adaptation à la carence nutritionnelle qui permettraient la survie à long terme chez *P. freudenreichii* CIRM-BIA138. Les voies réprimées indiquent probablement une réduction de l'activité métabolique de cette bactérie en vue de son entrée en dormance. Ces résultats ouvrent la voie à d'autres études plus ciblées pour mieux cerner les mécanismes impliqués dans cette stratégie d'adaptation à la survie à long terme chez *P. freudenreichii*.

# 2. Artigo 2: Adaptation of *Propionibacterium freudenreichii* to long-term survival in case of gradual nutritional shortage

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#### Abstract

#### Background

*Propionibacterium freudenreichii* is an Actinobacterium widely used in the dairy industry as a ripening culture for Swiss-type cheeses, for vitamin B12 production and some strains display probiotic properties. It is reportedly a hardy bacterium, able to survive the cheese-making process and digestive stresses.

#### Results

During this study, *P. freudenreichii* CIRM-BIA 138, which has a generation time of 5H in Yeast Extract Lactate medium at 30°C under microaerophylic conditions, was incubated for 11 days (9 days after entry into stationary phase) in a culture medium, without any adjunct during the incubation. The carbon and free amino acids sources available in the medium, and the organic acids produced by the strain, were monitored throughout growth and survival. Although lactate (the prefered carbon source for *P. freudenreichii*) was exhausted three days after inoculation, the strain sustained a high population level of 9.3 log<sub>10</sub> CFU/mL. Its physiological adaptation was investigated by RNA-seq analysis and revealed a complete disruption of metabolism at the entry into stationary phase as compared to exponential phase.

#### Conclusions

*P. freudenreichii* adapts its metabolism during entry into stationary phase by downregulating oxidative phosphorylation, glycolysis, and the Wood-Werkman cycle by exploiting new nitrogen (glutamate, glycine, alanine) sources, by down-regulating the transcription, translation and secretion of protein. Utilization of polyphosphates and rearrangement at chromosome level were suggested.

**Keywords:** *Propionibacterium freudenreichii*, long-term survival, stationary phase, RNA-seq, adaptation.

#### Background

*Propionibacterium freudenreichii* is an Actinobacterium widely used in the dairy industry and responsible for aroma development and opening in Swiss-type cheeses. Some strains are also used as probiotics because they produce bifidogenic compounds [1], they are resistant to digestive stress [2] and they may be endowed with anti-inflammatory capabilities and could be used to prevent inflammatory bowel diseases [3] [4]. *P. freudenreichii* core metabolism leads to propionic acid as the main end-product [5] [6].

The genetic bases for hardiness have already been well-documented. *P. freudenreichii* can produce ATP and NAD(P)H via fermentation utilizing an unique metabolic pathway (the Wood-Werkman cycle), leading to propionic acid. Under anaerobiosis, *P. freudenreichii* can also use electron acceptors other than  $O_2$  (such as humic acid or nitrate) to produce ATP during anaerobic respiration [7] [8]. Several genes involved in polyphosphate and pyrophosphate utilization are found in the genome of CIRM-BIA1, the type strain for *P. freudenreichii*, suggesting a storage of energy in these forms [9]. However, these genes have never yet been found to be induced under any growth conditions.

*P. freudenreichii* can stay alive and metabolically active for long periods even under stressful conditions like gastro-intestinal tract environment or in cold condition. When placed at stationary phase in the intestinal tract (colon) of a piglet, CIRM-BIA1 was able to ferment some atypical carbon sources present in the gut (gluconate and propanediol), enabling its survival under these harsh conditions. Both of these catabolic pathways were reconstructed in silico from the genomic data and were found to be fully expressed in the colon of piglets [10]. Another transcriptomic study on the adaptation of

*P. freudenreichii* strains to the cold in the absence of carbon shortage showed that strains slowed down their cellular metabolism, displayed cold stress responses, and rerouted their carbon metabolism toward trehalose and glycogen synthesis [11]. In addition, proteomic studies on different *P. freudenreichii* strains have highlighted the role of chaperones during exposure to acids, bile salts or NaCl [12] [13] [14]. The genomic analysis of *P. freudenreichii* strains thus revealed the duplication of several chaperone genes.

In a previous study [15], we carried out the phenotypic characterization of *P. freudenreichii* over an 11-day period without the addition of nutrients, and revealed different phases of growth, membrane permeabilization and entry into dormancy and a viable but non-cultivable (VBNC) state to ensure Long-Term Survival (LTS). How *P. freudenreichii* prepares the LTS phase remains unknown and no data are currently available on the metabolism of *P. freudenreichii* in stationary phase in the event of a gradual nutritional shortage.

The objective of the present study was therefore to identify the pathways used by *P*. *freudenreichii* to cope with starvation and enter to the LTS phase. The strategy adopted here combined the quantification of sugars, acids and free amino acids in the supernatant of an 11-day culture without the addition of nutrients, and an RNA-seq analysis of bacterial cells sampled at three different time points during *P*. *freudenreichii* culture. This work was designed to compare both biochemical quantifications (acids, amino-acids and sugars) and transcriptomic data between the exponential and stationary phases in order to gain an in-depth view of *P*. *freudenreichii* adaptation to LTS.
## **Results and Discussion**

The objective of the study was to identify the metabolic pathways used by *P*. *freudenreichii* to cope with starvation and 'prepare' entry into the LTS phase.

#### Enumeration, DO, pH

The results of bacterial enumerations and pH measurements (Figure 1A) confirmed that CIRM-BIA 138 survived well (around 8  $log_{10}$  CFU/mL) during 11 days, even under conditions of nutritional shortage, as it had been observed during a previous study [15]. When inoculated at 7  $log_{10}$  CFU/mL, the *P. freudenreichii* culture entered the exponential growth phase to reach a maximum population of 9.3  $log_{10}$  CFU/mL 3d post-inoculation, corresponding to entry into the stationary phase. The pH was very slightly affected by growth since the most acidic point was 6.7. At entry into stationary phase, a slight decline in the population was observed until it stabilized at around 8.5  $log_{10}$  CFU/mL as from 9d post-inoculation.

### Carbon source utilization, organic acid utilization and production

CIRM-BIA138 can degrade glucose, glycerol, mannose, galactose, inositol, erythritol, adonitol, esculine, lactose, xylitol, gluconate, D-fructose and L-arabinose [5]. In this study, CIRM-BIA138 was cultivated in Yeast Extract Lactate, a growth medium containing 120 mM lactate. Small quantities of several other carbon sources were also identified: 0.42 mM lactose, 0.15 mM glucose, 0.07 mM fructose, 0.03 mM mannose. Inositol and arabinose were not detected and probably below the detection limit of 0.5 mM. *P. freudenreichii* CIRM-BIA138 used lactose, glucose, mannose and fructose at the beginning of growth until they were completely exhausted at 24h post-inoculation (Figure 1B). These degradation capacities are in line with those previously described for

this strain [5]. The degradation of these sugars was concomitant with the release of pyruvate into the medium, this being one of their intermediary metabolites. From 24h post inoculation onwards, pyruvate was consumed by the strain. Levels of other compounds such as propionate, acetate and succinate rose gradually in the medium over time during growth, to reach a plateau at 3d post-inoculation (Figure1C). The stoichiometry of the conversion was 3 mol of lactate into 1 mol of acetate and 2 mol of propionate, and is in line with those described previously [16]. Lactate levels decreased as early as the onset of growth because of its consumption by *P. freudenreichii*. Lactate was exhausted 3d post-inoculation and pyruvate 11d post-inoculation. Among all the carbon sources quantified during this experiment, pyruvate was the only carbon source detectable in the medium when *P. freudenreichii* entered the stationary phase.

Overview of differentially expressed genes between exponential and stationary phases

After filtration to eliminate reads of poor quality, sequencing generated 100,450,770 reads (FastQC results), which included 13,502,207 non-aligned reads and 37,018,311 reads aligned on coding sequences. The remaining reads corresponded to those mapping on two different sequences (1,147,269 ambiguous reads according an htseq-count with a union parameter), on tRNA, rRNA, or non coding RNA (small RNA, non-sense RNA, 5' and 3 ' UTR). Only reads mapping on coding sequences were analysed further. Overall, 912 genes were found to be differentially expressed (adjusted pvalue<0.05) between 1d (exponential phase) and 3d post-inoculation (start of stationary phase) (458 genes induced (Additional file 1: Table S1), 454 genes repressed (Additional file 2: Table S2)). Differentially expressed genes (adjusted pvalues<0.05, pvalues available in Table S1, S2 and S3) represented 40% of the protein-coding genes in the CIRM-BIA138 genome. The

genes most affected by entry into stationary phase was those implicated in carbohydrate metabolism with 26 genes of this functional category being induced and 41 repressed. Translation was the second most affected category, with two genes induced and 56 repressed. Transport/binding was the third most affected category, with 28 genes induced and 19 repressed. Transcription was the fourth most affected category, with 29 genes induced and 17 repressed. Membrane bioenergetics was the fifth most affected category, with nine genes induced and 36 repressed. Amino acid metabolism category was also affected, with 11 genes induced and 32 repressed. Lastly, the metabolism of coenzyme and prosthetic groups was also affected, with 11 genes induced and 29 repressed (Figure 2). Most induction and repression observed from RNAseq analysis were confirmed by RTqPCR analysis (pavalue<0.05, Table S3). To validate RNAseq results, we performed RT-qPCR and we have included the 9d post inoculation sample. In most cases, the tendency (induction versus repression) seen at 3d post inoculation was confirmed at 9d. A massive number of differentially expressed genes (63 repressed and 177 induced) were annotated as having an 'unknown function'. 39% of induced genes at stationary phase were of unknown function. The genome of CIRM-BIA 138 contains 27 % of genes encoding protein of unknown function. So, results suggested that genes expressed at the entry into stationary phase are enriched in genes without function, thus reflecting our lack of knowledge on bacterial stationary phase metabolism. All induction and repression discussed below are statistically significant at adjusted pvalue<0.05 for RNAseq and at pvalue<0.05 for RT-qPCR.

# Slowdown of the metabolism leading to dormancy

RNA extraction and quantification throughout growth and survival showed that RNA levels gradually declined. The total/depleted RNA quantities corresponding to 2 mL of

culture were  $60\mu g/290$  ng at 1d post-inoculation (exponential growth phase) for 15 log<sub>10</sub> CFU, 22 µg/225 ng at 3d post-inoculation (entry into stationary phase) 19 log<sub>10</sub> CFU and 15  $\mu$ g/90 ng at 11d post-inoculation 18 log<sub>10</sub> CFU. The translation machinery was also reduced at 3d post-inoculation. Indeed, expression of the translation initiation factor IF-3, and the elongation factors Tu and G encoded by genes infC, tuf, fusA, were 2, 1.8 and 3-fold repressed, respectively. The expression of ribosomal proteins was repressed from 1.5 to 16 fold (see Table S2, for repression fold and pvalue of each gene). Taken together, these repressions reflected a slowdown of the cellular machinery in line with the process of dormancy recently described [15]. This slowdown can be explained by the scarcity of carbon sources when *P. freudenreichii* enters the stationary phase at 3d post-inoculation. The main glycolytic pathways producing reducing equivalents were repressed during this phase. In YEL medium, glycolysis, the pentose phosphate pathway and the Wood-Werkman cycle (fermentation) are three ways for P. freudenreichii to produce NADH and NADPH reducing equivalents. ATP and precursor metabolites are required for the biosynthesis of essential compounds (amino acids, purine, pyrimidine, glycerol 3 phosphate, fatty acids, N-acetyl glucosamine, vitamins). Here, under the conditions we applied, glucose was exhausted at 3d post-inoculation, and the genes involved in glycolysis were repressed. Some of them were particularly strongly down-regulated, such as sglT, the glucose transporter with a fold change of 4, and pfkA, pfp, fba1 and eno2 with fold changes of 2.2, 3.1, 1.9 and 3.4, respectively. The repression of sglT was confirmed by RT-qPCR results (pvalue<0.05) with a repression fold of 6 at 3d and 88 at 9d post-inoculation (Table S3).

The Wood-Werkman cycle is specific to certain propionic acid-producing bacteria. It plays a central role in propionic acid fermentation, the principal carbon metabolic pathway in dairy propionibacteria. This pathway provides major sources of reducing equivalents: NADPH required for biosynthesis reactions and NADH necessary for oxidative phosphorylation. Through this pathway, pyruvate is converted into propionate. Pyruvate is initially converted into succinate by the successive steps of the tricarboxylic cycle (TCA). At 3d post-inoculation, gene-set enrichment analysis identified the "Citrate Cycle" as one of the most down-regulated pathways (ko00020, fold change of 2.1, pval=2.7e-04). All the corresponding genes (mdh, fumC and sdh) were repressed, with fold changes of 1.9, 2.1 and 8.6, respectively, in stationary phase compared to exponential phase (see Table S2). Succinate is then converted into succinyl-CoA, methyl malonyl CoA, propanoyl CoA and propionate by specific enzymes. Transcripts corresponding to the 12S, 5S and 1.3S subunits of the well-studied methylmalonyl-CoA carboxyltransferase were repressed with fold changes of 2.3, 1.8 and 2.9, respectively. RT-qPCR results confirmed the repression of mmda, encoding the 12S subunit with a fold change of 2Down-regulation of the Wood-Werkman cycle is probably involved in maintaining the redox balance. Although glycolysis and the Wood Werkman cycle were down-regulated, all genes in the pentose phosphate pathway (except the rpiB3 gene, induced with a fold change of 2.9) were stably expressed between 1d and 3 d post-inoculation.

#### Changes in oxidative phosphorylation

Several transcripts (nuoA,B,C,D,E,F,I,J,K,L,M,N genes) encoding the different chains of NADH-quinone oxidoreductase (responsible for the release of electrons and H+ contained in the NADH molecule) involved in aerobic respiration were down-regulated at 3d post-inoculation, with a fold change of between 3.8 and 6.3. The PFCIRM138\_05930 gene encoding an iron-sulphur protein was also down-regulated, with a fold change of 6 in RNAseq analysis and 4.5 at 3d and 5.5 at 9d post- inoculation according to RT-qPCR results. Likewise, sdhA, A3, B, B3, C1, and C2, encoding the different subunits of succinate dehydrogenase, were down-regulated with fold changes of between 4.4 and 8.8 (for details, see Supplementary Table S1 and S2). Repression of sdhC1 was confirmed by RT-qPCR results with a repression with a fold of 9 (Table S3). By contrast, both the dmsC and dmsB genes, encoding the anaerobic dimethyl sulphoxide reductase chains C and B, were induced with fold changes of 13.7 and 4.7, respectively. The latter protein handles the final transfer of electrons to various sulphoxide and N-oxide compounds. Anaerobiosis-inducible dimethyl sulphoxide reductases play a key role in bacterial adaptation to anaerobic conditions in bacteria and serve as terminal reductases using DMSO as a terminal electron acceptor [17]. Nitrate, sulphur or ferrous ions can act as electron acceptors. Accordingly, the gene encoding the permease protein of the nitrate ABC transporter ssuC was induced with a fold change of 4.4, the gene citT2 encoding a sodium:sulphate symporter was induced with a fold change of 4.3, and the feuC and feuS genes encoding the ferrous ABC transporter were induced with fold-changes of 2.5 and 2.8. Some subunits of nitrate reductase, namely those encoded by narH and narJ, were induced by more than 22-fold. RT-qPCR results confirmed the induction of narH with a lesser fold of 3.5. During growth, the headspace of the tube contained air and was therefore a source of O2. Although the culture was grown without agitation, the medium contained traces of  $O_2$ , which were probably used as a terminal electron acceptor during the exponential growth phase. At 3d postinoculation, the CO<sub>2</sub> released by the bacteria likely saturated the headspace of the tube and the P. freudenreichii were in anoxic conditions. This might modify the redox balance due to the lower availability of reducing equivalents, and thus explain the down-regulation of oxidative phosphorylation. At the pathway level, gene-set enrichment analysis (see Materials and Methods) showed that oxidative phosphorylation was indeed the most significantly down-regulated (ko00190, fold change of 4.4 and pval=1.2e-12) of all of the pathways down-regulated during the study. A similar down-regulation had been observed when *P. freudenreichii* was placed in the colon of piglets under anoxic conditions [10]. To assess whether the oxydative repression is due to traces of O2 in the headspace or due to the metabolism at the entry into stationary phase, cultures growing in bioreactors under strictly controlled conditions can be considered.

The changes thus observed suggest profound metabolic reprogramming in response to starvation. Similarly, during our study, specific catabolic pathways were induced (see below).

#### Cell wall

The cell envelope of Gram positive bacteria such as *P. freudenreichii* comprises the inner cell (cytoplasmic) membrane and the cell wall of the bacterium composed of peptidoglycan and various associated compounds (proteins, polysaccharides, teichoic acids) that differ from one species to another. In *P. freudenreichii*, some strains are known to possess a surface exopolysaccharidic (EPS) layer composed of (1 3,1 2)- D-glucan [18].

Peptidoglycan (PG) is essential to maintaining cell shape and also providing mechanical protection against osmotic pressure. It is also involved in cell division process. PG is a three-dimensional network made up of N-acetylglucosamine (GlcNAc) and N-acetylmuramic acid(MurNAc), The carboxyl groups of MurNAc are substituted by a short peptides, interconnecting the chains together. The biosynthesis of PG includes cytoplasmic steps mediated by MurA to F, and thereafter the transfer through the membrane via the transferases MarY and MurG, and finally the binding of new material to cell wall. At entry into stationary phase,

the PG biosynthesis pathway is significantly down-regulated. The genes encoding MurA and MurB that are involved in the formation of UDP-N-acetylmuramate were down-regulated, with a fold changes of 1.8 for both. The murC, D, E, and F genes encoding cytoplasmic enzymes responsible for the sequential adjunction of aminoacids to UDP-N-acetylmuramate, leading to the formation of UDP-MurNAcpentapeptide, were also down-regulated (with fold changes of 2.3, 1.6, 1.2, and 2, respectively) as well as the gene ddlA reponsible for the formation of the D-alanyl-D-alanine dipeptide is down regulated (fold changes of 2,9). Two proteins MraY and MurG ensure the transfer of the phospho--MurNAc-pentapeptide moity of the UDP-MurNAc-pentapeptide to the membrane acceptor and the addition of GlcNAc, leading to the formation of GlcNAc-MurNAc-pentapeptide on the lipid carrier. The mraY gene encoding UDP- MurNAc-pentapeptide phosphotransferase was repressed with a fold change of 3.4, whereas expression of the murG gene remained stable. Finally, the enzymes responsible of the late steps of the biosynthesis of the PG are also down regulated: fstI (fold change of 1,9),mrc/ponA (FC of 1,5) Taken together, our results showed a down-regulation of peptidoglycan synthesis, which agreed with the growth arrest and entry into dormancy observed for CIRM-BIA138.

The CIRM-BIA138 strain produces a surface -D-glucan polysaccharide [18]. A single gtfF gene is responsible for synthesis of this polysaccharide. The gtfF gene is strongly induced in stationary growth phase compared to the exponential phase, with a fold change of 6.1, which might lead to an overproduction of surface -D-glucan polysaccharide. The induction was confirmed with a fold change of 3.3 at 3d but not at 9d post-inoculation by RT-qPCR results. Such EPS production might protect the bacteria against the unfavourable conditions encountered in stationary phase and

therefore prepare them for long-term survival, as has previously been shown in other food species such as Oenococcus oeni [19].

The cell wall of *P. freudenreichii* is also coated with proteins that are anchored via an SLH domain. SlpE, SlpF and SlpG are three of these proteins. Interestingly, during this study, the slpE gene was found to be strongly up-regulated (fold change of 12.9); whereas slpG and slpF were down-regulated (fold change of 2.8 and 2.2 respectively). According to RT-qPCR results, the induction of slpE is transient since it was confirmed at 3d with a fold change of 5.5 but not at 9d post-inoculation. As of yet, the physiological role of these proteins has not been elucidated, but these results suggest differing roles in long-term survival for SlpE and the other two proteins.

The bacterial membrane mostly comprises a protein-embedded phospholipid bilayer. In CIRM-BIA138, the fatty acid biosynthesis pathway was strongly repressed in stationary phase compare to exponential phase. The fabF and fabH genes encoding the enzyme charging the acetyl residue from acetyl-CoA to acylcarrier protein (Acp) were repressed, with respective fold changes of 6.3 and 6.9. RT-qPCR results confirmed repression of fabF with a fold-change of 8.2 at 3d and 6.6 at 9d post-inoculation. The acp gene was repressed, with a fold change of 5.9. In the same way, the fabD genes encoding the enzyme charging the malonyl residue from malonoyl-CoA to Acp was repressed, with a 3.7-fold change. Genes such as fabG or inhA, which are responsible for the further elongation of fatty acids, were repressed by 1.9 and 1.7, respectively. Bacteria can produce fatty acids anaerobically. Since most fatty acids in bacterial cells are used for membrane phospholipid synthesis, growth arrest at the entry into stationary phase at 3d compared to 1d post-inoculation (see CFU count, Figure 1) might limit the need for fatty acids, and lead to the down-regulation we observed here.

### **Diversification of nutrients**

At 3d post-inoculation, glucose, lactose, and lactate were exhausted. *P. freudenreichii* therefore needed to recruit other catabolic pathways to produce NADH and NADPH reducing equivalents, ATP and precursor metabolites required for the survival. Inositol and arabinose pathways were induced. Gene-set enrichment analysis revealed that 'Pentose and glucuronate interconversions' were the most markedly induced KEGG pathways (ko00040; fold change of 1.7, pval 0.003). In this large pathway, the degradation of arabinose appeared to be the only one to be induced.

#### <u>Arabinose</u>

Most P. freudenreichii strains can degrade L-arabinose, and the degradation pathway has previously described L-arabinose cell been [5]. enters the via а xylose/ribose/arabinose/galactoside ABC transporter encoded by rbsBA that is not differentially expressed. L-arabinose is sequentially converted to L-ribulose, L- ribulose 5-phosphate, and D-xylulose 5-phosphate by the action of the L-arabinose isomerase encoded by araA, L-ribulokinase encoded by araB and ribulose-5- phosphate 4epimerase encoded by araD and araD1, which in our study were induced 2.2, 2.2, 1.1 and 2.9 fold, respectively. However, biochemical quantification failed to detect arabinose which is probably present at a concentration lower than the detection limit of 0.5 mM (like glucose, lactose, fructose, mannose) in the medium.

## Inositol

Inositol is a six-fold cyclohexane alcohol. Inositol is found in many foods (particularly in fruits) and is probably present in the yeast extract contained in YEL culture medium. Inositol is transported into the cell by a transporter encoded by iolT1, iolT2 and iolT3, which were found to be induced with respective fold changes of 11, 3.2 and 1.7, at the entry into stationary phase compared to exponential phase (Table S2). The induction of iolT1 was induced 2.4 at 3d and 58.3 at 9d according to RT-qPCR results. In the cell, inositol is transformed into 2-keto-inositol by inositol dehydrogenase that is encoded by the iol, iolG2 genes, induced with fold changes of 5.5 and 2.1, respectively. 2-ketoinositol is then transformed into 2,3-di-keto-4-deoxy inositol by 2-keto inositol dehydratase encoded by iolE3, which was found to be induced by a 2.2 fold change. 2,3-di-keto-4-deoxy inositol is then transformed into 2-deoxy-5-keto gluconic acid by the product of the iolB gene repressed with a fold change of 2.1. The iolC and iolD genes, whose products convert 2-deoxy-5-keto gluconic acid into malonic semialdehyde, were not differentially expressed. The induction of inositol degradation was not detected by GAGE, probably because the steps in this pathways are not fully described in the KEGG orthology map for P. freudenreichii species. The inositol degradation pathway had previously been annotated manually in all sequenced strains able to degrade inositol [20], and this revealed a high number of paralogous genes at different steps in the pathway (Figure 3), suggesting that the pathway is essential for this species. During our study, we were able for the first time to demonstrate the expression and induction of the inositol degradation pathway in P. freudenreichii. Unfortunately, inositol was undetected in the culture medium (data not shown).

## Use of asparagine

Biochemical analysis of the culture medium showed that asparagine was totally consumed between 1d and 3d post-inoculation (1.3 mM consumption, Figure 4). The KEGG and Metacyc pathways report that in bacteria, asparagine is degraded into aspartate that is further transformed into fumarate or succinate. Despite asparagine exhaustion in the CIRM-BIA138 supernatant, the transcripts of asmA and asmB enabling the transformation of asparagine into aspartate were both repressed at 3d postinoculation compared to 1d post-inoculation, with fold changes of 2.1 and 1.5, respectively. In P. freudenreichii, the fate of asparagine between 1d and 3d postinoculation remains unknown [21]. The use of asparagine as an energy supply has previously been suggested [18]. Asparagine is described as being co-metabolized along with aspartate and lactate in type strain P. freudenreichii CIRM-BIA1 [22]. However, the quantification of free aspartate did not support this idea in the P. freudenreichii CIRM-BIA138 strain. The concentration of free aspartate in the medium was not significantly different between 1d and 3d post-inoculation (t-test, P value >0.05). The majority (i.e. 70% of the 100 P. freudenreichii isolates previously tested) displayed very low levels of aspartate activity [22]. In CIRM-BIA138, the lack of aspartate consumption could be explained by the lack of the dcuA gene enabling the transport of aspartate into the cell.

#### Putrescine

Putrescine is a deleterious biogenic amine that is often present in mature cheeses. In general, Swiss-type cheeses containing the *P. freudenreichii* species do not exhibit high levels of putrescine [24]. Whether this is due to the degradation of putrescine by *P. freudenreichii* remains to be demonstrated. The degradation pathway for putrescine has

not yet been described in P. freudenreichii. During the present study, the PFCIRM138\_100015 gene, putatively encoding a putrescine importer, was induced with a fold change of 3.7 at entry into stationary phase. In E. coli, putrescine degradation is supported by the gadA and gadB gene products and involves steps that are shared with those of glutamate. The glutamate decarboxylase encoded by gadB was 5-fold more induced at 3d compared to 1d post-inoculation. RT-qPCR results confirmed a strong and progressive induction of gadB with a fold change of 3 at 3d and of 114 at 9d postinoculation. The glutamate concentration did not differ statistically at 3d and 1d postinoculation (t-test, P value >0.05), suggesting that degraded glutamate arose from the prior degradation of putrescine. This induction was also consistent with that observed in E. coli for the gadA and gadB genes during entry into stationary phase. It has been shown that glutamate decarboxylase activity increases the survival of Listeria monocytogenes, because intracellular glutamate is stored and converted into aminobutyric acid (GABA), and extracellular glutamate is coupled to an antiporter in the gad system [25]. The CIRM-BIA138 genome contains two genes encoding a GABA transferase: PFCIRM138\_10030 (gabT), which was repressed with a fold change of 6.6, and PFCIRM138\_04365, which was induced with a fold change of 10.6. However, neither GABA nor putrescine were detected in the medium despite a quantification limit (data not shown). The induction of 25 μM of putrescine importer (PFCIRM138\_100015) and gadB at d3 and d9 is not sufficient to know whether these both genes were involved in the metabolism of stationary phase and whether glutamate metabolism could be linked to an hypothetical putrescine degradation by P. freudenreichii.

### Use of glycine

The dosage of of free glycine in the medium revealed a 0.42 mM consumption between 1d and 3d post-inoculation (Figure 4). The *dad*A2 gene encoding glycine oxidase, which catalyses glycine oxidative deamination was induced at 3d compared to 1d postinoculation, with a fold change of 13 according to RNAseq analysis but the induction was not statistically significant according to RT-qPCR result. This gene is the only one revealing a discrepency between RNAseq data and RT-qPCR analysis. However we can conclude to a maintain of dadA2 expression at 3d and even at 9d post-inoculation. During osmotic stress induced by a high NaCl concentration in a medium containing glycine betaine (also named trimethylglycine), it was found that *Propionibacterium acidipropionici* accumulates glycine betaine in order to balance osmolarity and enable adaptation [26]. Other studies have also shown that in *P. freudenreichii*, glycine and proline exert protective effects against high osmolarity throughout bacterial growth [21]. Internalization of glycine could be a way to cope with osmotic conditions encountered at the entry of stationnary phase.

## Use of alanine

Alanine was the free amino acid that was most consumed (1.17 mM between 1d and 3d post-inoculation). This was in line with induction of the *ald* gene encoding an alanine dehydrogenase (2.7 fold change), which deaminates L-alanine into pyruvate. The induction of *ald* throughout growth has already been observed in stationary phase induced by low temperature in the absence of nutrient shortage [6]. We showed here that such *ald* induction also occurred in a context of nutrient shortage.

## The role of proline

Biochemical analysis between 1d post-inoculation and 3d post-inoculation an almost two-fold increase in proline was measured in the surpernatant (0.87 mM released) (Figure 4). As no gene for proline synthesis was induced at 3d post-inoculation, this suggests that proline was released due to the degradation of proteins and peptides in the medium. This was consistent with the action of a proline iminopeptidase (Pip) which cleaves amino-terminal proline residues from peptides and has previously been described in cheese [27].

## Chaperone and detox

Protein chaperones protect other proteins against aggregation and denaturation. In *P. freudenreichii*, chaperones are strongly expressed in the case of acidic or bile salt stresses [31] or under cold conditions [6]. At 3d post-inoculation, the three sequences hsp20 1, hsp20 2 and hsp20 3 encoding heat shock proteins were induced with fold changes of 6.7, 6.9 and 2.5, respectively. The copZ gene encoding a copper chaperone was induced with a fold change of 2.5. The cspB gene encoding a cold shock protein was induced with a 2.7 fold change. The number of chaperones induced seems low when compared to the large number of chaperone-encoding genes described in the literature for *P. freudenreichii*. The limited induction of chaperone genes might reflect the efficient adaptation of *P. freudenreichii* to nutritional shortage. Alternatively, and unlike osmotic or heat-shock stress, nutritional shortage may not induce protein misfolding or denaturation and thus not cause the expression of many chaperone genes.

#### **Competence**

In several species, natural competence enables the bacteria to cope with stresses or nutritional shortage. There are three prevailing hypotheses regarding the benefits of DNA uptake and recombination in bacteria (for a review, see [32] [33]): (i) DNA is used as a nutrient source; (ii) DNA is used to improve the efficiency of natural selection (by acquiring new genetic information); and (iii) DNA is used as a template to repair damaged chromosomal DNA. In the case of P. freudenreichii, a gene encoding DNA transfer protein (PFCIRM138\_00630) was induced with a fold change of 14.7, and dprA encoding a DNA processing /uptake protein was induced with a fold change of 27.3 at 3d post inoculation. Two contiguous genes encoding mobilization protein (PFCIRM138\_07015, PFCIRM138\_07020) were induced with fold changes of 49.4 and 10.8 respectively, one gene (PFCIRM138\_07570) encoding a conjugative relaxase was induced with a 24-fold change and lastly the comEC gene involved in competence was induced with a fold change of 17.4 at 3 d post-inoculation. PFCIRM138\_07015 induction was confirmed by RT-qPCR result with a fold change of 23.6 at 3d and 26.7 at 9d post-inoculation. P. freudenreichii is known to have evolved more by recombination than by mutations [34] and possesses a high copy number of insertion sequences and integrase genes in its genome [9]. Experiments with DNA shuffling performed to improve the production yields of vitamin B12 also showed a high capacity for genome rearrangement in P. freudenreichii [35]. Here, several genes involved in natural competence were found to be induced under the conditions applied. However, neither natural competence nor conjugation has been described in this species to date. Cultures of CIRM-BIA138 in presence of high concentration of insertional plasmid (data not shown) failed to illustrate natural competence.

## Utilization of pyrophosphate

*P. freudenreichii* can accumulate inorganic polyphosphate (polyP) as an energy reserve whereas most bacteria utilize ATP [9]. The ability to use polyphosphate as an energy reserve has been shown to be specific to bacteria adapted to difficult environments. The CIRM-BIA138 genome possesses 20 genes encoding enzymes which use polyphosphate or pyrophosphate. At the start of the stationary phase during our study (at 3d postinoculation), three phosphorylases were induced when compared to exponential growth phase at 1d post-inoculation: (i) ppx5, encoding an exopolyphosphatase with a 19.8 fold change, (ii) ppa encoding an inorganic pyrophosphatase with a 1.6 fold change, and (iii) PFCIRM138\_07685 encoding a NUDIX hydrolase with a 3 fold change. The phoH gene, encoding a phosphate-starvation inducible protein, was induced with a 2.5 fold change. In Corynebacterium glutamicum, another Actinobacterium that is phylogenetically close to P. freudenreichii, previous microarray results showed an induction of the phoH gene with a fold change ranging from 1.1 to 6.8 under Pi-limiting conditions compared to non-limiting conditions [28]. In many species, strains with a mutation in the gene involved in polyphosphate (ppk) synthesis are unable to survive during stationary phase [29]. Taken together, these results suggested a possible phosphate exhaustion of the medium, causing P. freudenreichii to use polyphosphates stored in its cytoplasm during stationary phase to cover its phosphate requirements. Since the phosphate starvation response is critical for the persistence of Mycobacterium tuberculosis (another Actinobacterium) in the lung [30], the role of phosphate starvation in the entry of *P. freudenreichii* into dormancy needs to be further explored, e.g. by quantifying extracellular phosphate and intracellular polyphosphate and by directed mutagenesis on key enzymes in the polyphosphate synthesis pathway.

## Conclusions

At entry into stationary phase, *P. freudenreichii* adapts its metabolism to nutritional shortage and slows down its metabolism. Genes involved in oxidative phosphorylation and fermentation (via the WoodWerkman cycle) are repressed, in line with the lack of lactate in the medium, enabling the entry into dormancy. By contrast, *P. freudenreichii* diversifies its source of nutrients and appears to utilize amino acids which differ from those used during the exponential phase according to RNAseq analysis, RT-qPCR and amino acids quantification. To meet its energy needs *P. freudenreichii* probably utilizes polyphosphate, because several phosphatases were found to be induced during this study. These results therefore provide an analysis of *P. freudenreichii* adaptation during entry into stationary phase by means of comprehensive gene expression analysis using RNA-Seq combined with targeted biochemical quantifications. They shed light on important molecular mechanisms that might be involved in the long-term survival of *P. freudenreichii*, and open avenues for further investigation on its survival strategies.

#### Methods

## **Bacterial strains**

*P. freudenreichii* CIRM-BIA138 strain was used during this work. The strain was supplied by the International Centre for Microbial Resources-Bacteria of Food Interest (Centre International de Ressources Microbiennes-Bactéries d'Intérêt Alimentaire; INRA, Rennes).

#### Conditions for bacterial growth

The strain was cultured in YEL medium (pH=7.0) [36] with no agitation at 30°C for 11 days. The strain was grown in a different assay tube for each manipulation, thus preventing any interference of oxygen in the environment and maintaining the strain under microaerophilic conditions. At T0, the medium was inoculated at $10^7$  CFU/mL.

The growth kinetics were followed using Optical Density measurements with a Model DU 640 spectrophotometer (Beckman Coulter, Fullerton, California, USA) at 650 nm (OD<sub>650</sub>), and the CFU (CFU/mL) were counted using the micromethod described previously [37]. *P. freudenreichii* enumerations were carried out on YEL agar at 30°C under microaerophilic conditions, until the visualization of colonies (6 days). Two technical replicates and two biological replicates were performed for growth curves analysis, and three biological replicates were performed for RNA-seq and biochemical analysis. Aliquots of the culture were sampled for RNA extraction, pH measurements and biochemical analysis (see above) on the day of inoculation, and 24h (1 day), 72h (3 days) and 216h (9 days) post-inoculation.

#### RNA extraction and quality control

1 ml of each culture was mixed with 2 volumes of RnaProtect (Qiagen, Hilden, Germany), left for 5 min. at room temperature and then centrifuged (8,000 g, 10 min at room temperature). The supernatant was removed and the pellet stored at -80°C until total RNA was extracted. The pellets were thawed on ice, suspended in 200 µL lysis buffer (50 mM Tris-HCl, 1 mM EDTA; pH 8.0) containing 20 mg/mL lysozyme (MP Biomedicals, Illkirch, France) and 50 U/mL mutanolysin (Sigma, Saint Quentin Fallavier, France), and incubated for 15 min at 24°C. The suspensions were then transferred to two millilitre tubes containing 50 mg zirconium beads (diameter: 0.1 mm; BioSpec Products, Bartlesville, OK) and 100 µL SDS (10%). The tubes were shaken twice for 90 s at 30 Hz with a bead beater (MM301; Retsch, Haan, Germany), being chilled on ice for 2 min. between the shaking steps. RNA extraction was then performed using an RNeasy minikit (Qiagen) and the Qiacube extraction robot (Qiagen), according to the manufacturer's instructions. RNA were suspended in 50 µL RNase-free water and treated with DNase (DNA-free; Ambion, Cambridgeshire, United Kingdom) according to the supplier's instructions, and then stored at -80°C until use. The RNA was then quantified and the contamination of RNA by proteins was assessed spectrophotometrically using a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies, Inc., Rockland, DE). RNA quality was evaluated using an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA). All the RNA samples from spent medium had a RIN value higher than 7.5, indicative of good rRNA integrity. RNA was then depleted using RiboZero Magnetic kit for Gram positive bacteria (Epicentre, Madison, WI, USA) according to the manufacturer's instructions. Depleted RNA quality was evaluated using an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA) and nanoarrays for prokaryotes.

#### Sequencing of mRNA

The cDNA libraries were prepared from the depleted RNA for each of the three repetitions of the exponential phase (1d post-inoculation), stationary phase (3d postinoculation) and long-term survival (9d post-inoculation) and then prepared for sequencing using the Illumina TruSeq Stranded mRNA LT Sample Preparation kit. The nine oriented RNA-seq libraries were prepared using the TruSeq Stranded mRNA Sample Preparation kit, according to the manufacturer's instructions. Qualitative and quantitative library validations were performed on a DNA 1000 Agilent chip as well as using quantitative PCR with SYBR Green (Applied Biosystem 7500). The libraries were sequenced on an Illumina HiSeq 2000 as 50 bp reads using the Sequence By Synthesis technique at the Montpellier GenomiX facility. Image analyses and base calling were performed using the Illumina HiSeq Control Software and Real-Time Analysis component. Demultiplexing was performed using Illumina's sequencing analysis software (CASAVA 1.8.2). Data quality was assessed using FastQC from the Babraham Institute and the Illumina software SAV (Sequencing Analysis Viewer). The concentration in the samples corresponding to long-term survival (9d post inoculation) were too low for sequencing and revealed contamination with genomic DNA, so they were not further analysed.

## RNA-seq analysis

Sequences were mapped with Bowtie [38] and counted with htseq-count [39] using the ABIMS Roscoff platform. A list of differentially expressed genes was generated using an R software package: SARtools https://github.com/PF2-pasteur-fr/SARTools developed by Institut Pasteur Paris that embedded Deseq2 [40] and EdgeR [41] (adjusted Pvalues<0.05). A comparison of the results obtained with either Deseq2 or

EdgeR produced a more exhaustive list using EdgeR. Most of the differentially expressed genes from Deseq2 were present in the EdgeR results. For this reason, only the results obtained using EdgeR are discussed below.

The protein sequences of the strain were subjected to a search against the Kyoto Encyclopedia of Genes and Genomes Pathway database (KEGG) using GhostKOALA (http://www.kegg.jp/ghostkoala/) to retrieve KEGG orthology (KO) identifiers. 1143 out of 2304 genes were annotated automatically. Differentially expression data were associated with these KO identifiers and underwent gene set enrichment analysis on KEGG orthology pathways using the GAGE package [42] of R software to highlight pathways that were significantly differentially expressed (pval <0.1).

#### **RT-qPCR** validation

*cst*A, *eno*1 and *sda*A were used as housekeeping genes. Relative quantification relates the PCR signal of the target transcript at 3d and 9d post-inoculation to that of inoculation time as control. Before, RT-qPCR, a supplementary DNAse treatment was applied to total RNA from 0, 3d and 9d post-inoculation samples. cDNA were obtained from a retrotranscription step using iScript cDNA synthesis kit (Bio-Rad, Marne la Coquette, France). cDNAs were amplified as follow: each PCR mixture included 5µl of 1/50 diluted cDNA, 3µl of 200 nM of primers, 8 µl of IQ Sybr Green supermix (Bio-Rad, Marne la Coquette, France). Amplifications were carried out on a CFX96 Real Time System (Bio-Rad) for 5 minutes at 95°C and 40 cycles of 2 steps consisting of 15 seconds at 95°C and 60 seconds at 60°C. The relative quantification of the mRNA levels of the target genes was determined using CFX Manager. The amount of target was normalized to *cst*A, *eno*1 and *sda*A genes because they were revealed as stably expressed according to the software: Delta Cq = Cq (target gene) – Cq (housekeeping gene), where Cq represents the cycle number required to reach a defined threshold target abundance.

#### Metabolome analysis

The samples used for this study had previously been frozen at -20°C. These analyses were performed on the following samples: (i) YEL culture medium, (ii) exponential (1d post-inoculation), (iii) stationary (3d post-inoculation), (iv) late stationary (9d post-inoculation) and (v) long-term survival (11d post-inoculation), with three biological replicates of each sample. The YEL culture medium was used as a control.

# ) Amino acid assays

Proteins and peptides in the samples were precipitated by adding solid sulphosalicylic acid to 5% (w/vol), holding at 4°C for 1 h, and centrifuged (5,000 g, 15 min). The supernatant was filtered through a 0.45 µm pore size filter and diluted with a 0.2 mol/L lithium citrate buffer (pH 2.2) before injection. Amino acid analyses were then carried out using cation exchange chromatography on a Biochrom 30 AA analyser (Biochrom Ltd, Cambridge, UK) according to the method described by Moore et al. (1958) with lithium citrate buffers as eluents and the ninhydrin post-column reaction system. Data were recovered using the control software Biosys and chromatographic data were processed with EZChrom Elite.

) Quantification of lactate, pyruvate, acetate, citrate and propionate

Lactic, propionic, acetic, succinic and pyruvic acids were quantified in culture supernatants using an HPLC Aminex A-6 ion exchange column (Bio-Rad, Hercules,

CA) at 60°C with 0.005 M H2SO4 at an eluent flow rate of 0.4 ml min-1. Acids were detected by UV (210 nm) and /or refractometry (RI2031 plus, Jasco).

#### • Quantification of arabinose and inositol

Concentrations were measured in culture supernatants by high performance liquid chromatography (Agilent Technologies 1200 Series, Waldbronn, Germany) using a HPX87H+ Biorad column and the following conditions: a temperature of 48 °C, eluent H2SO4 (5 mM) at a flow rate of 0.5 mL/min, and dual detection (refractometer and UV).

# ) Quantification of glucose, mannose, fructose and lactose

The supernatant of culture medium was deproteinized using Vivaspin 10 kDa filters. Glucose, mannose, fructose, and lactose were separated on a CarboPacMA1 (4 x 250 mm) analytical column (preceded by a corresponding guard column 50 x 4 mm) with 16 mM NaOH as the eluent and a flow rate of 1 mL/min using an ICS-3000 chromatography/detector module (Dionex) and a range of 1  $\mu$ C. A solution containing each of the four sugars (glucose, mannose, fructose and lactose) (Sigma-Aldrich) at 2, 5,10, 20 and 40 mg/L (linearity range) was used as a standard. Detection and quantification were performed using amperometry and expressed in  $\mu$ C.

## Availability of data and materials

The dataset supporting the conclusions of this article is available in the GEO repository, [GSE75553] in http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE75553].

## **Competing interests**

The authors declare that they have no competing interests.

#### **Authors' contributions**

FA and HF wrote the paper. SMD and AT participated in its compilation. GH, EB, and MBM quantified the amino acids, sugars and organic acids. HP compiled the library and performed the sequencing. MR undertook the demultiplexing and quality control of the reads obtained. MNM cared for the strain. SP performed quantitative PCR. AN performed the statistical analyses. HF mapped the RNA-Seq data and reconstructed the pathways. AM, VA and YLL supervised the work.

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**Figure 1:** DO, CFU, pH, sugar utilization and organic acid consumption and production during the growth and long term survival over 11 days of CIRM-BIA138 in YEL medium. (A) Bacterial enumeration (CFU/mL), and pH (B) Quantification of sugars (glucose: pink stars, mannose: purple bold pipes, fructose: light blue dashes, lactose: green pipes) for conversion to pyruvate (blue bow tie). (C) Quantification of acids (succinate: dark blue square, lactate: orange diamond, acetate: red circle, propionate: green triangle). The vertical dotted line indicates entry into stationary phase (3d post-inoculation).



**Figure 2**: Number of induced and repressed genes in the stationary phase (3d postinoculation) compared to the exponential phase (1d post-inoculation), classified by metabolic function. In blue and orange: induced and repressed genes, respectively. Differential comparison of groups (each containing three repetitions) were performed gene by gene using a modified t-test. Genes were declared as being differentially expressed (DE) with a P value 0.005.



**Figure 3**: Schematic view of the metabolism at the entry into stationary phase compared to exponential phase. Induced / repressed / 'non differentially expressed' genes are symbolized with green / red / black arrows respectively. (P value 0.005).



**Figure 4**: Concentrations of free amino acids in the medium (mM) during growth and stationary phases. In dark blue at the time of inoculation, in red at 1-day post inoculation, in green at 3-day post inoculation, in purple at 9-day post-inoculation, in light blue at 11-day post-inoculation. Significant consumptions discussed in the text (serine, asparagine, glycine, alanine) correspond to the difference in concentration at 1d and 3d post inoculation (t-test, Pvalue <0.05).

**Table S1** : Induced genes in stationary phase compared to exponential phase.

				log2Fold	_	
Name	Descript	Function	FC	Change	pvalue	padj
mraZ	Protein mraZ	1.1 Cell wall	1,839	0,879	0,01458547	0,03763326
gtfF	Polysaccharide synthase	1.1 Cell wall	6,169	2,625	0,0011783	0,00431847
slpE	Surface protein with SLH domain	1.1 Cell wall	12,903	3,69	2,7762E-07	2,965E-06
PFCIRM13 8 06855	Hypothetical outer membrane protein	1.1 Cell wall	4,992	2,32	0,00269909	0,00896146
gtfD	Glycosyl transferase, family 2	1.1 Cell wall	2,274	1,185	1,7102E-05	0,00011405
PFCIRM13			,	, , , , , , , , , , , , , , , , , , ,	,	,
8_07670	nucleoside-diphosphate-sugar epimerases	1.1 Cell wall	2,194	1,134	2,3011E-05	0,00014509
		1.10				
PFCIRM13		Transformation/compet	44750	0.000	0.44005.05	0 00040047
8_00630	DNA transfert protein	ence	14,752	3,883	2,1136E-05	0,00013617
		1.10 Transformation/compat				
dorA	DNA proposing / uptako protoin	Transformation/compet	27 262	4 774		1 20505 06
ирід	DNA processing / uplake protein		27,302	4,774	1,0090E-07	1,2959E-00
DECIDM13		Transformation/compet				
8 07015	Mobilisation protein	ence	49 447	5 628	4 7645E-40	5 2124E-37
0_07010		1 10	+5,++1	0,020	4,70432 40	5,21242 57
PECIRM13		Transformation/compet				
8 07020	Mobilisation protein	ence	10.855	3.44	6.5035E-22	1.423E-19
0_0.020		1.10	,	0,11	0,00001 11	.,
PFCIRM13		Transformation/compet				
8 07570	Conjugative relaxase domain protein, TrwC/Tral family	ence	24,025	4,586	2,7937E-06	2,4065E-05
		1.10	,	,	,	,
		Transformation/compet				
comEC	ComE operon protein 3	ence	17,39	4,12	1,2834E-06	1,2E-05
PFCIRM13		1.2 Transport/binding				
8_00130	Permease	proteins and	20,821	4,38	7,3817E-07	7,1465E-06

		lipoproteins				
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_00900	ABC transporter (Permease)	lipoproteins	12,247	3,614	7,9844E-05	0,0004261
		1.2 Transport/binding				
		proteins and				
sufS	Cysteine desulphurases, SufS	lipoproteins	1,956	0,968	0,00877986	0,02440956
		1.2 Transport/binding				
	ABC-type transport system involved in Fe-S cluster assembly, ATPase	proteins and				
sufC	component, SufC	lipoproteins	1,926	0,945	0,01558108	0,039707
		1.2 Transport/binding				
		proteins and				
sufD	FeS assembly protein SufD	lipoproteins	2,114	1,08	0,00534908	0,01603257
		1.2 Transport/binding				
		proteins and				
phnL	ATP binding protein of ABC transporter	lipoproteins	15,223	3,928	1,0113E-06	9,5378E-06
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_02385	permease protein	lipoproteins	50,478	5,658	1,6649E-06	1,5115E-05
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_02395	ABC transporter related precursor	lipoproteins	12,372	3,629	0,00015522	0,00075138
		1.2 Transport/binding				
PFCIRM13	Binding-protein-dependent transport systems inner membrane	proteins and				
8_03530	component	lipoproteins	2,246	1,168	0,00035727	0,00156031
		1.2 Transport/binding				
		proteins and				
yceJ	Transporter protein	lipoproteins	16,874	4,077	2,9259E-05	0,00017882
		1.2 Transport/binding				
PFCIRM13	Binding-protein-dependent transport systems inner membrane	proteins and				
8_05125	component	lipoproteins	10,809	3,434	0,00017228	0,00081767
		1.2 Transport/binding				
PFCIRM13	Binding-protein-dependent transport systems inner membrane	proteins and				
8_05130	component	lipoproteins	13,72	3,778	3,2365E-05	0,00019401
PFCIRM13		1.2 Transport/binding				
8_05560	Transporter protein	proteins and	6,834	2,773	0,00103039	0,00382765

		lipoproteins				
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_06235	Phosphocarrier protein, HPr family 8.A.8.1.1	lipoproteins	1,736	0,796	0,02059267	0,04945857
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_06470	ABC-type transporter, permease component	lipoproteins	3,067	1,617	0,00136804	0,00490701
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_06475	BP protein of siderophore ABC transporter (ISVH:undef)	lipoproteins	3,578	1,839	0,00071747	0,00280327
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_07645	transmembrane efflux protein	lipoproteins	19,201	4,263	1,4006E-05	9,6067E-05
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_08270	ABC transporter ATP-binding protein	lipoproteins	8,213	3,038	0,00100895	0,00375439
		1.2 Transport/binding				
		proteins and				
lmrB	Permease (Lincomycin resistance protein )	lipoproteins	13,929	3,8	0,00012318	0,00061956
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_08660	secretory protein	lipoproteins	10,949	3,453	0,00020008	0,00093742
		1.2 Transport/binding				
PFCIRM13	Arsenite efflux pump ACR3 and related permeases (Arsenite efflux	proteins and				
8_09385	pump ACR3 or related permease)	lipoproteins	3,001	1,586	0,0013183	0,00477556
		1.2 Transport/binding				
PFCIRM13		proteins and				
8_09450	major facilitator superfamily drug:H+ antiporter 2.A.1.3.x	lipoproteins	6,824	2,771	2,1795E-19	2,3844E-17
		1.2 Transport/binding				
		proteins and				
cynX	Cyanate MSF transporter	lipoproteins	3,305	1,725	0,00830715	0,02339258
		1.2 Transport/binding				
		proteins and				
pimH	Integral membrane efflux protein	lipoproteins	4,234	2,082	0,00105323	0,00390588
		1.2 Transport/binding				
hfIC2	Stomatin/prohibitin	proteins and	3,522	1,816	3,0476E-09	4,9394E-08
		lipoproteins				
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		1.2 Transport/binding				
PFCIRM13		proteins and	ı			
8_10665	ABC transporter ATP binding	lipoproteins	2,084	1,06	0,00756706	0,02162374
		1.2 Transport/binding				
PFCIRM13		proteins and	ı			
8_10695	IM-ABC protein of drug, protein or lipid ABC transporter (DPL:LLP)	lipoproteins	3,161	1,66	0,00117113	0,00430662
		1.2 Transport/binding				
PFCIRM13		proteins and	1			
8_12260	transporter, MFS superfamily protein	lipoproteins	11,237	3,49	7,5623E-05	0,00040654
PFCIRM13		1.2.3 Transport/binding				
8_05030	Cobalt permease	of inorganic ions	2,753	1,461	0,01803761	0,04454435
PFCIRM13		1.2.3 Transport/binding				
8_05035	ABC-type transporter, ATPase component	of inorganic ions	2,48	1,31	0,00253841	0,00849243
		1.2.3 Transport/binding				
mntH	Manganese transport protein	of inorganic ions	7,261	2,86	8,9333E-05	0,00047203
PFCIRM13		1.2.3 Transport/binding				
8_08475	nitrate transporter	of inorganic ions	1,88	0,911	0,00799075	0,02261805
	· · · ·	1.2.3 Transport/binding				
ssuC	permease protein of nitrate ABC transporter	of inorganic ions	4,413	2,142	0,00047049	0,00196081
PFCIRM13		1.2.3 Transport/binding				
8_08620	Heavy metal transport/detoxification protein	of inorganic ions	3,759	1,91	6,4909E-07	6,4263E-06
PFCIRM13		1.2.3 Transport/binding	1			
8_08625	Cation-transporting ATPase	of inorganic ions	4,248	2,087	5,1528E-08	6,5548E-07
PFCIRM13		1.2.3 Transport/binding				
8_09635	divalent anion:Na+ symporter 2.A.47.3.x	of inorganic ions	2,399	1,262	5,8332E-05	0,00032393
		1.2.3 Transport/binding				
feuS	binding protein of iron ABC transporter	of inorganic ions	2,805	1,488	0,00559075	0,01668836
		1.2.3 Transport/binding				
feuC	inner membrane protein of iron ABC transporter	of inorganic ions	2,468	1,303	0,01488094	0,03826028
PFCIRM13		1.2.3 Transport/binding				
8_11450	Sodium:sulfate symporter transmembrane region	of inorganic ions	4,306	2,106	0,0059313	0,01753638
PFCIRM13		1.2.4 Transport/binding				
8_04135	permease of the major facilitator superfamily	of carbohydrates	3,582	1,841	2,0847E-06	1,8467E-05
PFCIRM13	Membrane permease (sugar transporter)	1.2.4 Transport/binding	9.736	3.283	6.3679E-21	8.7081E-19

8_04640		of carbohydrates				
	PTS system mannose-specific EIIBCA component (EIIBCA-Man) (EII-	1.2.4 Transport/binding				
ptsG/ptsM	Man/EIII-Man)	of carbohydrates	2,025	1,018	0,00072156	0,00281422
		1.2.4 Transport/binding				
ptsl	PTS enzyme I	of carbohydrates	2,309	1,207	0,0001149	0,00058876
		1.2.4 Transport/binding				
yaaU	metabolite transport protein YaaU	of carbohydrates	4,656	2,219	0,00050151	0,00205875
		1.2.4 Transport/binding				
gudP	D-galactonate transporter	of carbohydrates	10,093	3,335	5,6868E-05	0,00031741
		1.2.4 Transport/binding				
ioIT1	ioIT1 (Major myo-inositol transporter ioIT1)	of carbohydrates	11,044	3,465	1,9084E-05	0,00012577
	inner membrane protein of ribose/xylose/arabinose/galactoside ABC	1.2.4 Transport/binding				
araH2	transporter	of carbohydrates	3,838	1,94	0,0083652	0,02352578
PFCIRM13		1.2.5 Transport/binding				
8_00815	Glutamate/gamma-aminobutyrate anti-porter (membrane protein)	of amino-acids	8,595	3,104	3,0355E-05	0,0001833
PFCIRM13		1.2.5 Transport/binding				
8_04335	polar amino acid ABC transporter, ATP-binding protein	of amino-acids	33,689	5,074	8,5244E-07	8,2165E-06
PFCIRM13		1.2.5 Transport/binding				
8_04340	polar amino acid ABC transporter, binding protein component	of amino-acids	10,43	3,383	0,00016304	0,00078057
PFCIRM13		1.2.5 Transport/binding				
8_04345	Polar amino acid ABC transporter, inner membrane subunit	of amino-acids	14,928	3,9	3,0824E-05	0,00018528
PFCIRM13		1.2.5 Transport/binding				
8_04350	polar amino acid ABC transporter, inner membrane protein	of amino-acids	16,067	4,006	3,041E-05	0,0001833
		1.2.5 Transport/binding				
lysE	Lysine exporter protein, membran protein	of amino-acids	4,953	2,308	0,01065826	0,02875496
PFCIRM13		1.2.5 Transport/binding				
8_07335	polar amino acid ABC transporter, binding protein	of amino-acids	3,294	1,72	0,00149863	0,00531441
PFCIRM13		1.2.5 Transport/binding				
8_07665	Amino acid permease-associated region	of amino-acids	2,317	1,212	0,00040015	0,00171335
PFCIRM13		1.2.5 Transport/binding				
8_10015	Putrescine importer	of amino-acids	3,541	1,824	1,1329E-09	2,0486E-08
		1.2.6 Transport/binding				
		of nucleosides,				
		nucleotides, purines				
cytX	Hydroxymethylpyrimidine transporter CytX	and pyrimidines	3,892	1,961	0,01324767	0,03458939

PFCIRM13		1.3 Sensors (signal				
8_08430	sensor kinase - Membrane protein	transduction	5,331	2,414	2,2078E-05	0,00014042
PFCIRM13		1.3 Sensors (signal				
8_10375	Sensory transduction histidine kinase	transduction)	1,555	0,637	0,00918295	0,02536905
PFCIRM13		1.3 Sensors (signal				
8_11140	Two-component system sensor kinase	transduction)	1,953	0,966	0,00048526	0,00201087
		1.4 Membrane				
		bioenergetics (electron				
PFCIRM13		transport chain and				
8_01505	Dioxygenase	ATP synthase)	1,987	0,99	0,00871079	0,02434126
		1.4 Membrane				
		bioenergetics (electron				
PFCIRM13		transport chain and				
8_07220	Cytochrome c biogenesis protein transmembrane region	ATP synthase)	13,347	3,738	9,8363E-05	0,00051243
		1.4 Membrane				
		bioenergetics (electron				
		transport chain and				
dmsC	Anaerobic dimethyl sulfoxide reductase chain C	ATP synthase)	13,75	3,781	0,000144	0,00070486
		1.4 Membrane				
		bioenergetics (electron				
		transport chain and				
dmsB	Anaerobic dimethyl sulfoxide reductase, chain B	ATP synthase)	4,764	2,252	0,00987035	0,02696171
		1.4 Membrane				
		bioenergetics (electron				
PFCIRM13		transport chain and				
8_08930	NAD(P)H-dependent FMN reductase	ATP synthase)	2,962	1,567	3,4375E-06	2,8707E-05
		1.4 Membrane				
		bioenergetics (electron				
	Coenzyme F420-dependent N5,N10-methylene	transport chain and				
mer	tetrahydromethanopterin reductase	ATP synthase)	3,124	1,643	2,7322E-07	2,9448E-06
		1.4 Membrane				
		bioenergetics (electron				
PFCIRM13		transport chain and				
8_09375	alkylglycerone-phosphate synthase (FAD linked oxidase-like protein)	ATP synthase)	10,084	3,334	2,5391E-14	9,9207E-13
PFCIRM13	NADPH:quinone reductase and related Zn-dependent	1.4 Membrane				
8_10905	oxidoreductases	bioenergetics (electron	1,718	0,781	0,01159156	0,030892

		transport chain and				
		1 4 Mombrane				
		hisepergetics (electron				
PECIRM13		transport chain and				
8 11130	NADPH guinone reductase related Zn-dependent oxidoreductase	ATP synthase)	2 623	1 391	2 2365E-05	0 00014184
PFCIRM13			2,020	1,001	2,20002 00	0,00011101
8 06860	Peptidase C60, sortase A and B precursor	1.6 Protein secretion	22.196	4.472	1.2374E-05	8.7054E-05
PFCIRM13			,	,	,	_,
8 07165	Tat pathway signal sequence domain protein	1.6 Protein secretion	15,425	3,947	1,3403E-05	9,2323E-05
		10.2 Protein of		· · ·		
		unknown function				
PFCIRM13		similar to proteins from				
8_00105	protein of unknown function	other organisms	2,054	1,039	0,01220287	0,03224623
PFCIRM13		2 Intermediary				
8_05230	Zinc-containing alcohol dehydrogenase	metabolism	15,475	3,952	3,329E-23	9,1049E-21
		2.1.1 Specific				
		carbohydrate metabolic				
gntK	gluconokinase	pathway	2,572	1,363	1,1071E-05	7,9608E-05
		2.1.1 Specific				
		carbohydrate metabolic				
galK1	Galactokinase	pathway	1,999	1	0,01117885	0,03001144
		2.1.1 Specific				
PFCIRM13		carbohydrate metabolic				
8_00835	Inositol 2-dehydrogenase	pathway	7,458	2,899	0,00060532	0,00242127
		2.1.1 Specific				
PFCIRM13		carbohydrate metabolic				
8_00840	sugar (xylose) phosphate isomerase/epimerase	pathway	14,326	3,841	0,00016245	0,00077946
		2.1.1 Specific				
		carbohydrate metabolic				
iolH	IoIH (Myo-inositol catabolism IoIH protein) (isomerase:epimerase)	pathway	23,005	4,524	3,6816E-05	0,00021713
		2.1.1 Specific				
		carbohydrate metabolic				
glgA	glycosyltransferase (glycogen synthase)	pathway	2,264	1,179	4,0563E-05	0,00023604
		2.1.1 Specific				
ioIT2	ioIT2 (myo-inositol transporter ioIT2)	carbohydrate metabolic	3,219	1,686	0,00447323	0,01370789

		pathway				
		2.1.1 Specific				
		carbohydrate metabolic				
iol	myo-inositol 2-dehydrogenase	pathway	5,559	2,475	0,00085922	0,00326954
		2.1.1 Specific				
PFCIRM13		carbohydrate metabolic				
8_03930	Glycoside hydrolase	pathway	2,345	1,229	0,0184331	0,04526557
		2.1.1 Specific				
		carbohydrate metabolic				
adh	alcohol dehydrogenase	pathway	10,194	3,35	0,00036667	0,001595
		2.1.1 Specific				
PFCIRM13		carbohydrate metabolic				
8_04360	hydroxyacid dehydrogenase	pathway	14,233	3,831	7,0972E-05	0,00038627
		2.1.1 Specific				
		carbohydrate metabolic				
aldB	Lactaldehyde dehydrogenase	pathway	2,665	1,414	1,13E-05	8,0798E-05
		2.1.1 Specific				
PFCIRM13		carbohydrate metabolic				
8_06500	Beta-glucosidase	pathway	1,621	0,696	0,01711601	0,04270219
		2.1.1 Specific				
PFCIRM13		carbohydrate metabolic				
8_07275	hexulose-6-phosphate isomerase	pathway	3,863	1,95	0,00708094	0,02033216
		2.1.1 Specific				
	4-alpha-glucanotransferase (Amylomaltase) (Disproportionating	carbohydrate metabolic				
malQ2	enzyme) (D-enzyme)	pathway	1,529	0,613	0,01739521	0,04319038
		2.1.1 Specific				
		carbohydrate metabolic				
galE3	UDP-glucose 4-epimerase	pathway	11,493	3,523	0,00010621	0,0005494
		2.1.1 Specific				
	iolG2 (Myo-inositol catabolism IolG2 protein) (Inositol 2-	carbohydrate metabolic				
iolG2	dehydrogenase)	pathway	2,114	1,08	0,0116371	0,03097563
		2.1.1 Specific				
		carbohydrate metabolic				
ioIE3	Myo-inositol catabolism protein	pathway	2,26	1,176	0,00100895	0,00375439
		2.1.1 Specific				
araD1	L-ribulose-5-phosphate 4-epimerase	carbohydrate metabolic	2,869	1,52	0,007871	0,02233689

		pathway				
		2.1.1 Specific				
		carbohydrate metabolic				
araA	L-arabinose isomerase	pathway	2,187	1,129	0,01676094	0,04215279
		2.1.2 Main glycolytic				
fba2	Fructose-bisphosphate aldolase class I	pathways	3,651	1,868	5,6726E-09	8,6795E-08
		2.1.2 Main glycolytic				
rpiB3	Ribose-5-phosphate isomerase 3	pathways	2,879	1,526	3,1493E-06	2,6503E-05
		2.1.2 Main glycolytic				
acyP	Putative acylphosphatase	pathways	5,76	2,526	0,0016336	0,00574651
		2.1.2 Main glycolytic				
araB	L-ribulokinase	pathways	2,158	1,11	1,2915E-05	8,999E-05
		2.1.2 Main glycolytic				
eno2	Enolase 2	pathways	3,442	1,783	1,2064E-08	1,7598E-07
		2.1.4 Substrate-specific				
PFCIRM13		entries to carbohydrate				
8_07350	hydroxypyruvate isomerase	metabolic pathway	15,014	3,908	1,0201E-05	7,4646E-05
		2.2 Metabolism of				
	3-isopropylmalate dehydratase large subunit (Isopropylmalate	amino acids and				
leuC	isomerase) (Alpha-IPM isomerase) (IPMI)	related molecules	2,413	1,271	0,00049172	0,00202233
		2.2 Metabolism of				
		amino acids and				
gadB	Glutamate decarboxylase	related molecules	4,997	2,321	2,3166E-06	2,0194E-05
		2.2 Metabolism of				
	Chorismate synthase (5-enolpyruvylshikimate-3-phosphate	amino acids and				
aroF	phospholyase)	related molecules	1,569	0,65	0,01874808	0,0457311
		2.2 Metabolism of				
	S-adenosylmethionine synthetase (Methionine adenosyltransferase)	amino acids and				
metK	(AdoMet synthetase) (MAT)	related molecules	1,608	0,685	0,02029457	0,04884985
		2.2 Metabolism of				
		amino acids and				
ald	Alanine dehydrogenase	related molecules	2,668	1,416	6,0251E-05	0,0003329
		2.2 Metabolism of				
		amino acids and				
dadA2	DadA, Glycine/D-amino acid oxidases	related molecules	12,994	3,7	4,9033E-06	3,9442E-05

		2.2 Metabolism of				
PFCIRM13		amino acids and				
8_04365	aminotransferase	related molecules	10,633	3,41	0,00021657	0,00100392
		2.2 Metabolism of				
PFCIRM13		amino acids and				
8_05235	Carbamoylphosphate synthase large subunit CarB	related molecules	4,228	2,08	7,2777E-06	5,5677E-05
	3-phosphoshikimate 1-carboxyvinyltransferase	2.2 Metabolism of			-	
	(phosphoenolpyruvate:3-phosphoshikimate5-O-(1-carboxyvinyl)-	amino acids and				
aroA	transferase) (EPSP synthase)	related molecules	1,672	0,742	0,00276818	0,00912165
		2.2 Metabolism of	,	,	,	,
		amino acids and				
dapA1	Dihydrodipicolinate synthetase	related molecules	38.828	5.279	2.6405E-07	2.8602E-06
		2.2 Metabolism of		-,		
		amino acids and				
leuA1	2-isopropylmalate synthase	related molecules	1.676	0.745	0.00347887	0.01090968
		2.3 Metabolism of	.,	-,		-,
		nucleotides and nucleic				
nnx	Ppx/GppA phosphatase family	acids	1 62	0 696	0 02002399	0 04827567
		2 3 Metabolism of	1,02	0,000	0,02002000	0,01021001
		nucleotides and nucleic				
nrdR	Transcriptional repressor prdR	acids	1 728	0 789	0 01925179	0 04669946
mart		2.3 Metabolism of	1,720	0,100	0,01020110	0,01000010
	Phosphomethylpyrimidine kinase/ hydroxymethylpyrimidine	nucleotides and nucleic				
thiD1	nhosphokinase	acids	4 07	2 025	0.0111065	0 02989055
		2.3 Metabolism of	4,07	2,020	0,0111000	0,02000000
		nucleotides and nucleic				
dat	Deoxyguanosinetrinhosnhate trinhosnhohydrolase (dGTPase)	acide	1 822	0 866	0 00312000	0 00006850
ugi		2.3 Metabolism of	1,022	0,000	0,00312333	0,00330033
		2.5 Metabolishi of				
prt	nhachharibacultransfaraca		6 779	2 761	0.00026501	0.00110208
pr		2.2 Motobolism of	0,770	2,701	0,00020501	0,00119308
		2.5 Metabolishi ol				
neoB2	A deputer uncipate luces		11 011	2 574	0.00021726	0.00100400
puadz			11,911	3,374	0,00021720	0,00100499
pf1655	Carboxylic ester hydrolase	2.4 Metabolism of lipids	1,511	0,595	0,01741038	0,04319038
pf169	Carboxylic ester hydrolase	2.4 Metabolism of lipids	2,308	1,207	0,00380039	0,01182825

PFCIRM13						
8_03645	Esterase family protein	2.4 Metabolism of lipids	65,916	6,043	4,4128E-11	1,0495E-09
	Crotonobetainyl-CoA dehydrogenase (Crotonobetainyl-CoA				–	
caiA	reductase)	2.4 Metabolism of lipids	15,443	3,949	1,006E-05	7,386E-05
caiB	Crotonobetainyl-CoA:carnitine CoA-transferase	2.4 Metabolism of lipids	3,281	1,714	0,00683535	0,0197566
caiC	Crotonobetaine/carnitine-CoA ligase	2.4 Metabolism of lipids	12,384	3,63	4,7644E-06	3,8467E-05
	Undecaprenyl pyrophosphate synthase (di-trans,poly-cis-					
uppS	decaprenylcistransferase)	2.4 Metabolism of lipids	1,789	0,839	0,00412185	0,01270227
glxK/garK	Glycerate kinase GlxK/GarK	2.4 Metabolism of lipids	2,794	1,482	2,8322E-08	3,8731E-07
plsC2	PIsC, Phosphate acyltransferases	2.4 Metabolism of lipids	2,069	1,049	0,00527008	0,01581748
		2.5 Metabolism of				
		coenzymes and				
chll	Magnesium chelatase, subunit Chll	prosthetic groups	8,331	3,058	5,9295E-05	0,00032845
		2.5 Metabolism of				
PFCIRM13		coenzymes and				
8_04810	Isochorismatase hydrolase	prosthetic groups	1,848	0,886	0,01286699	0,03379709
		2.5 Metabolism of				
		coenzymes and				
hemX	Membrane-bound transport protein (HemX protein)	prosthetic groups	16,735	4,065	2,6276E-05	0,00016287
		2.5 Metabolism of				
		coenzymes and				
pdxA2	4-hydroxythreonine-4-phosphate dehydrogenase	prosthetic groups	2,549	1,35	0,00083498	0,00319352
		2.5 Metabolism of				
		coenzymes and				
pncA	pyrazinamidase/nicotinamidase	prosthetic groups	2,501	1,322	3,4305E-06	2,8707E-05
		2.5 Metabolism of				
		coenzymes and				
cbiO3	ABC transporter, ATP-binding protein	prosthetic groups	5,571	2,478	0,00229833	0,00776041
		2.5 Metabolism of				
	ABC-type transporter, permease components (ABC-type cobalt	coenzymes and				
cbiQ1	transport system, permease component CbiQ)	prosthetic groups	9,615	3,265	0,00026681	0,00119871
		2.5 Metabolism of				
		coenzymes and				
mog	Molybdopterin biosynthesis Mog protein	prosthetic groups	2,792	1,481	0,00011522	0,00058903
thiC	thiamine-phosphate diphosphorylase	2.5 Metabolism of	2,361	1,239	0,0156067	0,039707

1		i i				
		coenzymes and				
		prosthetic groups				
		2.5 Metabolism of				
PFCIRM13		coenzymes and				
8_09275	NADH-flavin reductase	prosthetic groups	1,801	0,849	0,0066313	0,01926864
		2.5 Metabolism of				
		coenzymes and				
bioB	Biotin synthase	prosthetic groups	7,499	2,907	4,262E-27	1,5542E-24
		2.6 Metabolism of				
ppx5	Exopolyphosphatase	phosphate	19,842	4,31	9,256E-06	6,8885E-05
		2.6 Metabolism of				
рра	Inorganic pyrophosphatase	phosphate	1,579	0,659	0,01239683	0,03267982
PFCIRM13		2.6 Metabolism of				
8_07685	NUDIX hydrolase	phosphate	3,022	1,595	0,00035576	0,00155682
		2.6 Metabolism of				
phoH	Phosphate starvation-inducible protein PhoH-like protein	phosphate	2,521	1,334	4,7091E-05	0,00027115
•		2.8 Metabolism of			,	
		nitrogen/nitrate and				
ntcA2	Global nitrogen regulator NtcA	nitrite	1,732	0,792	0,00180183	0,00626775
		2.8 Metabolism of	,	,	,	,
PFCIRM13		nitrogen/nitrate and				
8 04265	Nitrate reductase, alpha subunit	nitrite	5.948	2.572	0.00096256	0.00361871
		2.8 Metabolism of		, -		-,
		nitrogen/nitrate and				
narH	Respiratory nitrate reductase, beta subunit	nitrite	22.415	4.486	3.9715E-07	4.1183E-06
		2.8 Metabolism of	,	.,	0,01102 01	.,
		nitrogen/nitrate and				
narJ	Nitrate reductase delta subunit	nitrite	22,951	4.52	3.0582E-06	2.5835E-05
		2.8 Metabolism of	,	.,•=	0,00012.00	_,0000_00
		nitrogen/nitrate and				
narl	Respiratory nitrate reductase, gamma subunit	nitrite	16.346	4 031	7 471E-06	5 6957E-05
nan		2.8 Metabolism of	10,010	1,001	7,1712.00	0,00072.00
		nitrogen/nitrate and				
alnB	Nitrogen regulatory protein P-II	nitrite	4 518	2 176	0 00150854	0 00534091
gind Las EQ			4,010	2,170	0,00100004	0,00007091
dnaE2	UNA polymerase III alpha subunit	3.1 DNA replication	2,304	1,204	0,00804479	0,0227416

priA	Primosomal protein N	3.1 DNA replication	2,979	1,575	5,5968E-09	8,6311E-08
gyrB2	DNA gyrase subunit B	3.1 DNA replication	2,435	1,284	1,9573E-05	0,00012784
hels	Helicase	3.1 DNA replication	2,469	1,304	5,1448E-07	5,2115E-06
PFCIRM13						
8_08320	helicase	3.1 DNA replication	2,774	1,472	6,7193E-08	8,2594E-07
dnaB	Replicative DNA helicase	3.1 DNA replication	1,552	0,635	0,01620308	0,04103201
PFCIRM13	Membrane protease subunits, stomatin/prohibitin homologs	3.10 Protein				
8_09485	(Membrane protease subunit, stomatin/prohibitin homolog)	degradation	2,516	1,331	1,4334E-05	9,77E-05
		3.10 Protein				
pepC	Aminopeptidase C, Bleomycin hydrolase,	degradation	1,601	0,679	0,0092136	0,02542164
		3.2 DNA restriction and				
		modification (and				
tagA	3-Methyladenine DNA glycosylase I	repair)	3,915	1,969	0,01295853	0,03397163
		3.2 DNA restriction and				
		modification (and				
sms	DNA repair protein	repair)	1,904	0,929	0,00144176	0,00513773
		3.2 DNA restriction and				
		modification (and				
sbcD	SbcD, DNA repair exonuclease	repair)	2,785	1,478	2,8782E-06	2,4504E-05
		3.2 DNA restriction and				
PFCIRM13		modification (and				
8_10795	exonuclease	repair)	1,828	0,87	0,01437546	0,03722308
		3.3 DNA				
PFCIRM13		recombination, and				
8_00325	Recombinase	repair	77,776	6,281	6,6498E-07	6,4955E-06
		3.3 DNA				
		recombination, and				
gin	Resolvase	repair	1,823	0,866	0,01302721	0,03409513
		3.3 DNA				
PFCIRM13		recombination, and				
8_00650	Recombinase	repair	7,929	2,987	9,3006E-18	7,2678E-16
		3.3 DNA				
		recombination, and				
recG	ATP-dependent DNA helicase RecG	repair	1,74	0,799	0,00307427	0,0098197
dinP	DNA-directed DNA polymerase	3.3 DNA	5,915	2,564	0,0006119	0,00243761

		recombination, and repair				
		3.3 DNA				
PFCIRM13		recombination, and				
8 01865	Helicase protein	repair	1,806	0.853	0,00091498	0,00345169
		3.3 DNA				
		recombination, and				
recA	RecA (Recombinase A)	repair	2,019	1,014	0,0023503	0,00790909
		3.3 DNA				
PFCIRM13		recombination, and				
8_03835	Bifunctional DNA primase/polymerase	repair	18,618	4,219	6,2207E-43	1,3611E-39
		3.3 DNA				
		recombination, and				
rmuC	DNA recombination protein	repair	3,851	1,945	2,0143E-10	4,3208E-09
		3.3 DNA				
		recombination, and				
sbcC	ATP-dependent dsDNA exonuclease SbcC	repair	1,712	0,776	0,00302292	0,00971241
		3.3 DNA				
	AlkA, 3-methyladenine DNA glycosylase/8-oxoguanine DNA	recombination, and				
alkA	glycosylase	repair	7,574	2,921	0,00061275	0,00243761
		3.3 DNA				
PFCIRM13		recombination, and				
8_10165	ATP-dependent DNA helicase	repair	2,875	1,524	6,3437E-07	6,3091E-06
		3.3 DNA				
		recombination, and				
urvD	UvrD/REP helicase / ATP-dependent DNA helicase	repair	1,534	0,617	0,0186472	0,04558667
		3.3 DNA				
		recombination, and				
holA	DNA polymerase III, delta subunit	repair	3,066	1,617	1,1811E-07	1,3969E-06
		3.5.1 Transcription				
hrdD2	RNA polymerase principal sigma factor HrdD	initiation	6,278	2,65	3,3471E-10	6,909E-09
		3.5.1 Transcription				
hrdD1	RNA polymerase principal sigma factor HrdD	initiation	8,762	3,131	0,00020197	0,00094223
		3.5.1 Transcription				
rpoE	RNA polymerase sigma -70 factor, sigma-E factor	initiation	1,875	0,907	0,01422102	0,03686681
marR3	Transcriptional regulator, MarR family	3.5.2 Transcription	3,689	1,883	0,01723654	0,04295393

		regulation				
PFCIRM13		3.5.2 Transcription				
8_00405	Transcriptional regulator	regulation	3,214	1,684	6,4352E-06	4,993E-05
		3.5.2 Transcription				
arsR1	Transcriptional regulator, ArsR family protein	regulation	6,938	2,794	1,0324E-05	7,5048E-05
		3.5.2 Transcription				
dtxR	Iron-dependent repressor	regulation	1,512	0,596	0,01689512	0,0424186
PFCIRM13		3.5.2 Transcription				
8_00890	transcription regulator	regulation	3,484	1,801	6,3071E-06	4,9285E-05
		3.5.2 Transcription				
arsR4	Transcriptional regulator, ArsR family	regulation	4,063	2,022	3,0203E-08	4,1047E-07
		3.5.2 Transcription				
pspA	Phage shock protein A	regulation	1,849	0,887	0,00592449	0,01753638
PFCIRM13		3.5.2 Transcription				
8_02505	Helix-turn-helix XRE-family like proteins.	regulation	11,769	3,557	4,248E-17	2,6556E-15
		3.5.2 Transcription				
arsR3	Transcriptional regulators (Bacterial regulatory protein, arsR family)	regulation	9,029	3,174	0,00028321	0,00126462
		3.5.2 Transcription				
tetR1	Transcriptional regulator, TetR family	regulation	3,006	1,588	7,0767E-05	0,00038613
		3.5.2 Transcription				
merR2	Transcriptional regulator, MerR	regulation	38,449	5,265	8,6754E-07	8,3253E-06
		3.5.2 Transcription				
hemR	HemR	regulation	3,469	1,795	0,00848484	0,02380106
		3.5.2 Transcription				
regX	#N/D	regulation	1,621	0,697	0,02065629	0,04955698
		3.5.2 Transcription				
padR	PadR, Transcriptional regulator PadR-like family	regulation	7,325	2,873	5,5452E-21	8,0885E-19
PFCIRM13		3.5.2 Transcription				
8_06840	transcriptional regulator ArsR	regulation	6,121	2,614	2,917E-09	4,763E-08
PFCIRM13		3.5.2 Transcription				
8_07310	Transcriptional regulator PadR family	regulation	3,263	1,706	5,5842E-05	0,00031248
		3.5.2 Transcription				
lacl2	Transcriptional regulator, Lacl family protein	regulation	3,31	1,727	0,00174572	0,00611924
PFCIRM13		3.5.2 Transcription				
8_08435	Two-component system, regulatory protein	regulation	2,948	1,56	0,00297648	0,00959137

		3.5.2 Transcription				
lysR1	Transcriptional regulator, LysR family protein	regulation .	4,075	2,027	7,0714E-06	5,4288E-05
PFCIRM13		3.5.2 Transcription				
8_09520	Regulatory protein, MarR	regulation	3,825	1,936	0,00578025	0,01720705
PFCIRM13		3.5.2 Transcription				
8_10090	two-component system response regulator	regulation	2,183	1,126	0,01610133	0,04082237
PFCIRM13		3.5.2 Transcription				
8_10370	anti-sigma factor	regulation	1,776	0,829	0,00445461	0,01366998
		3.5.2 Transcription				
whiB2	transcription factor WhiB	regulation	2,029	1,021	0,00235321	0,00790909
		3.5.2 Transcription				
tetR2	Transcriptional regulator, TetR/AcrR family	regulation	1,757	0,813	0,01694616	0,04247217
PFCIRM13		3.5.2 Transcription				
8_11280	Silent information regulator protein Sir2 NAD-dependent deacetylase	regulation	2,118	1,083	0,00929407	0,02561138
PFCIRM13		3.5.2 Transcription				
8_12035	MarR-family transcriptional regulator	regulation	4,754	2,249	0,00176852	0,00617148
		3.5.2 Transcription				
hrcA	Heat-inducible transcription repressor hrcA	regulation	4,506	2,172	4,2318E-12	1,2183E-10
	DNA-directed RNA polymerase omega chain (RNAP omega subunit)	3.5.3 Transcription				
rpoZ	(Transcriptase omega chain) (RNA polymerase omega subunit)	elongation	1,805	0,852	0,00879443	0,02441906
PFCIRM13		3.5.4 Transcription				
8_10430	Transcription termination factor Rho	termination	2,007	1,005	0,0006899	0,00271004
PFCIRM13						
8_10140	DEAD/DEAH box helicase domain protein	3.6 RNA modification	1,599	0,677	0,01455087	0,03763167
trmU	tRNA (5-methylaminomethyl-2-thiouridylate)-methyltransferase	3.6 RNA modification	2,571	1,362	0,00022788	0,00104969
infB	Translation initiation factor IF-2	3.7 Protein synthesis	2,009	1,007	0,00195859	0,00671962
		3.7.1 Ribosomal	,	,	,	
rpIS	50S ribosomal protein L19	proteins	1,882	0,912	0,00473468	0,01438816
•		3.7.1 Ribosomal		,		
rpmB	Ribosomal protein L28	proteins	3,235	1,694	2,0683E-05	0,00013389
•		3.7.1 Ribosomal		,		
rpsO	30S ribosomal protein S15	proteins	2,799	1,485	9,3926E-06	6,9429E-05
	•	3.7.1 Ribosomal				
rplK	50S ribosomal protein L11	proteins	1,802	0,85	0,01296449	0,03397163
rpmA	50S ribosomal protein L27	3.7.1 Ribosomal	2,126	1,088	0,00050893	0,00208527

		proteins				
PFCIRM13	translation initiation inhibitor involved in amino acid or purine	3.7.3 Translation				
8_04330	metabolism	initiation	5,469	2,451	0,00589094	0,01748898
		3.7.5 Translation				
prfA	Peptide chain release factor 1 (RF-1)	termination	2,113	1,079	0,00119823	0,00437685
PFCIRM13		3.7.6 Nonribosomal				
8_06630	Isochorismatase hydrolase	protein synthesis	2,021	1,015	0,01212049	0,03206726
trxA4	Thioredoxine	3.8 Protein modification	3,519	1,815	2,0155E-05	0,00013125
gst	Glutathione S-transferase	3.8 Protein modification	2,489	1,315	3,3022E-05	0,00019634
trxB	Thioredoxin reductase	3.8 Protein modification	2,696	1,431	0,00071228	0,00278797
PFCIRM13						
8_06730	AarF	3.8 Protein modification	3,156	1,658	1,2202E-09	2,1583E-08
PFCIRM13						
8_07240	Thioredoxin	3.8 Protein modification	16,925	4,081	5,1511E-05	0,00029198
htrA4	Serine protease	3.8 Protein modification	3,344	1,742	7,2864E-09	1,0995E-07
msrA	peptide-methionine (S)-S-oxide reductase	3.8 Protein modification	1,818	0,862	0,01140032	0,03048767
PFCIRM13						
8_12360	Amino acid-binding ACT	3.8 Protein modification	1,909	0,933	0,00133699	0,00482729
copZ	Copper chaperone CopZ	3.9 Protein folding	2,555	1,353	2,856E-05	0,00017553
hsp20 3	Heat shock protein 20 3 (20 kDa chaperone 3)	3.9 Protein folding	6,775	2,76	2,3131E-15	1,1247E-13
hsp20 1	Heat shock protein 20 1 (20 kDa chaperone 1)	3.9 Protein folding	6,955	2,798	6,2693E-15	2,8578E-13
hsp20 2	Heat shock protein 20 2 (20 kDa chaperone 2)	3.9 Protein folding	2,555	1,353	0,00099365	0,00371643
		4.1 Adaptation to			,	
cspB	Cold shock protein CspB	atypical conditions	2,664	1,414	2,4027E-06	2,0779E-05
PFCIRM13		4.1 Adaptation to				
8_04080	drug exporters of the RND superfamily	atypical conditions	1,768	0,822	0,00882506	0,02447305
		4.1 Adaptation to				
osmC	OsmC protein	atypical conditions	2,669	1,416	4,9954E-05	0,00028463
PFCIRM13		4.1 Adaptation to				
8_04225	Multidrug resistance transporter, MFS superfamily protein	atypical conditions	16,316	4,028	1,252E-05	8,78E-05
PFCIRM13		4.1 Adaptation to			· · ·	
8_04645	Cyclase family protein	atypical conditions	35,816	5,163	6,7683E-11	1,5589E-09
katA	Catalase	4.1 Adaptation to	2,129	1,09	0,00057545	0,00232305

		atypical conditions				
		4.1 Adaptation to				
dps	Starvation-inducible DNA-binding protein	atypical conditions	13,789	3,785	7,7027E-21	9,9139E-19
		4.1 Adaptation to				<b>-</b>
pspC	Possible stress-response transcriptional regulator protein PspC	atypical conditions	4,094	2,034	2,6719E-08	3,6767E-07
сорА	Copper-Cadmimum efflux ATPase	4.2 Detoxification	1,695	0,762	0,00195938	0,00671962
drrA	Daunorubicin resistance ATP-binding protein DrrA	4.2 Detoxification	20,788	4,378	4,2764E-06	3,4913E-05
PFCIRM13						
8_03890	Drug resistance transporter	4.2 Detoxification	6,494	2,699	0,00060844	0,00242933
PFCIRM13						
8_04715	Aldo/keto reductases, related to diketogulonate reductase	4.2 Detoxification	3,531	1,82	1,9954E-08	2,8167E-07
ahpF	Alkyl hydroperoxide reductase subunit F	4.2 Detoxification	1,883	0,913	0,01456765	0,03763167
PFCIRM13						
8_06410	Beta-lactamase-like	4.2 Detoxification	6,756	2,756	0,00266337	0,00886978
matE	Multi antimicrobial extrusion protein MatE	4.2 Detoxification	8,165	3,029	0,00011774	0,0005991
PFCIRM13						
8_06835	Hemerythrin HHE cation binding region	4.2 Detoxification	4,86	2,281	5,3397E-11	1,2429E-09
	Iron/Manganese superoxide dismutase (Superoxide dismutase					
sodA	[Mn/Fe]) (SODM)	4.2 Detoxification	9,425	3,237	1,7633E-14	7,0149E-13
PFCIRM13						
8_09705	Drug resistance transporter	4.2 Detoxification	4,418	2,143	0,00087606	0,00332205
pcaC1	4-carboxymuconolactone decarboxylase	4.2 Detoxification	7,442	2,896	0,00347011	0,01090891
PFCIRM13		4.3 Antibiotic				
8_00720	Isochorismatase hydrolase	production	1,742	0,8	0,00459481	0,01400202
PFCIRM13		4.4 Phage-related				
8_03830	Primase	function	18,861	4,237	1,6746E-32	9,3895E-30
PFCIRM13		4.4 Phage-related				
8_03840	Gp42	function	6,283	2,651	5,6222E-08	7,0697E-07
PFCIRM13						
8_00080	Transposase	4.5 Transposon and IS	7,242	2,856	5,3402E-05	0,00030115
tnpA	Transposase	4.5 Transposon and IS	2,485	1,314	1,721E-05	0,00011411
PFCIRM13						
8_06175	Transposase for IS3514b	4.5 Transposon and IS	15,427	3,947	1,0869E-09	1,9985E-08
tnp	Transposase	4.5 Transposon and IS	1,905	0,93	0,00868827	0,02430937

tnp7109-55	Transposase	4.5 Transposon and IS	3,073	1,62	5,4012E-05	0,0003038
TnpD	Transposase IS3/IS911	4.5 Transposon and IS	6,567	2,715	1,9076E-06	1,7106E-05
PFCIRM13		•				
8_00060	Dehydrogenase	4.6 Miscellaneous	6,654	2,734	0,00075901	0,00293411
PFCIRM13						
8_02755	acetyltransferase	4.6 Miscellaneous	12,363	3,628	0,00075805	0,00293411
PFCIRM13						
8_03090	LMBE-related protein	4.6 Miscellaneous	1,705	0,77	0,01026047	0,02781214
PFCIRM13						
8_05685	Zinc-containing alcohol dehydrogenase superfamily	4.6 Miscellaneous	3,604	1,85	0,00585402	0,01740297
PFCIRM13						
8_07990	GCN5-related N-acetyltransferase	4.6 Miscellaneous	1,742	0,801	0,00487411	0,01476462
PFCIRM13						
8_08590	Methyltransferase	4.6 Miscellaneous	8,446	3,078	0,00074434	0,00288761
PFCIRM13						
8_09515	Acetyltransferase	4.6 Miscellaneous	4,163	2,058	0,01734942	0,04313696
PFCIRM13						
8_10260	acetyltransferase	4.6 Miscellaneous	5,637	2,495	8,6977E-15	3,8061E-13
PFCIRM13						
8_10305	Glycosyl transferase	4.6 Miscellaneous	14,016	3,809	9,6463E-05	0,00050373
PFCIRM13						
8_10670	Methyltransferase	4.6 Miscellaneous	3,148	1,654	0,00083633	0,00319352
PFCIRM13						
8_10815	acetyltransferase	4.6 Miscellaneous	6,42	2,682	1,804E-05	0,00011925
PFCIRM13						
8_11765	Metal dependent phosphohydrolase	4.6 Miscellaneous	11,77	3,557	1,3418E-06	1,2493E-05
obg	GTPase	4.6 Miscellaneous	1,61	0,687	0,01497509	0,03845716
PFCIRM13						
8_12295	Metal-dependent hydrolase	4.6 Miscellaneous	3,582	1,841	0,00239099	0,00802374
		5.1 Protein of unknown				
		function similar to other				
PFCIRM13		proteins from the same				
8_00330	protein of unknown function	organism	58,871	5,87 <u>9</u>	4,0881E-06	3,3627E-05
PFCIRM13		5.1 Protein of unknown				
8_00620	protein of unknown function	function similar to other	81,837	6,355	2,8665E-09	4,7158E-08

		proteins from the same				
		5 1 Protein of unknown				
		function similar to other				
PECIRM13		proteins from the same				
8 00960	Hypothetical membrane protein	organism	1 732	0 792	0.0169054	0 0424186
0_00000		5.1 Protein of unknown	1,702	0,702	0,0100004	0,0424100
		function similar to other				
PECIRM13		proteins from the same				
8 03905	protein of unknown function	organism	5 637	2 4 9 5	0 00121989	0 00444855
0_00000		5 1 Protein of unknown	0,007	2,400	0,00121303	0,00444000
		function similar to other				
DECIDM13		proteins from the same				
8 05220	Hypothetical secreted protein	organism	2 715	1 1 1 1	1 68/9E-05	0 00011300
0_03220		5.2 Protein of unknown	2,715	1,771	1,00492-00	0,00011303
		function similar to				
		notoing from other				
	Hypothetical mombrane protein	organisms	7 6 9 7	2 0 4 2	0 00021502	0.00120026
0_00000		5.2 Drotoin of unknown	7,007	2,942	0,00031392	0,00139920
		5.2 FIOLEIN OF UNKNOWN				
		nunction similar to				
	Hypothetical protein	proteins nom other	2 4 1 2	1 071	0 00202700	0 01217242
0_00310		5 2 Drotoin of unknown	2,413	1,271	0,00392799	0,01217343
		5.2 Protein of unknown				
		runction similar to				
PFCIRM13	protoin of unknown function	proteins from other	40.050	5 225		
8_00355		organisms	40,353	5,335	5,1216E-07	5,2115E-06
		5.2 Protein of unknown				
DEOIDIMA		function similar to				
PFCIRM13		proteins from other	0.070	4 500		
8_00410	Hypothetical protein	organisms	2,872	1,522	8,5136E-06	6,4013E-05
		5.2 Protein of unknown				
DEOIDMAG		function similar to				
PFCIRM13		proteins from other	1 000	o ===	0.04550454	0.000707
8_00535	Hypothetical protein	organisms	1,689	0,757	0,01559171	0,039707
PFCIRM13		5.2 Protein of unknown				
8_00585	protein of unknown function	function similar to	20,592	4,364	1,6995E-05	0,00011371

		proteins from other				
		5.2 Protein of unknown				
		function similar to				
PECIRM13		proteins from other				
8 00590	protein of unknown function	organisms	2 6 1 7	1 388	2 1856E-05	0 00013942
0_00000		5.2 Protein of unknown	2,017	1,000	2,10002.00	0,00010012
		function similar to				
PECIRM13		proteins from other				
8 00605	protein of unknown function	organisms	13 578	3 763	5 7204E-05	0 00031848
0_00000		5.2 Protein of unknown	10,070	0,700	0,72042 00	0,00001040
		function similar to				
PECIRM13		proteins from other				
8 00765	Hypothetical protein	organisms	2 107	1 075	0 00036578	0 00159427
0_00700		5.2 Protein of unknown	2,107	1,070	0,00000070	0,00100427
		function similar to				
DECIDM13		proteins from other				
8 01260	Hypothetical protein	organisme	1 81	0.856	0 00204630	0 00607/31
0_01200		5.2 Protein of unknown	1,01	0,000	0,00204033	0,00037431
		function similar to				
		notoing from other				
8 01840	Hypothetical membrane protein	organisms	3 3 1 3	1 728	4 0837E-06	3 0707E-05
0_01040		5.2 Protoin of unknown	3,313	1,720	4,9037 E-00	3,97972-05
		5.2 FIOLEIII OF UNKNOWN				
		nunction similar to				
PFCIRIVI 13	Hypothetical protein	proteins from other	1 0 1	0.956	0 00027026	0 00050506
0_01925		5 2 Drotoin of unknown	1,01	0,000	0,00037930	0,02355555
		5.2 Protein of unknown				
		runction similar to				
PFCIRM13	Ly mothetical protein	proteins from other	2 251	4 000	0 10725 05	0 000 47796
0_01975		5 2 Dratain of unknown	2,301	1,233	9,1072E-05	0,00047786
		5.2 Protein of unknown				
		nunction similar to				
	Ly mothetical protain	proteins from other	1 1 5 1	0 1 5 4		1 10165 00
0_02050	nypothetical protein		4,451	2,154	5,8951E-10	1,1216E-08
PECIRM13		5.2 Protein of unknown	0.404	4.047	0.00000.400	0.0040500
8_03095	Hypothetical protein	function similar to	2,491	1,317	0,00038423	0,0016582

		proteins from other				
		5.2 Protein of unknown				
		function similar to				
PECIRM13		proteins from other				
8 03570	Hypothetical protein	organisms	11 8/17	3 566	1 365E-20	1 6502E-18
0_03370		5.2 Protein of unknown	11,047	5,500	1,3032-20	1,03321-10
		function similar to				
		protoing from other				
8 03720	Hypothetical protein	organisms	2 300	1 262	0.01883607	0.04588504
0_03720		5 2 Protoin of unknown	2,399	1,202	0,01003097	0,04300304
		5.2 FIOLEIN OF UNKNOWN				
		nunction similar to				
PFCIRIVI 13	Hypothetical protein	organisms	2 061	1 096	0.0020585	0 00079262
0_03075		5 2 Protoin of unknown	3,901	1,900	0,0030385	0,00978382
		5.2 FIOLEIII OF UNKNOWN				
		nunction similar to				
PFCIRIVI 13	Hypothetical protein	proteins from other	2 400	1 221	2 75065 05	0 00017009
0_04010		5 2 Drotoin of unknown	2,499	1,321	2,7590E-05	0,00017008
		5.2 Protein of unknown				
		runction similar to				
PFCIRM13	Ly mothetical protein	proteins from other	4 560	2.40	4 00565 11	0 62115 10
8_04055	Hypothetical protein	organisms	4,562	2,19	4,0056E-11	9,6311E-10
		5.2 Protein of unknown				
DEOIDIMA		function similar to				
PFCIRM13		proteins from other	0.500	0 700	0.0004.0400	0.00070504
8_04395	Hypothetical protein	organisms	6,539	2,709	0,00016438	0,00078531
		5.2 Protein of unknown				
DEOIDIAA		function similar to				
PFCIRM13		proteins from other	0.400	0.7		
8_04790	protein of unknown function	organisms	6,498	2,7	1,5613E-15	7,9444E-14
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other	1010	o oo-	0 4040 <del>5</del> 00	
8_05045	BioY family protein. Membrane protein	organisms	4,248	2,087	2,1616E-08	3,0317E-07
PFCIRM13		5.2 Protein of unknown				
8_05295	Hypothetical protein	function similar to	2,134	1,094	0,00083032	0,00318167

		proteins from other				
		5.2 Protein of unknown				
		function similar to				
PECIRM13		proteins from other				
8 06350	Hypothetical protein	organisms	1 977	0 983	0 00401692	0 01243144
0_00000		5.2 Protein of unknown	1,077	0,000	0,00401002	0,01240144
		function similar to				
PECIRM13		proteins from other				
8 06370	Hypothetical protein	organisms	1 736	0 796	0.0186894	0 04563884
0_00070		5.2 Protein of unknown	1,730	0,750	0,0100004	0,04000004
		function similar to				
DECIDM13		proteins from other				
8 06070	protein of unknown function	organisms	1 71	2 2/5	0 00620485	0 01822300
0_00370		5.2 Protein of unknown	4,74	2,243	0,00020403	0,01022303
		function similar to				
		notoing from other				
9 07000	protoin of unknown function	organisms	2 702	1 /2/	7 44045 05	0 00040245
8_07000		5 2 Drotoin of unknown	2,702	1,434	7,4494E-05	0,00040245
		5.2 Protein of unknown				
		runction similar to				
PFCIRM13	protoin of unknown function	proteins from other	0.400	4 777		4 20025 07
8_07025	protein of unknown function	organisms	3,428	1,777	9,4008E-09	1,3993E-07
		5.2 Protein of unknown				
DEOIDIAA		function similar to				
PFCIRM13		proteins from other	0.044	4 400	0 00044444	0 00050770
8_07065	protein of unknown function	organisms	2,644	1,403	0,00011444	0,00058776
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_07110	protein of unknown function	organisms	2,558	1,355	0,0001788	0,00084337
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_07195	protein of unknown function	organisms	15,105	3,917	1,1091E-05	7,9608E-05
PFCIRM13		5.2 Protein of unknown				
8_07680	Hypothetical membrane protein	function similar to	1,965	0,975	0,00015077	0,00073472

		proteins from other				
		5.2 Protoin of unknown				
		5.2 FIOLEIN OF UNKNOWN				
DECIDM13		proteins from other				
	Hypothetical protein		1 095	0 000	0 00022275	0 001 17225
6_07755		5 2 Drotoin of unknown	1,900	0,969	0,00033375	0,00147225
		5.2 Protein of unknown				
		runction similar to				
PFCIRM13		proteins from other	4 050	0.004	0.04000450	0.04400004
8_08135	cobait ABC transporter	organisms	4,859	2,281	0,01622152	0,04103201
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_08205	Hypothetical protein	organisms	1,835	0,876	0,00220236	0,00748255
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_08265	Hypothetical membrane protein	organisms	4,718	2,238	0,00757028	0,02162374
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_08425	Hypothetical membrane protein	organisms	5,262	2,396	0,00296868	0,00958036
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8 08510	protein of unknown function	organisms	2.793	1.482	0.00022247	0.00102691
		5.2 Protein of unknown	,	, -	- ,	
		function similar to				
PECIRM13		proteins from other				
8 08615	Hypothetical protein	organisms	4 077	2 028	2 6115E-08	3 6164E-07
0_00010		5.2 Protein of unknown	.,011	2,020	2,01102 00	0,01012 01
		function similar to				
PECIRM13		proteins from other				
8 08655	Hypothetical protein	organisms	25 004	4 644	7 6362E-06	5 7813E-05
PECIRM13		5.2 Protein of unknown	20,007		1,00021 00	5,7010E 00
8 08665	Hypothetical membrane protein	function similar to	11 764	3 556	0 0001404	0 00060187
0_00000	ן דיארט וויפוויט איז	TUTICIUM SITTIAL IO	11,704	3,000	0,0001404	0,00009107

		proteins from other organisms				
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8 08685	Hypothetical membrane protein	organisms	17,315	4,114	8,5125E-05	0,00045193
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_08690	Hypothetical protein	organisms	6,558	2,713	0,00303668	0,00972803
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_09090	Hypothetical membrane protein	organisms	2,448	1,292	2,116E-05	0,00013617
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_09425	Hypothetical membrane protein	organisms	9,849	3,3	0,0001539	0,0007483
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_09600	protein of unknown function	organisms	6,055	2,598	2,8939E-17	1,8623E-15
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_09605	protein of unknown function	organisms	2,469	1,304	2,0402E-05	0,00013246
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_10080	Hypothetical transmembrane protein	organisms	1,606	0,684	0,00811851	0,0229204
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_10150	Hypothetical protein	organisms	1,779	0,831	0,00957208	0,02624524
PFCIRM13		5.2 Protein of unknown				
8_10325	Hypothetical protein	function similar to	33,738	5,076	3,0234E-05	0,00018325

		proteins from other organisms				
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8 10340	protein of unknown function	organisms	8,002	3	0,0002369	0,0010776
		5.2 Protein of unknown				
		function similar to				
		proteins from other				
dedA	protein of unknown function	organisms	2,589	1,373	0,0190562	0,04627633
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_10635	Hypothetical protein	organisms	6,25	2,644	1,5135E-14	6,1324E-13
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_10675	Hypothetical protein	organisms	3,512	1,812	0,00010459	0,00054226
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_10820	Hypothetical membrane protein	organisms	19,805	4,308	2,7134E-05	0,00016771
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_11125	Hypothetical protein	organisms	2,24	1,163	2,328E-05	0,00014637
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_11265	Hypothetical protein	organisms	1,811	0,857	0,00369512	0,01151699
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8_11285	Hypothetical membrane protein	organisms	5,125	2,357	0,00362367	0,01131041
PFCIRM13		5.2 Protein of unknown				
8_11965	Hypothetical protein	function similar to	6,126	2,615	0,00077752	0,00298983

		proteins from other organisms				
		5.2 Protein of unknown				
		function similar to				
PFCIRM13		proteins from other				
8 12135	Hypothetical protein	organisms	30.564	4.934	1.7357E-06	1.5693E-05
		5.2 Protein of unknown	,	,	,	,
		function similar to				
PFCIRM13		proteins from other				
8 12190	membrane protein	organisms	1,905	0,93	0,00135085	0,00485328
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_00045	Hypothetical secreted protein	proteins	7,742	2,953	0,00137045	0,00490759
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_00340	Protein of unknown function	proteins	36,263	5,18	1,8097E-06	1,6295E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_00350	Protein of unknown function	proteins	18,263	4,191	1,9215E-05	0,00012625
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_00495	Hypothetical protein	proteins	2,522	1,335	6,3462E-06	4,9415E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_00540	Hypothetical protein	proteins	3,007	1,588	9,3287E-07	8,8745E-06
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_00685	Hypothetical protein	proteins	6,202	2,633	0,00040607	0,00173531
PFCIRM13		6 Protein of unknown				
8_00775	Hypothetical protein	function, without	1,796	0,845	0,02003396	0,04827567

		similarity to other				
		6 Protein of unknown				
		function without				
PECIRM13		similarity to other				
8 00860	Protein of unknown function	proteins	8,159	3.028	0.00100393	0.00374847
0_00000		6 Protein of unknown	0,100	0,020	0,00100000	0,0001 10 11
		function without				
PECIRM13		similarity to other				
8 01055	Hypothetical protein	proteins	3.515	1.813	9.0062E-06	6.7255E-05
0_01000		6 Protein of unknown	0,010	.,	0,00012 00	0,12002.00
		function, without				
PFCIRM13		similarity to other				
8 01100	Hypothetical protein	proteins	40.584	5.343	6.0332E-07	6.0277E-06
		6 Protein of unknown		_ ,	- )	-,
		function, without				
PFCIRM13		similarity to other				
8 01105	Hypothetical protein	proteins	3,755	1,909	1,0305E-05	7,5048E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_01110	Hypothetical protein	proteins	2,533	1,341	0,00034502	0,00151891
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_01255	Protein of unknown function	proteins	2,826	1,499	0,01473584	0,03797647
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_01415	Hypothetical protein	proteins	14,972	3,904	1,2887E-05	8,999E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_01860	Hypothetical protein	proteins	3,019	1,594	0,01039925	0,02812554
PFCIRM13		6 Protein of unknown				
8_01945	Hypothetical protein	function, without	5,918	2,565	0,0004425	0,0018652

		similarity to other				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 01985	Protein of unknown function	proteins	22,893	4,517	5,3057E-05	0,00029997
		6 Protein of unknown			,	
		function, without				
PFCIRM13		similarity to other				
8_02570	Hypothetical protein	proteins	2,069	1,049	0,00084363	0,00321579
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_02790	Hypothetical protein	proteins	7,426	2,893	2,7242E-20	3,1372E-18
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_02795	Protein of unknown function	proteins	6,412	2,681	0,0004137	0,00176018
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_02945	Protein of unknown function	proteins	6,159	2,623	0,00644455	0,01887641
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_03785	Hypothetical protein	proteins	8,007	3,001	0,00023102	0,00105967
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_03800	Hypothetical protein	proteins	9,542	3,254	0,00034867	0,0015319
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				<b>_</b>
8_03815	Protein of unknown function	proteins	8,461	3,081	2,3555E-17	1,718E-15
PFCIRM13		6 Protein of unknown				
8_03820	Hypothetical protein	function, without	46,981	5,554	4,7714E-31	2,088E-28

		similarity to other				
		6 Protein of unknown				
		function without				
PFCIRM13		similarity to other				
8 03825	Protein of unknown function	proteins	38,799	5.278	2.4245E-21	4.509E-19
		6 Protein of unknown		-, -	, -	,
		function, without				
PFCIRM13		similarity to other				
8 03845	Protein of unknown function	proteins	3,066	1,616	1,6862E-07	1,973E-06
		6 Protein of unknown			,	
		function, without				
PFCIRM13		similarity to other				
8_03915	Hypothetical protein	proteins	36,161	5,176	3,5589E-07	3,7437E-06
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_03920	Hypothetical protein	proteins	10	3,322	0,00042381	0,00179363
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_04020	Hypothetical protein	proteins	1,733	0,793	0,00990972	0,02700182
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_04075	Hypothetical protein	proteins	5,993	2,583	0,00011576	0,0005904
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_04470	Hypothetical protein	proteins	3,182	1,67	1,1097E-05	7,9608E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_04505	Hypothetical protein	proteins	7,362	2,88	0,00045722	0,0019128
PFCIRM13		6 Protein of unknown				
8_04700	Hypothetical protein	function, without	1,861	0,896	0,00303494	0,00972803

		similarity to other				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 04710	Hypothetical protein	proteins	2.875	1.524	2.1237E-05	0.00013626
		6 Protein of unknown	,	,	,	,
		function, without				
PFCIRM13		similarity to other				
8 04740	Hypothetical protein	proteins	15,023	3,909	1,6108E-07	1,8949E-06
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_04775	protein of unknown function	proteins	3,834	1,939	0,0032166	0,01021468
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_05060	Hypothetical protein	proteins	2,658	1,41	0,00327233	0,01036158
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_05135	Protein of unknown function	proteins	8,584	3,102	0,00029126	0,00129528
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_05225	Hypothetical protein	proteins	11,52	3,526	1,0332E-18	9,5632E-17
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_05240	Hypothetical protein	proteins	2,958	1,565	0,01699199	0,04248969
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_05605	Hypothetical protein	proteins	5,103	2,351	0,00026846	0,0012012
PFCIRM13		6 Protein of unknown				
8_05680	Hypothetical protein	function, without	3,232	1,693	0,00613998	0,01805681

		similarity to other				
		6 Protein of unknown				
		function without				
PFCIRM13		similarity to other				
8 06165	Hypothetical protein	proteins	5.277	2.4	6.5848E-09	1.0005E-07
		6 Protein of unknown	- /	,	-,	,
		function, without				
PFCIRM13		similarity to other				
8 06170	Protein of unknown function	proteins	7,138	2,836	0,00014478	0,00070711
		6 Protein of unknown		,		
		function, without				
PFCIRM13		similarity to other				
8_06315	Hypothetical protein	proteins	18,264	4,191	1,4142E-05	9,6699E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_06360	Protein of unknown function	proteins	2,088	1,062	0,00187852	0,00648298
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_06635	Hypothetical protein	proteins	1,534	0,618	0,01811254	0,04467896
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_06780	Protein of unknown function	proteins	29,47	4,881	5,3936E-06	4,2603E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_06785	Protein of unknown function	proteins	10,352	3,372	0,00023019	0,0010581
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_06790	Protein of unknown function	proteins	26,06	4,704	1,4786E-05	0,00010016
PFCIRM13		6 Protein of unknown				
8_06815	Hypothetical protein	function, without	2,101	1,071	0,0006017	0,0024112

		similarity to other				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 06820	Hypothetical protein	proteins	8.02	3.004	1.8565E-16	1.0689E-14
		6 Protein of unknown	- , -	- ,	,	,
		function, without				
PFCIRM13		similarity to other				
8 06875	Hypothetical protein	proteins	2,551	1,351	0,00132553	0,00479382
		6 Protein of unknown	,	,	,	,
		function, without				
PFCIRM13		similarity to other				
8 06955	Hypothetical protein	proteins	5,932	2,568	0,00212844	0,00724264
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_06990	protein of unknown function	proteins	4,812	2,267	8,3428E-08	9,9748E-07
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_07105	Hypothetical protein	proteins	4,688	2,229	3,5802E-13	1,224E-11
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_07155	Protein of unknown function	proteins	6,541	2,71	0,00073586	0,00286486
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_07160	protein of unknown function	proteins	18,185	4,185	3,7496E-05	0,00021995
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_07190	protein of unknown function	proteins	10,206	3,351	0,0001789	0,00084337
PFCIRM13		6 Protein of unknown				
8_07200	protein of unknown function	function, without	9,477	3,244	0,00016107	0,00077454

		similarity to other proteins				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 07255	Protein of unknown function	proteins	17,52	4,131	2,5936E-05	0,00016168
		6 Protein of unknown	,			
		function, without				
PFCIRM13		similarity to other				
8 07290	Protein of unknown function	proteins	14,434	3,851	0,00051761	0,00211294
		6 Protein of unknown	,	,		,
		function, without				
PFCIRM13		similarity to other				
8 07475	Protein of unknown function	proteins	4,931	2,302	0,0041776	0,01285597
		6 Protein of unknown	,	,	,	,
		function, without				
PFCIRM13		similarity to other				
8 07580	protein of unknown function	proteins	7,856	2,974	3,1621E-08	4,2708E-07
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 07625	Protein of unknown function	proteins	14,139	3,822	1,3418E-05	9,2323E-05
		6 Protein of unknown	,	,	,	,
		function, without				
PFCIRM13		similarity to other				
8_07895	Hypothetical protein	proteins	2,517	1,332	0,00099312	0,00371643
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_07905	protein of unknown function	proteins	8,36	3,064	3,7729E-07	3,9498E-06
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_08415	Hypothetical protein	proteins	17,829	4,156	0,00020056	0,00093766
PFCIRM13		6 Protein of unknown		-		
8_08440	Hypothetical protein	function, without	11,537	3,528	4,2507E-06	3,4834E-05

		similarity to other				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 08445	Protein of unknown function	proteins	35.068	5.132	2.2273E-07	2.4613E-06
		6 Protein of unknown		-, -	,	,
		function, without				
PFCIRM13		similarity to other				
8 08450	Protein of unknown function	proteins	5,264	2,396	0,00290053	0,00945808
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 08455	Protein of unknown function	proteins	15,732	3,976	7,589E-05	0,00040698
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_08535	Hypothetical protein	proteins	6,391	2,676	0,00179745	0,00626246
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_08575	Hypothetical protein	proteins	4,394	2,136	1,0378E-14	4,4524E-13
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_08580	Hypothetical protein	proteins	4,685	2,228	2,066E-06	1,8375E-05
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_08585	Hypothetical protein	proteins	4,993	2,32	0,00048621	0,00201103
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_08680	Hypothetical protein	proteins	5,209	2,381	0,00161249	0,00568136
PFCIRM13		6 Protein of unknown				
8_08975	Hypothetical protein	function, without	2,048	1,034	0,01846948	0,04530406

		similarity to other				
		6 Proteins				
		function without				
PECIRM13		similarity to other				
8 09120	Hypothetical protein	proteins	6 3 1 9	2 66	2 2515E-13	8 2105E-12
0_03120		6 Protein of unknown	0,010	2,00	2,20102 10	0,21002 12
		function without				
PECIRM13		similarity to other				
8 09125	Protein of unknown function	proteins	10 349	3 371	1 5412E-17	1 1628E-15
0_00120		6 Protein of unknown	10,040	0,071	1,04126 17	1,10202 10
		function without				
PECIRM13		similarity to other				
8 09230	Hypothetical protein	proteins	12 115	3 599	2 8213E-06	2 4208E-05
0_00200		6 Protein of unknown	12,110	0,000	2,02102 00	2,12002 00
		function without				
PECIRM13		similarity to other				
8 09235	Hypothetical protein	proteins	12.44	3.637	1.5483E-12	5.0407E-11
0_00200		6 Protein of unknown	,	0,001	.,0:002 .2	0,0101 = 11
		function without				
PFCIRM13		similarity to other				
8 09340	Hypothetical protein	proteins	9.116	3.188	0.00049039	0.00202096
		6 Protein of unknown		-,		
		function, without				
PFCIRM13		similarity to other				
8 09610	Protein of unknown function	proteins	20,668	4,369	5,2006E-07	5,2437E-06
		6 Protein of unknown		,	,	
		function, without				
PFCIRM13		similarity to other				
8_09655	Protein of unknown function	proteins	7,342	2,876	0,00023686	0,0010776
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_09660	Hypothetical protein	proteins	3,091	1,628	3,761E-05	0,00022003
PFCIRM13		6 Protein of unknown				
8_09675	Hypothetical protein	function, without	3,674	1,877	0,00295749	0,00958036

		similarity to other				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_09695	Protein of unknown function	proteins	5,343	2,418	0,00257269	0,00859397
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_09700	protein of unknown function	proteins	28,614	4,839	4,0353E-08	5,2555E-07
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_10335	protein of unknown function	proteins	4,065	2,023	0,00876188	0,02439059
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_10470	Hypothetical protein	proteins	2,212	1,145	7,5248E-05	0,00040552
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				0 000 17505
8_10730	Hypothetical secreted protein	proteins	7,306	2,869	9,032E-05	0,00047505
		6 Protein of unknown				
DEOIDIAA		function, without				
PFCIRM13		similarity to other	4 050	0.040		
8_10740	Hypothetical secreted protein	proteins	4,653	2,218	5,8076E-08	7,2612E-07
		6 Protein of unknown				
DEOIDIMA		function, without				
PFCIRM13		similarity to other	0.000	4 405	0.01000001	0.00704040
8_10825	Hypothetical protein	proteins	2,289	1,195	0,0100081	0,02721613
		6 Protein of unknown				
		iunction, without				
9 10045	Hypothetical protein		1 5 9 7	0.666	0.01200227	0 02605729
0_10940 DECIDM40		6 Protoin of unknown	1,007	0,000	0,01309227	0,03003728
	Ly mothetical protain	6 FIOLEIN OF UNKNOWN	E 204	2 404	0.00044004	0.00000050
0_11080	nypolnelical protein	runction, without	5,∠ŏ1	∠,401	0,00244981	0,00820856

		similarity to other proteins				
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 11150	Hypothetical protein	proteins	1,785	0,836	0,01560696	0,039707
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 11315	Hypothetical protein	proteins	3,456	1,789	0,00118544	0,00433735
		6 Protein of unknown	,	,	,	,
		function, without				
PFCIRM13		similarity to other				
8 11410	Hypothetical membrane protein	proteins	7,12	2,832	0,00039519	0,00169879
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 11585	Hypothetical protein	proteins	5,681	2,506	0,00231847	0,00781634
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 11770	Protein of unknown function	proteins	3,746	1,905	0,00488555	0,01476462
		6 Protein of unknown	,	,	,	,
		function, without				
PFCIRM13		similarity to other				
8 11920	Protein of unknown function	proteins	3,976	1,991	0,01790837	0,04427515
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8 12160	Hypothetical protein	proteins	4,339	2,117	0,00289119	0,00945188
		6 Protein of unknown				
		function, without				
PFCIRM13		similarity to other				
8_12320	Protein of unknown function	proteins	2,612	1,385	0,01647353	0,0416199
PFCIRM13		6.2 Protein of unknown			-	•
8_07965	protein of unknown function	function similar to	2,414	1,271	0,00021458	0,0009968

		proteins from other organisms				
PFCIRM13		7.2 Protein of unknown function similar to proteins from other				
8_07250	protein of unknown function	organisms	2,252	1,171	0,01141197	0,03048767
PFCIRM13		8.2 Protein of unknown function similar to proteins from other				
8_08485	protein of unknown function	organisms	2,193	1,133	0,00017041	0,00081055
PFCIRM13		9.2 Protein of unknown function similar to proteins from other				
8_07140	protein of unknown function	organisms	2,139	1,097	0,01049979	0,0283624
**Table S2**: Repressed genes in stationnary phase compared to exponential phase.

					log2Fold		
Name	Descript	Function	FC	1/FC	Change	pvalue	padj
	dTDP-4-dehydrorhamnose reductase / dTDP-4-	1 Cell envelope and					
rmIC (cps)	dehydrorhamnose 3,5-epimerase	cellular processes	0,349	2,86532951	-1,52	3,4728E-10	7,0388E-09
glmU	UDP-N-acetylglucosamine pyrophosphorylase	1.1 Cell wall	0,63	1,58730159	-0,667	0,01478679	0,03806294
	UDP-N-acetylmuramateL-alanine ligase (UDP-N-						
murC	acetylmuramoyl-L-alanine synthetase)	1.1 Cell wall	0,425	2,35294118	-1,234	2,2735E-06	1,9898E-05
	UDP-N-acetylmuramoylalanineD-glutamate ligase						
	(UDP-N- acetylmuramoyl-L-alanyl-D-glutamate						
murD	synthetase) (D-glutamic acid- adding enzyme)	1.1 Cell wall	0,596	1,67785235	-0,746	0,00654238	0,01908631
	Phospho-N-acetylmuramoyl-pentapeptide-transferase						
mraY	(UDP- MurNAc-pentapeptide phosphotransferase)	1.1 Cell wall	0,291	3,43642612	-1,783	2,9208E-10	6,145E-09
	UDP-N-acetylmuramoyl-tripeptideD-alanyl-D-alanine						
	ligase (UDP- MurNAc-pentapeptide synthetase) (D-						
murF	alanyl-D-alanine-adding enzyme)	1.1 Cell wall	0,507	1,97238659	-0,979	0,00069815	0,00273757
	UDP-N-acetylmuramoylalanyl-D-glutamate2,6-						
	diaminopimelate ligase (UDP-N-acetylmuramyl-tripeptide						
_	synthetase) (Meso- diaminopimelate-adding enzyme)						
murE	(UDP-MurNAc-tripeptide synthetase)	1.1 Cell wall	0,527	1,89753321	-0,924	0,0006803	0,00268196
	Cell division protein FtsI (penicillin-binding protein 2)						
ftsl	(Peptidoglycan glycosyltransferase)	1.1 Cell wall	0,515	1,94174757	-0,957	0,00032724	0,00144646
PFCIRM1							
38_02375	cell-wall peptidases, NIpC/P60 family secreted protein	1.1 Cell wall	0,192	5,20833333	-2,384	3,529E-18	2,8598E-16
ddIA	D-alanineD-alanine ligase (D-alanylalanine synthetase)	1.1 Cell wall	0,336	2,97619048	-1,576	2,9725E-10	6,1942E-09
dpm	dolichyl-phosphate beta-D-mannosyltransferase	1.1 Cell wall	0,367	2,72479564	-1,448	2,8712E-05	0,00017597
	UDP-N-acetylglucosamine 1-carboxyvinyltransferase						
MurA	(Enoylpyruvate transferase)	1.1 Cell wall	0,555	1,8018018	-0,849	0,00063216	0,00251028
PFCIRM1							
38_03660	Rhamnosyltransferase, Glycosyl transferase family 2	1.1 Cell wall	0,597	1,67504188	-0,744	0,00348714	0,01091539
pbpA	Penicillin-binding protein	1.1 Cell wall	0,318	3,14465409	-1,654	1,8129E-07	2,0987E-06
mrc/ponA	penicillin-binding protein (peptidoglycan	1.1 Cell wall	0,645	1,5503876	-0,632	0,01277471	0,03363546

	glycosyltransferase)						
slpG	Surface layer protein G	1.1 Cell wall	0,355	2,81690141	-1,494	1,0062E-08	1,4876E-07
	UDP-N-acetylenolpyruvoylglucosamine reductase (UDP-						
murB	N- acetylmuramate dehydrogenase)	1.1 Cell wall	0,551	1,81488203	-0,859	0,00189094	0,00650532
slpF	surface layer protein F (S-layer protein F)	1.1 Cell wall	0,447	2,23713647	-1,162	1,3114E-05	9,1088E-05
gtfB	Glycosyltransferase	1.1 Cell wall	0,577	1,73310225	-0,794	0,00456797	0,01393965
tagG2	ABC-type polysaccharide export systems	1.1 Cell wall	0,379	2,63852243	-1,401	3,5163E-06	2,9253E-05
	ABC-type polysaccharide export systems, permease						
tagG1	component	1.1 Cell wall	0,518	1,93050193	-0,948	0,00012993	0,00065053
gtfA	Glycosyltransferase, family 2	1.1 Cell wall	0,344	2,90697674	-1,539	4,5625E-08	5,907E-07
glf, rfb	UDP-galactopyranose mutase	1.1 Cell wall	0,472	2,11864407	-1,083	0,00054009	0,00218836
_	Undecaprenyl-phosphate alpha-N-						
tagO	acetylglucosaminyltransferase	1.1 Cell wall	0,539	1,85528757	-0,89	0,00115377	0,00425707
		1.2 Transport/binding					
PECIRM1		proteins and					
38 00755	Hypoxanthine/guanine permease 2.A.40.7.2	lipoproteins	0,521	1,9193858	-0,94	0.02000989	0,04827567
		1.2		,	- , -	-,	- ,
		Transport/binding					
PFCIRM1		proteins and					
38_00850	Transporter, major facilitator family 2.A.1.52.3	lipoproteins	0,181	5,52486188	-2,469	4,6211E-15	2,1513E-13
		1.2 Transport/binding					
PFCIRM1		proteins and					
38 01645	ATP-binding protein ABC transporter	lipoproteins	0,366	2,73224044	-1,451	0,00309331	0,00986612
		1.2	,	,	,		,
		Transport/binding					
PFCIRM1		proteins and					
38_02940	transporter	lipoproteins	0,34	2,94117647	-1,557	5,2582E-09	8,2179E-08
		1.2 Transport/binding					
PFCIRM1	BP protein of monosaccharide ABC transporter	proteins and					
38_03110	(MOS:undef)	lipoproteins	0,325	3,07692308	-1,623	1,2191E-05	8,6046E-05
PFCIRM1	Alpha-ketoglutarate permease 2.A.1.6.2	1.2	0,399	2,50626566	-1,326	0,0029664	0,00958036

38_03760		Transport/binding proteins and lipoproteins					
PFCIRM1 38_04250	ABC transporter permease protein	1.2 Transport/binding proteins and lipoproteins	0,496	2,01612903	-1,011	0,00665541	0,01928747
PFCIRM1 38_04695	Xanthine/uracil permease	1.2 Transport/binding proteins and lipoproteins	0,489	2,04498978	-1,031	0,00875932	0,02439059
PFCIRM1 38_04945	Redox protein export	1.2 Transport/binding proteins and lipoproteins	0,62	1,61290323	-0,69	0,01649198	0,0416199
PFCIRM1 38_05530	ATP-binding protein of ABC transporter	1.2 Transport/binding proteins and lipoproteins	0,462	2,16450216	-1,113	4,8661E-05	0,00027872
PFCIRM1 38 05955	ABC transporter ATP-binding protein	1.2 Transport/binding proteins and lipoproteins	0,499	2,00400802	-1,002	0,00328542	0.010388
PFCIRM1 38 07330	polar amino acid ABC transporter, binding protein component	1.2 Transport/binding proteins and lipoproteins	0.45	2,22222222	-1,153	0,00013505	0.00066855
oppD	ABC2 protein of oligopeptide ABC transporter	1.2 Transport/binding proteins and lipoproteins	0,283	3,5335689	-1,82	1,2652E-12	4,1943E-11
annC	IM protain of aligonantida ARC transporter (ODN/updat)	1.2 Transport/binding proteins and	0.200	2 24449161	1 740	4 45275 12	1 26555 10
оррВ	IM protein of oligopeptide ABC transporter (OPN:undef)	1.2	0,299	2,79329609	-1,742	2,4867E-08	3,4656E-07

		Transport/binding					
		lipoproteins					
		1.2					
		Transport/binding					
PFCIRM1		proteins and					
38_09060	Permease component of cobalt ABC transporter	lipoproteins	0,482	2,0746888	-1,054	0,01199629	0,03181561
		1.2					
	PD protoin of motallia action (7n2 + Mn2 +) APC	I ransport/binding					
	bP protein of metallic cation (212+ M12+) ABC	lipoprotoine	0 205	2 52164557	1 2/1	0.00015926	0.00076272
36_09645			0,395	2,55104557	-1,341	0,00015820	0,00070273
		Transport/binding					
PFCIRM1		proteins and					
38 10105	Preprotein translocase SecA subunit	lipoproteins	0,5	2	-1,001	0,00168797	0,00592823
		1.2					
		Transport/binding					
PFCIRM1		proteins and					
38_11820	ABC2 protein of ABC transporter (ART:REG)	lipoproteins	0,567	1,76366843	-0,819	0,00148618	0,00527884
		1.2.1					
1.10	Sec-independent protein translocase protein TatC	Transport/binding of	0.400	0 4 4 5 0 0 0 7 5	4 4 9 9		0 00040454
tatC	nomolog	proteins/peptides	0,466	2,14592275	-1,103	1,5475E-05	0,00010451
		T.Z.1					
htr∆2	secreted serine protesse, trypsin-like serine protesses	nroteins/pentides	0 4 1 4	2 41545894	-1 273	0 00044328	0.0018652
		121	0,414	2,41040004	1,275	0,00044020	0,0010002
		Transport/binding of					
bopA	solute binding protein of the ABC transport system	proteins/peptides	0,471	2,12314225	-1,087	0,00053912	0,00218836
•		1.2.1					
PFCIRM1		Transport/binding of					
38_11405	ATP-binding component ABC transporter	proteins/peptides	0,572	1,74825175	-0,805	0,01324727	0,03458939
		1.2.3					
		Transport/binding of					
teoB	⊢errous iron uptake protein B 9.A.8.1.x	inorganic ions	0,367	2,72479564	-1,445	0,00036855	0,00159997
PFCIRM1		1.2.3	0.000	4 57000704	0.050	0.00007004	0.00000000
38_02880	TIKA, potassium transporter	Transport/binding of	0,636	1,5/232/04	-0,653	0,00697931	0,02006666

		inorganic ions					
		1.2.3					
		Transport/binding of					
mntH	Mn2+ and Fe2+ transporter	inorganic ions	0,139	7,1942446	-2,852	6,1578E-12	1,6842E-10
		1.2.3					
		Transport/binding of					
cbiO1	Cobalt import ATP-binding protein CbiO	inorganic ions	0,32	3,125	-1,644	0,00088694	0,00335749
		1.2.3					
		Transport/binding of					
cbiQ2	Cobalt transport protein CbiQ	inorganic ions	0,306	3,26797386	-1,707	0,00039985	0,00171335
		1.2.3					
		Transport/binding of					
cbiN	Cobalt transport protein CbiN	inorganic ions	0,266	3,7593985	-1,912	9,1322E-11	2,0389E-09
		1.2.3					
		Transport/binding of					
cbiM	Cobalt transport protein CbiM	inorganic ions	0,285	3,50877193	-1,811	2,6279E-09	4,3893E-08
		1.2.3					
PFCIRM1		Transport/binding of					
38_11535	Cation transport protein	inorganic ions	0,48	2,08333333	-1,06	0,00686414	0,01981363
		1.2.4					
		Transport/binding of					
glpF	Glycerol uptake facilitator protein,	carbohydrates	0,12	8,33333333	-3,056	3,9947E-12	1,1811E-10
		1.2.4					
		Transport/binding of					
galP	Sodium:galactoside symporter	carbohydrates	0,403	2,48138958	-1,311	0,00042013	0,0017815
		1.2.4					
PFCIRM1		Transport/binding of					
38_02775	sugar transporter, major facilitator superfamily	carbohydrates	0,286	3,4965035	-1,806	0,00280826	0,00922833
		1.2.4					
PFCIRM1	IM protein of monosaccharide ABC transporter	Transport/binding of	0.040	0.00500054	4 50	0.00400000	
38_03105	(MOS:undef)	carbohydrates	0,349	2,86532951	-1,52	0,00189062	0,00650532
		1.2.4					
		I ransport/binding of	0.057	0.00405050	4 050		4 57075 00
וומצ	L-lactate permease	carbonydrates	0,257	3,89105058	-1,959	8,415E-10	1,5/3/E-08
		1.2.4	0.000	0 0000 4505	4 000		
CSDX	Alpha-ketoglutarate permease	I ransport/binding of	0,309	3,23624595	-1,696	1,2403E-09	2,171E-08

		carbohydrates					
		1.2.4					
	Sodium/glucose cotransporter (Na(+)/glucose symporter)	Transport/binding of					
slgT	2.A.21.3.2	carbohydrates	0,247	4,048583	-2,016	2,1526E-05	0,00013771
		1.2.4					
PFCIRM1	PTS system, Lactose/Cellobiose or galactitol specific IIB	Transport/binding of					
38_09590	subunit	carbohydrates	0,475	2,10526316	-1,075	0,01727657	0,04300471
		1.2.5					
	ATP-binding protein of glutamate ABC transporter (PAO :	Transport/binding of					
gluA	undef : Glutamate)	amino-acids	0,603	1,65837479	-0,729	0,01697461	0,04248969
		1.2.5					
PFCIRM1		Transport/binding of					
38_05630	Oligopeptide transporter, OPT family protein.	amino-acids	0,529	1,89035917	-0,918	0,00026243	0,00118634
		1.2.5					
PFCIRM1		Transport/binding of					
38_05890	Amino acid permease. membrane protein	amino-acids	0,425	2,35294118	-1,235	1,715E-05	0,00011405
		1.2.5					
PFCIRM1		Transport/binding of					
38_06700	ABC-type transport systems, periplasmic component	amino-acids	0,189	5,29100529	-2,401	2,252E-06	1,9791E-05
		1.2.5					
		Transport/binding of					
metN	Methionine import ATP-binding protein metN	amino-acids	0,189	5,29100529	-2,405	1,6269E-11	4,0915E-10
		1.2.5					
PFCIRM1		Transport/binding of					
38_06710	ABC transporter, permease protein	amino-acids	0,19	5,26315789	-2,397	2,9778E-14	1,143E-12
		1.2.5					
PFCIRM1	polar amino acid ABC transporter, inner membrane	Transport/binding of					
38_07320	component	amino-acids	0,289	3,46020761	-1,79	3,7466E-12	1,123E-10
		1.2.5					
PFCIRM1		Transport/binding of					
38_07325	polar amino acid ABC transporter, binding protein	amino-acids	0,482	2,0746888	-1,053	1,1802E-05	8,3568E-05
		1.2.5					
PFCIRM1	binding protein of oligopeptide ABC transporter (OPN :	I ransport/binding of					
38_10275	undet : Oligopeptides)	amino-acids	0,341	2,93255132	-1,55	8,149E-08	9,7967E-07
PFCIRM1		1.2.5					
38_10280	permease protein of oligopeptide ABC transporter	Transport/binding of	0,538	1,85873606	-0,895	0,01001421	0,02721876

		amino-acids					
		1.2.5					
		Transport/binding of					
argK	Kinase ArgK	amino-acids	0,36	2,77777778	-1,473	6,4451E-05	0,00035521
		1.2.6					
		Transport/binding of					
		nucleosides,					
		nucleotides, purines					
carA	Carbamoyl-phosphate synthase small chain	and pyrimidines	0,605	1,65289256	-0,726	0,00517053	0,01554
		1.2.6					
		Transport/binding of					
		nucleosides,					
		nucleotides, purines					
codB	Permease for cytosine/purines, uracil, thiamine, allantoin	and pyrimidines	0,365	2,73972603	-1,452	0,00053094	0,00216329
		1.2.6					
		Transport/binding of					
		nucleosides,					
	Adenylate kinase (EC 2.7.4.3) (ATP-AMP	nucleotides, purines					
adk	transphosphorylase)	and pyrimidines	0,358	2,79329609	-1,482	8,9583E-06	6,7126E-05
PFCIRM1		1.3 Sensors (signal					
38_03945	Forkhead-associated protein	transduction)	0,634	1,57728707	-0,658	0,01006328	0,02731819
PFCIRM1		1.3 Sensors (signal					
38_03950	Forkhead-associated protein	transduction)	0,375	2,66666667	-1,416	4,6256E-05	0,00026704
PFCIRM1	Sensor protein, ATP-binding region, ATPase-	1.3 Sensors (signal					
38_06685	like:Histidine kinase	transduction)	0,492	2,03252033	-1,023	0,00116254	0,00428222
PFCIRM1		1.3 Sensors (signal					
38_09785	Sensor protein, ATPase-like:Histidine kinase	transduction)	0,321	3,1152648	-1,638	1,1946E-10	2,6138E-09
PFCIRM1		1.3 Sensors (signal					
38_09790	two-component response regulator	transduction)	0,421	2,37529691	-1,249	4,3636E-06	3,5493E-05
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP					
sdhB3	Succinate dehydrogenase	synthase)	0,172	5,81395349	-2, <u>53</u> 9	9,6739E-11	2,138E-09
		1.4 Membrane					
sdhA3	Succinate dehydrogenase flavoprotein subunit	bioenergetics	0,218	4,58715596	-2,195	2,5141E-11	6,1121E-10

		(electron transport chain and ATP					
		Synthase)					
		hippenergetics					
		(electron transport					
		chain and ATP					
sdhC2	Succinate dehydrogenase cytochrome B-558 subunit	synthase)	0 227	4 40528634	-2 136	2 2604E-12	6 966E-11
501102		1 4 Membrane	0,221	4,40020004	2,100	2,20046 12	0,0002 11
		bioenergetics					
		(electron transport					
		chain and ATP					
mgtE	magnesium (Mg2+) transporter	synthase)	0,615	1,62601626	-0,702	0,01194171	0.0317093
0		1.4 Membrane	,	,	,	,	,
		bioenergetics					
		(electron transport					
		chain and ATP					
fdxA	Ferredoxin	synthase)	0,35	2,85714286	-1,516	7,4299E-08	8,9815E-07
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP					
nuoN	NADH dehydrogenase I chain N	synthase)	0,165	6,06060606	-2,595	2,7031E-13	9,3881E-12
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP					
nuoM	NADH dehydrogenase I chain M	synthase)	0,157	6,36942675	-2,67	1,707E-12	5,3858E-11
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP					
nuoL	NADH dehydrogenase (quinone)	synthase)	0,201	4,97512438	-2,314	2,59E-09	4,3592E-08
		1.4 Membrane					
		bioenergetics					
nuoK	NADH dehydrogenase I chain K	(electron transport	0,199	5,02512563	-2,329	1,5742E-11	4,005E-10

		chain and ATP					
		1 4 Membrane					
		bioenergetics					
		(electron transport					
	NADH-quinone oxidoreductase chain J (NADH	chain and ATP					
nuoJ	dehydrogenase I, chain J)	synthase)	0,237	4,21940928	-2,075	3,6112E-08	4,7886E-07
		1.4 Membrane			,		
		bioenergetics					
		(electron transport					
	NADH-quinone oxidoreductase subunit I (NADH	chain and ATP					
nuol	dehydrogenase I subunit I) (NDH-1 subunit I)	synthase)	0,246	4,06504065	-2,023	1,2231E-09	2,1583E-08
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	NADH-quinone oxidoreductase subunit H (NADH	chain and ATP					
nuoH	dehydrogenase I subunit H)	synthase)	0,314	3,18471338	-1,673	1,2256E-06	1,1509E-05
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	NADH-quinone oxidoreductase chain G (NADH	chain and ATP					
nuoG	dehydrogenase I, chain G)	synthase)	0,288	3,47222222	-1,795	5,0931E-06	4,0523E-05
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	NADH-quinone oxidoreductase chain F (NADH	chain and ATP					
nuoF	dehydrogenase I, chain F) (NDH-1, chain F)	synthase)	0,196	5,10204082	-2,354	1,2923E-08	1,8726E-07
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP					
nuoE	NADH-quinone oxidoreductase chain E	synthase)	0,217	4,60829493	-2,206	2,5645E-12	7,7934E-11
		1.4 Membrane					
		bioenergetics					
	NADH-quinone oxidoreductase chain D (EC 1.6.99.5)	(electron transport					
nuoD	(NADH dehydrogenase I, chain D)	chain and ATP	0,164	6,09756098	-2,607	4,9573E-12	1,3906E-10

		synthase)					
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	NADH-quinone oxidoreductase chain C (NADH	chain and ATP					
nuoC	dehydrogenase I, chain C)	synthase)	0,171	5,84795322	-2,545	6,6857E-15	2,9854E-13
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	NADIL suisses autidas dustas adais D	chain and ATP	0.004	0 70707070	4 000		4 04005 40
NUOB	NADH-quinone oxidoreductase chain B	synthase)	0,264	3,78787879	-1,922	5,9342E-12	1,6436E-10
		1.4 Membrane					
		bioenergetics					
ημοΑ	NADH-quipope oxidoreductase chain	synthase)	0 206	4 85436893	-2 276	3 8733E-15	1 8423E-13
1100/1		1 4 Membrane	0,200	4,00400000	2,210	0,07002 10	1,04202 10
		bioenergetics					
		(electron transport					
PFCIRM1		chain and ATP					
38 05515	electron transfer oxidoreductase	synthase)	0,332	3,01204819	-1,589	3,2108E-08	4,31E-07
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP					
dhbC	Menaquinone-specific isochorismate synthase	synthase)	0,549	1,82149362	-0,865	0,00276458	0,00912165
		1.4 Membrane					
		bioenergetics					
		(electron transport					
PFCIRM1		chain and ATP					
38_05930	iron-sulfur protein	synthase)	0,168	5,95238095	-2,571	2,6088E-13	9,2066E-12
		1.4 Membrane					
		bioenergetics					
Arth 1111	EAD linked evidese domain protein	chain and ATP	0.284	2 52112676	1 915	5 2072E 07	5 2166E 06
AIUI_4141	FAD iinkeu uxiuase uumain protein	synunase)	0,∠04	3,32112070	-1,010	5,29120-01	5,3100 <b>⊑-</b> 00

		1.4 Membrane bioenergetics					
		(electron transport					
adhC1	Sussingto debudrogenego subunit C	chain and ATP	0.116	0.0000000	2 4 0 7		
SUNCT		Synthase)	0,116	0,02000900	-3,107	4,209E-21	0,0719E-19
		hippenergetics					
		(electron transport					
		chain and ATP					
sdhA	Succinate dehydrogenase, subunit A	synthase)	0,113	8,84955752	-3,147	1,8194E-18	1,5923E-16
		1.4 Membrane			-		
		bioenergetics					
		(electron transport					
		chain and ATP					
sdhB	Succinate dehydrogenase, subunit B	synthase)	0,115	8,69565217	-3,114	5,1373E-16	2,8101E-14
		1.4 Membrane					
		bioenergetics					
		(electron transport					
		chain and ATP	0 477	0.00040000	1 000	0.0004.04.47	0.00004.000
MOXR	A Pases MoxR family	synthase)	0,477	2,09643606	-1,069	0,00012147	0,00061238
		1.4 Memorane					
		bioenergetics					
atoF	ATP synthase B chain (E0E1 ATP synthase subunit B)	synthase)	0 554	1 80505415	-0.852	0.00200057	0 00682877
aipi		1 4 Membrane	0,004	1,00000410	0,002	0,00200007	0,00002011
		bioenergetics					
		(electron transport					
		chain and ATP					
atpH	ATP synthase delta chain	synthase)	0,509	1,96463654	-0,975	0,0001822	0,00085548
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	ATP synthase subunit alpha (ATPase subunit alpha)	chain and ATP					
atpA	(ATP synthase F1 sector subunit alpha)	synthase)	0,412	2,42718447	-1,279	2,3908E-05	0,00014989
atpG	ATP synthase gamma chain (ATP synthase F1 sector	1.4 Membrane	0,41	2,43902439	-1,287	1,1699E-05	8,311E-05

	gamma subunit)	bioenergetics (electron transport					
		chain and ATP					
		synthase)					
		1.4 Membrane					
		bioenergetics					
		(electron transport					
otoD	ATP synthase subunit beta (ATPase subunit beta) (ATP	chain and ATP	0.42	2 2255011	1 210	0.00046280	0 00102701
aipD	Synthase FT Sector Suburnit Deta)	1 / Membrane	0,43	2,3233014	-1,219	0,00040369	0,00193701
		bioenergetics					
		(electron transport					
	ATP synthase epsilon chain (ATP synthase F1 sector	chain and ATP					
atpC	epsilon subunit)	synthase)	0,477	2,09643606	-1,068	7,1146E-05	0,00038627
		1.4 Membrane					
		bioenergetics					
		(electron transport					
	Dummente eurotheses (Dummente fleure destine eurothese due teses	chain and ATP	0.074	0.0000000	4 005		
nitj'i	Pyruvate synthase/Pyruvate-flavodoxin oxidoreductase	synthase)	0,271	3,6900369	-1,885	6,8721E-07	6,6828E-06
		hipenergetics					
	FAD-dependent pyridine nucleotide-disulphide	(electron transport					
PFCIRM1	oxidoreductase:4Fe-4S ferredoxin, iron-sulfur	chain and ATP					
38_10760	binding:Aromatic-ring hydroxylase	synthase)	0,293	3,41296928	-1,77	4,7968E-08	6,1737E-07
		1.4 Membrane					
		bioenergetics					
		(electron transport					
PFCIRM1		chain and ATP	0 500	4 7000007	0 700	0.04000004	0.04550007
38_12020	Fe-S oxidoreductases family 2	synthase)	0,588	1,70068027	-0,766	0,01863881	0,04558667
totB	sec-independent protein translocase protein Tate	1.6 Protoin cocrotion	0.490	2 04 409079	1 021	0.0029112	0 01194502
		1.0 Protein secretion	0,409	2,04490970	-1,031	0,0030112	0,01104302
yajc		1.6 Protein secretion	0,608	1,64473684	-0,717	0,0077198	0,02193626
secD	Protein-export membrane protein secD	1.6 Protein secretion	0,397	2,51889169	-1,333	4,5803E-07	4,705E-06
secF	Protein-export membrane protein secF	1.6 Protein secretion	0,196	5,10204082	-2,35	4,9262E-17	2,994E-15
secY	Preprotein translocase SecY subunit	1.6 Protein secretion	0,413	2,42130751	-1,275	2,778E-07	2,965E-06

ftsQ	Cell division protein FtsQ	1.7 Cell division	0,375	2,66666667	-1,415	0,00013391	0,00066741
PFCIRM1							
38_04995	ATPase involved in cell division	1.7 Cell division	0,479	2,08768267	-1,061	2,4266E-05	0,0001517
	Aldehyde dehydrogenase (Succinate-semialdehyde	2.1 Metabolism of					
	dehydrogenase) (NAD-dependent aldehyde	carbohydrates and					
gab	dehydrogenase)	related molecules	0,529	1,89035917	-0,919	0,00593897	0,01753638
0		2.1 Metabolism of					,
		carbohvdrates and					
pfp (pfk)	pyrophosphate phosphofructokinase	related molecules	0,318	3,14465409	-1,652	2,173E-07	2,4253E-06
	Dihydrolipoyl dehydrogenase (E3 component of alpha	2.1 Metabolism of					
	keto acid dehydrogenase complexes) (Dihydrolipoamide	carbohydrates and					
lpd	dehydrogenase)	related molecules	0,504	1,98412698	-0,989	0,0008952	0,00338291
		2.1 Metabolism of	,	,	,	,	,
PFCIRM1		carbohydrates and					
38 04560	Aldose 1-epimerase	related molecules	0,357	2,80112045	-1,488	6,0157E-08	7,4363E-07
		2.1 Metabolism of	- ,	,	,		,
		carbohvdrates and					
iolT3	ioIT3 (myo-inositol transporter ioIT3)	related molecules	0,564	1,77304965	-0,827	0,01230528	0,03247763
		2.1 Metabolism of	,	,	,	,	,
		carbohvdrates and					
pta	Phosphate acetyltransferase	related molecules	0,23	4,34782609	-2,122	8,9026E-14	3,3015E-12
-1		2.1 Metabolism of	,	,	,	,	,
		carbohydrates and					
ackA	Acetate kinase	related molecules	0,204	4,90196078	-2,291	8,1723E-19	8,1277E-17
	Glucosaminefructose-6-phosphate aminotransferase (	2.1 Metabolism of	-, -	,	, -	-,	- )
	Hexosephosphate aminotransferase. D-fructose-6-	carbohvdrates and					
glmS	phosphate amidotransferase)	related molecules	0.613	1,63132137	-0,706	0,0068971	0,01988255
0		2.1.1 Specific	,	,	,	,	,
	Glycerol kinase (ATP:glycerol 3-phosphotransferase)	carbohvdrate					
alpK	(Glvcerokinase) (GK)	metabolic pathway	0.157	6.36942675	-2.674	4.4048E-10	8.6826E-09
51		2.1.1 Specific	-, -		7 -	,	-,
	Citrate lyase beta chain (Citrase beta chain) (Citrate(Pro-	carbohydrate					
citE	3S)-lyase beta chain)	metabolic pathway	0,582	1,71821306	-0,782	0,0029122	0,00948198
		2.1.1 Specific	,	,	,	,	,
		carbohydrate					
lacZ	Beta-galactosidase (Lactase) LacZ	metabolic pathway	0.308	3,24675325	-1,699	9,7588E-07	9,2434E-06

	Anaerobic alvcerol-3-phosphate debydrogenase subunit	2.1.1 Specific					
alpB	B	metabolic pathway	0.501	1.99600798	-0.998	0.00012019	0.00060873
3.F -		2.1.1 Specific		.,	-,		-,
	Anaerobic glycerol-3-phosphate dehydrogenase subunit	carbohydrate					
alpC	C	metabolic pathway	0,462	2,16450216	-1,116	3,0035E-05	0,00018255
01		2.1.1 Specific					
	Trehalose-6-phosphate synthase (Alpha, alpha-trehalose-	carbohydrate					
otsA	phosphate synthase)	metabolic pathway	0,616	1,62337662	-0,698	0,00748217	0,02142799
		2.1.1 Specific		,	,		
rmIA		carbohydrate					
(rfbA)	Glucose-1-phosphate thymidylyltransferase	metabolic pathway	0,629	1,58982512	-0,669	0,00798454	0,02261805
		2.1.1 Specific					
		carbohydrate					
galK2	Galactokinase	metabolic pathway	0,541	1,84842884	-0,886	0,00017825	0,00084337
		2.1.1 Specific					
		carbohydrate					
araM	L-arabinose utilization protein	metabolic pathway	0,355	2,81690141	-1,493	8,2974E-05	0,00044172
		2.1.1 Specific					
iolA	ioIA (Myo-inositol catabolism IoIA protein) (Methylmalonic	carbohydrate					
(msdA)	acid semialdehyde dehydrogenase)	metabolic pathway	0,539	1,85528757	-0,893	0,00264533	0,00882316
		2.1.1 Specific					
		carbohydrate					
iolB	iolB (Myo-inositol catabolism IolB protein)	metabolic pathway	0,464	2,15517241	-1,108	0,00048368	0,00200815
		2.1.1 Specific					
PFCIRM1	Methylmalonyl-CoA carboxytransferase 5S subunit.	carbohydrate					
38_05815	(transcarboxylase 5S) 505 bp	metabolic pathway	0,546	1,83150183	-0,873	0,0054605	0,01634416
		2.1.1 Specific					
	Methylmalonyl-CoA carboxytransferase 12S subunit	carbohydrate					
mmdA	(EC2.1.3.1) (Transcarboxylase 12S subunit). 610 bp	metabolic pathway	0,417	2,39808153	-1,261	0,00012662	0,00063544
		2.1.1 Specific					
PFCIRM1		carbohydrate					
38_05825	Methylmalonyl-CoA carboxyltransferase 12S subunit	metabolic pathway	0,447	2,23713647	-1,161	0,00023995	0,00108923
		2.1.1 Specific					
	Methylmalonyl-CoA carboxytransferase, 1.3S subunit	carbohydrate					
bccp	(Transcarboxylase, 1.3S subunit). 123bp	metabolic pathway	0,344	2,90697674	-1,54	6,6959E-06	5,1769E-05

		2.1.1 Specific					
dalE2	LIDP-alucose 4-enimerase	metabolic pathway	0 549	1 82149362	-0.866	0 00038876	0 00167442
gailez		2 1 1 Specific	0,040	1,02140002	0,000	0,0000010	0,00107442
PFCIRM1		carbohydrate					
38 07970	inositol-1-phosphate synthase	metabolic pathway	0,319	3,13479624	-1,65	1,9053E-07	2,1827E-06
		2.1.1 Specific			,	,	
		carbohydrate					
treS	Trehalose synthase	metabolic pathway	0,501	1,99600798	-0,997	0,00011277	0,00058058
		2.1.1 Specific					
		carbohydrate					
рссВ	Propionyl-CoA carboxylase beta chain	metabolic pathway	0,319	3,13479624	-1,65	6,2064E-10	1,1707E-08
		2.1.1 Specific					
	Galactose-1-phosphate uridylyltransferase (UDP-	carbohydrate					
galT	glucosehexose-1-phosphate uridylyltransferase)	metabolic pathway	0,539	1,85528757	-0,893	0,00560477	0,0167074
		2.1.2 Main glycolytic					
tpi1	triosephosphate isomerase 1	pathways	0,505	1,98019802	-0,985	0,00658728	0,0191917
		2.1.2 Main glycolytic	0.440	0 00747440		0 00000 404	0 00407004
рдк	Phosphoglycerate kinase	pathways	0,449	2,22717149	-1,154	0,00023461	0,00107391
milia	Dumunete kinese 1	2.1.2 Main glycolytic	0.47	0 40705057	4 000	0.00052044	0.0004.0000
рукт	Pyruvate kinase i	pathways	0,47	2,12765957	-1,088	0,00053814	0,00218836
amp	Phosphoglycerate mutase (D-phosphoglycerate 2,3-	2.1.2 Main glycolytic	0 5 0 7	1 07020650	0.070	0.00100660	0.0044049
grip	phosphomulase)	2.1.2 Main alveabria	0,507	1,97236659	-0,979	0,00123669	0,0044948
Idb1	L Lactate debudrogenase	Z. I.Z Main glycolylic	0.466	2 14502275	1 101	6 19125 06	1 94765 05
		2 1 2 Main diveolutio	0,400	2,14592275	-1,101	0,10132-00	4,04700-00
fba1	Fructose-bisphosphate aldolase class II	2.1.2 Main grycorytic	0.53	1 88679245	-0.915	0 00770202	0 02101421
1501		2 1 2 Main alvcolvtic	0,00	1,00070240	0,010	0,00770202	0,02101421
ofkA	6-phosphofructokinase	pathways	0 457	2 18818381	-1 13	7 0413E-05	0 00038516
phot		2 1 2 Main glycolytic	0,101	2,10010001	1,10	1,01102 00	0,00000010
gmpB	phosphoglycerate mutase	pathways	0,464	2,15517241	-1,107	0,00493771	0,01488114
mdh	Malate dehydrogenase	2.1.3 TCA cycle	0,522	1,91570881	-0,938	0,00276718	0,00912165
fumC	Fumarate hydratase, class-II	2.1.3 TCA cycle	0,47	2,12765957	-1,089	0,00044967	0,00188482
icd	Putative isocitrate/isopropylmalate dehydrogenase	2.1.3 TCA cycle	0,485	2,06185567	-1,045	0,0004114	0,00175468
cat	Coenzyme A transferase (Putative succinyl-CoA or	2.1.4 Substrate-	0,421	2,37529691	-1,247	0,00011893	0,00060373

	butyryl-CoA:coenzyme A transferase)	specific entries to					
		carbohydrate					
		netabolic pathway					
		2.1.4 Substrate-					
rmIR		specific entries to					
(rfbB)	DTDP-alucoso 4.6-dobydrataso	metabolic pathway	0 402	2 03252033	-1 023	6 605E-05	0 00036806
		2.2 Metabolism of	0,492	2,03232033	-1,025	0,0956-05	0,00030800
		amino acids and					
ardH	Argininosuccinate Ivase (Arginosuccinase)	related molecules	0 479	2 08768267	-1.061	3 7005E-05	0 00021766
argin	Arginine biosynthesis bifunctional protein arg.l	2.2 Metabolism of	0,470	2,00700207	1,001	0,7000E 00	0,00021700
	(Glutamate N-acetyltransferase) (Ornithine	amino acids and					
arg.J	acetyltransferase) (Ornithine transacetylase) (OATase)	related molecules	0.347	2 88184438	-1 526	2 9771E-07	3 162E-06
u go	N-acetyl-gamma-glutamyl-phosphate reductase (AGPR)	2.2 Metabolism of	0,011	2,00101100	1,020	2,01112 01	0,1022 00
	(N- acetyl-glutamate semialdehyde dehydrogenase)	amino acids and					
argC	(NAGSA dehydrogenase)	related molecules	0.256	3,90625	-1,963	1,7231E-12	5,3858E-11
- 0		2.2 Metabolism of	,	,	,	,	,
		amino acids and					
hisG	ATP phosphoribosyltransferase	related molecules	0,633	1,57977883	-0,66	0,00677543	0,01960931
		2.2 Metabolism of					
		amino acids and					
hisE	Phosphoribosyl-ATP pyrophosphohydrolase	related molecules	0,501	1,99600798	-0,998	0,00289431	0,00945188
		2.2 Metabolism of					
		amino acids and					
trpD	Anthranilate phosphoribosyltransferase	related molecules	0,271	3,6900369	-1,882	4,6178E-13	1,5544E-11
		2.2 Metabolism of					
PFCIRM1		amino acids and					
38_02155	Lysine decarboxylase	related molecules	0,463	2,15982721	-1,112	1,1428E-05	8,1446E-05
		2.2 Metabolism of					
PFCIRM1		amino acids and					
38_02175	aminotransferase	related molecules	0,317	3,15457413	-1,658	8,9783E-11	2,0252E-09
		2.2 Metabolism of					
	Histidinol-phosphate aminotransferase (Imidazole acetol-	amino acids and					
nisC	phosphate transaminase)	related molecules	0,518	1,93050193	-0,948	0,00016639	0,00079316
		2.2 Metabolism of					
hisB	Imidazolegiycerol-phosphate dehydratase (IGPD)	amino acids and	0,44	2,27272727	-1,186	0,00010079	0,00052382

		related molecules					
	Phosphoribosyl isomerase A (1-(5-phosphoribosyl)-5-[(5-						
	phosphoribosylamino)methylideneamino] imidazole-4-						
	carboxamide isomerase) (Phosphoribosylformimino-5-	2.2 Metabolism of					
	aminoimidazole carboxamide ribotide isomerase) (N-(5 -	amino acids and					
hisA	phosphoribosyl)anthranilate isomerase) (PRAI)	related molecules	0,5	2	-1,001	0,00037469	0,00162341
		2.2 Metabolism of					
carB	Carbamoyl-phosphate synthase large chain (Carbamoyl-	amino acids and					
(pyrA)	phosphate synthetase ammonia chain)	related molecules	0,397	2,51889169	-1,332	5,4695E-05	0,00030685
		2.2 Metabolism of					
	methionine synthase (5-methyltetrahydrofolate:L-	amino acids and					
metH	homocysteine S-methyltransferase)	related molecules	0,474	2,10970464	-1,076	1,9411E-05	0,00012716
	Ketol-acid reductoisomerase (Acetohydroxy-acid	2.2 Metabolism of					
	isomeroreductase) (Alpha-keto-beta-hydroxylacil	amino acids and					
ilvC	reductoisomerase)	related molecules	0.208	4.80769231	-2.262	6.3096E-17	3.7312E-15
		2.2 Metabolism of		.,	_,		
		amino acids and					
ilvD	Dihydroxy-acid dehydratase	related molecules	0.46	2 17391304	-1 122	2 6212E-05	0 00016287
		2.2 Metabolism of	0,10	_,	.,	_,0 00	0,000.020.
		amino acids and					
asnB	Asparagine synthase (Glutamine-hydrolyzing)	related molecules	0.672	1 48809524	-0.574	0 0171151	0 04270219
done		2.2 Metabolism of	0,012	1,10000021	0,07 1	0,0171101	0,01210210
		amino acids and					
ansA			0.472	2 11864407	-1 082	8 953E-05	0 000/7203
ansA		2.2 Motobolism of	0,472	2,11004407	-1,002	0,355⊑-05	0,00047203
		amino acide and					
ask	Aspartokipaso (Aspartato kipaso)	related molecules	0.557	1 70533214	-0.845	0.00405315	0.01252584
asn		2.2 Motobolism of	0,337	1,79000214	-0,043	0,00403313	0,01232304
0.00	avetaina avethaaa 2	amino acius anu	0.057	17 5 429506	4 4 2 5	4 47045 44	6 1000E 10
Cysz		Related molecules	0,057	17,5436596	-4,135	1,4704⊏-14	0,10332-13
DECIDINA		2.2 Metabolism of					
PFCIRMI		amino acids and	0.444	0.05005005	4 4 70	0 004 404 50	0.00540070
38_09945	alanine racemase	related molecules	0,444	2,25225225	-1,172	0,00146153	0,00519973
		2.2 Metabolism of					
		amino acids and					
tyrB	Aspartate transaminase (Aminotransferase)	related molecules	0,418	2,3923445	-1,259	3,3558E-05	0,00019898

		2.2 Metabolism of					
Tdep	1-aminohutyrate aminotransferase	related molecules	0 152	6 5780/737	-2 71/	5 2032E-10	5 4212E-17
gabi		2.2 Metabolism of	0,152	0,57034757	-2,714	5,20522-13	5,4212L-17
		amino acids and					
lvsA	Diaminonimelate decarboxylase (DAP decarboxylase)	related molecules	0 552	1 8115942	-0.857	0 00051744	0 00211294
1937		2.2 Metabolism of	0,002	1,0110042	0,007	0,00001744	0,00211234
		amino acids and					
thrA/hom	Homoserine dehydrogenase	related molecules	0 4 1 1	2 43309002	-1 283	1 7656E-07	2 0549E-06
		2.2 Metabolism of	0,411	2,40000002	1,200	1,70002 07	2,00402.00
	Phosphoserine phosphatase/homoserine	amino acids and					
serB/thrH	phosphotransferase bifunctional protein	related molecules	0.36	2 77777778	-1 475	5 9958E-08	7 4363E-07
0012, 1111	dihydrolipovllysine-residue (2-		0,00	2,	.,	0,00002 00	1,10002 01
	methylpropanovl)transferase Lipoamide acyltransferase						
	component of branched-chain alpha-keto acid	2.2 Metabolism of					
	dehvdrogenase complex (E2) (Dihvdrolipoamide	amino acids and					
bkdB	branched chain transacylase)	related molecules	0,375	2,66666667	-1,416	2,0794E-07	2,3399E-06
	2-oxoisovalerate dehydrogenase subunit beta (EC		,	,	,	,	,
	1.2.4.4) (Branched- chain alpha-keto acid dehydrogenase	2.2 Metabolism of					
	E1 component beta chain) (BCKDH E1-beta) Pyruvate	amino acids and					
bkdA2	dehydrogenase E1 component subunit beta	related molecules	0,341	2,93255132	-1,554	1,5683E-08	2,2428E-07
	2-oxoisovalerate dehydrogenase subunit alpha	2.2 Metabolism of					
	(Branched- chain alpha-keto acid dehydrogenase E1	amino acids and					
bkdA1	component alpha chain) (BCKDH E1-alpha)	related molecules	0,524	1,90839695	-0,932	0,00010967	0,00056593
		2.2 Metabolism of					
		amino acids and					
argG	Argininosuccinate synthase	related molecules	0,324	3,08641975	-1,625	2,3102E-07	2,5274E-06
		2.2 Metabolism of					
PFCIRM1		amino acids and					
38_11035	Chorismate mutase	related molecules	0,446	2,24215247	-1,166	7,3782E-05	0,00039959
		2.2 Metabolism of					
		amino acids and					
cysE	Serine acetyltransferase	related molecules	0,55	1,81818182	-0,863	0,00324044	0,01027548
		2.2 Metabolism of					
		amino acids and					
cys1	Cysteine synthase 1	related molecules	0,585	1,70940171	-0,774	0,00184305	0,0063706

		2.3 Metabolism of					
ors	Ribose-phosphate pyrophosphokinase	nucleonaes and	0 337	2 96735905	-1 571	3 3471E-09	5 3455E-08
pis		2.3 Metabolism of	0,007	2,30733300	1,071	0,047 TE 00	5,5455E 00
	Glutamine amidotransferase subunit pdxT (Glutamine	nucleotides and					
Txba	amidotransferase glutaminase subunit pdxT (Ciduannie	nucleic acids	0 44	2 27272727	-1 184	0.00300551	0 00967066
ралт		2.3 Metabolism of	0,11	2,21212121	1,101	0,00000001	0,00007000
		nucleotides and					
purM	Phosphoribosylformylglycinamidine cyclo-ligase	nucleic acids	0 628	1 59235669	-0.67	0 01564276	0 03975188
punn		2.3 Metabolism of	0,020	1,00200000	0,01	0,01001210	0,00010100
		nucleotides and					
rnd	Ribonuclease D (3 -5 exonuclease)	nucleic acids	0.386	2 59067358	-1 374	7 0629E-08	8 629E-07
		2.3 Metabolism of	0,000	_,0000.000	.,01 1	.,	0,0202 01
	Adenine phosphoribosyltransferase (AMP:diphosphate	nucleotides and					
apt	phospho-D-ribosvltransferase)	nucleic acids	0.32	3.125	-1.645	4.6044E-07	4.7077E-06
		2.3 Metabolism of	- / -	-, -	,	,	,
		nucleotides and					
pvrD	Dihydroorotate dehydrogenase 2	nucleic acids	0.537	1.86219739	-0.898	0.00077684	0.00298983
		2.3 Metabolism of		/	- ,	- /	- ,
	Uracil phosphoribosyltransferase (UMP	nucleotides and					
upp	pyrophosphorylase) (UPRTase)	nucleic acids	0,526	1,90114068	-0,926	0,01783752	0,04414987
	ribose-phosphate pyrophosphokinase (RPPK)	2.3 Metabolism of					
PFCIRM1	(Phosphoribosyl pyrophosphate synthetase) (P-Rib-PP	nucleotides and					
38_04525	synthetase) (PRPP synthetase)	nucleic acids	0,411	2,43309002	-1,283	4,5375E-07	4,683E-06
		2.3 Metabolism of					
		nucleotides and					
pyrE	Orotate phosphoribosyltransferase (OPRT) (OPRTase)	nucleic acids	0,602	1,66112957	-0,733	0,01344317	0,03505799
		2.3 Metabolism of					
		nucleotides and					
hpt	Hypoxanthine phosphoribosyltransferase	nucleic acids	0,324	3,08641975	-1,628	3,8731E-08	5,105E-07
		2.3 Metabolism of					
		nucleotides and					
gpt	Putative purine phosphoribosyltransferase	nucleic acids	0,444	2,25225225	-1,17	2,2522E-06	1,9791E-05
		2.3 Metabolism of					
PFCIRM1		nucleotides and					
38_10750	Dihydroorotate dehydrogenase	nucleic acids	0,272	3,67647059	-1,88	9,3166E-06	6,9101E-05

	Inosine-5 -monophosphate dehydrogenase (IMP	2.3 Metabolism of nucleotides and					
guaB2	dehydrogenase) (IMPDH) (IMPD) / GMP reductase	nucleic acids	0,476	2,10084034	-1,07	0,00014279	0,00070049
		2.3 Metabolism of					
	Phosphoribosylaminoimidazolecarboxamide	nucleotides and					
purH	formyltransferase (AICAR transformylase)	nucleic acids	0,398	2,51256281	-1,33	3,3022E-05	0,00019634
		2.4 Metabolism of					
ugpQ2	Glycerophosphoryl diester phosphodiesterase	lipids	0,241	4,14937759	-2,052	3,7194E-10	7,4661E-09
		2.4 Metabolism of					
cdsA	Phosphatidate cytidylyltransferase	lipids	0,516	1,9379845	-0,955	0,00058516	0,0023579
		2.4 Metabolism of					
inhA	Enoyl-[acyl-carrier-protein] reductase (NADH)	lipids	0,591	1,69204738	-0,758	0,00825452	0,02327434
		2.4 Metabolism of					
fabG	3-oxoacyl-[acyl-carrier protein] reductase	lipids	0,529	1,89035917	-0,92	0,00967845	0,02650153
		2.4 Metabolism of					
plsC3	Acyltransferase PIsC	lipids	0,592	1,68918919	-0,756	0,00480094	0,01456929
PFCIRM1		2.4 Metabolism of					
38_02490	Phosphatidylglycerophosphate synthase	lipids	0,631	1,58478605	-0,665	0,01122571	0,0301003
	CDP-diacylglycerolglycerol-3-phosphate 3-	2.4 Metabolism of					
pgs2	phosphatidyl-transferase	lipids	0,376	2,65957447	-1,409	0,00013478	0,00066855
		2.4 Metabolism of					
PPA2234	Long-chain-fatty-acidCoA ligase/synthetase	lipids	0,574	1,74216028	-0,802	0,0029171	0,00948383
		2.4 Metabolism of					
pf1861	Carboxylic ester hydrolase	lipids	0,485	2,06185567	-1,045	9,3149E-05	0,00048759
fabF	3-oxoacyl-[acyl-carrier-protein] synthase (Beta-ketoacyl-	2.4 Metabolism of					
(fabB)	ACP synthase)	lipids	0,159	6,28930818	-2,652	2,6775E-17	1,8307E-15
		2.4 Metabolism of					
acpP	Acyl carrier protein (ACP)	lipids	0,169	5,91715976	-2,569	1,049E-18	9,5632E-17
		2.4 Metabolism of					
fabH	3-oxoacyl-(Acyl-carrier-protein) synthase III	lipids	0,145	6,89655172	-2,782	1,2113E-22	2,9449E-20
		2.4 Metabolism of					
fabD	Malonyl CoA-acyl carrier protein transacylase	lipids	0,271	3,6900369	-1,884	4,0889E-12	1,1929E-10
		2.4 Metabolism of					
accA	Acetyl-CoA carboxylase	lipids	0,598	1,6722408	-0,742	0,00763265	0,0217451
PFCIRM1		2.5 Metabolism of					
38_00065	2-dehydropantoate 2-reductase	coenzymes and	0,324	3,08641975	-1,624	4,8218E-09	7,645E-08

I			prosthetic groups					
			2.5 Metabolism of					
			coenzymes and					
	pntB	NADH dehvdrogenase	prosthetic groups	0,345	2,89855072	-1,533	4,9616E-06	3,9766E-05
	•	, ,	2.5 Metabolism of					
			coenzymes and					
	pntA	NAD(P)(+) transhydrogenase (AB-specific).	prosthetic groups	0,331	3,02114804	-1,595	9,0372E-07	8,6347E-06
ĺ	•	Phosphopantetheine adenvlvltransferase (Pantetheine-	2.5 Metabolism of	,	,	,	,	,
		phosphate adenylyltransferase) (PPAT) (Dephospho-CoA	coenzymes and					
	coaD	pyrophosphorylase)	prosthetic groups	0,55	1,81818182	-0,862	0,00465722	0,01417245
		1-deoxy-D-xylulose 5-phosphate reductoisomerase (DXP						
		reductoisomerase) (1-deoxyxylulose-5-phosphate	2.5 Metabolism of					
		reductoisomerase) (2-C- methyl-D-erythritol 4-phosphate	coenzymes and					
	dxr	synthase)	prosthetic groups	0,569	1,75746924	-0,814	0,00346053	0,01089446
		· · ·	2.5 Metabolism of			,		
			coenzymes and					
	pdxS	Pyridoxal biosynthesis lyase pdxS	prosthetic groups	0,342	2,92397661	-1,549	2,0466E-07	2,3202E-06
ĺ	•		2.5 Metabolism of					
			coenzymes and					
	thiL	Thiamine monophosphate kinase	prosthetic groups	0,61	1,63934426	-0,714	0,00273067	0,00905258
			2.5 Metabolism of					
		CobU Bifunctional cobalamin biosynthesis pyrophosphate	coenzymes and					
	cobU	enzyme	prosthetic groups	0,355	2,81690141	-1,493	0,00937282	0,02575884
			2.5 Metabolism of					
			coenzymes and					
	cbiA	Cobyrinic acid A,C-diamide synthase, CbiA	prosthetic groups	0,487	2,05338809	-1,037	0,003179	0,01010995
			2.5 Metabolism of					
			coenzymes and					
	cobA2	Cob(I)alamin adenosyltransferase	prosthetic groups	0,414	2,41545894	-1,273	0,01829595	0,04502985
			2.5 Metabolism of					
			coenzymes and					
	cbiP/cobQ	Cobyric acid synthase CbiP/CobQ	prosthetic groups	0,464	2,15517241	-1,108	0,00590639	0,01751107
			2.5 Metabolism of					
			coenzymes and					
	idsA	Heptaprenyl diphosphate synthase component II	prosthetic groups	0,194	5,15463918	-2,363	2,839E-17	1,8623E-15

		2.5 Metabolism of					
h a ra A		coenzymes and	0.000	2 24440404	4 744		0 00145 00
nemA	Glutamyl-tRINA reductase	prostnetic groups	0,299	3,34448161	-1,741	5,6016E-09	8,6311E-08
	Demokratika e negode envigence (DDO) (Ukudaren mertkedkilene	2.5 Metabolism of					
	Porphobilinogen deaminase (PBG) (Hydroxymethylbilane	coenzymes and	0.000	0.04004040	4 50	0.00405004	0 00 4500 40
nemC	synthase) (HMBS) (Pre-uroporphyrinogen synthase)	prostnetic groups	0,332	3,01204819	-1,59	0,00125684	0,00456048
		2.5 Metabolism of					
		coenzymes and					
hemE	Uroporphyrinogen decarboxylase	prosthetic groups	0,381	2,62467192	-1,391	0,00492605	0,01486647
		2.5 Metabolism of					
		coenzymes and					
hemY	Protoporphyrinogen oxidase (PPO) HemY	prosthetic groups	0,314	3,18471338	-1,671	0,00044139	0,00186438
		2.5 Metabolism of					
	Delta-aminolevulinic acid dehydratase (Porphobilinogen	coenzymes and					
hemB	synthase)	prosthetic groups	0,267	3,74531835	-1,903	4,8843E-05	0,00027903
		2.5 Metabolism of					
		coenzymes and					
ribC (ribE)	Riboflavin synthase alpha chain	prosthetic groups	0,494	2,0242915	-1,017	0,00267583	0,00889775
		2.5 Metabolism of					
	GTP cyclohydrolase II protein, Riboflavin biosynthesis	coenzymes and					
ribAB	protein	prosthetic groups	0,402	2,48756219	-1,316	0,00013753	0,00067926
		2.5 Metabolism of					
		coenzymes and					
cobA	CobA Uroporphyrinogen III methyltransferase	prosthetic groups	0,312	3,20512821	-1,678	7,9359E-05	0,00042454
		2.5 Metabolism of					
		coenzymes and					
chID	ChID, Mg-chelatase subunit ChID	prosthetic groups	0,5	2	-1,001	0,01779438	0,04409299
		2.5 Metabolism of					
		coenzymes and					
dkgA	2,5-diketo-D-gluconate reductase A	prosthetic groups	0,447	2,23713647	-1,161	0,00047263	0,00196598
-	Phosphoribosyltransferase/nitroreductase (fusion gene)	2.5 Metabolism of					
bluB/cobT	(Nicotinate-nucleotide-dimethylbenzimidazole	coenzymes and					
2	phosphoribosyltransferase)	prosthetic groups	0,339	2,94985251	-1,562	1,9317E-08	2,7445E-07
		2.5 Metabolism of					
	Methylenetetrahydrofolate dehydrogenase (Bifunctional	coenzymes and					
folD	protein)	prosthetic groups	0,397	2,51889169	-1,333	2,2664E-07	2,4919E-06

		2.5 Metabolism of					
	Naphthoate synthase (Dihydroxynaphthoic acid	coenzymes and					
menB	synthetase)	prosthetic groups	0,46	2,17391304	-1,12	2,9891E-05	0,00018218
		2.5 Metabolism of					
		coenzymes and					
cbiL	CbiL Precorrin-2 C20-methyltransferase	prosthetic groups	0,43	2,3255814	-1,218	0,00013492	0,00066855
		2.5 Metabolism of					
		coenzymes and					
cbiF	CbiF Precorrin-4 C11-methyltransferase	prosthetic groups	0,332	3,01204819	-1,592	1,5932E-05	0,00010726
		2.5 Metabolism of					
		coenzymes and					
cysG/cbiX	CysG/CbiX	prosthetic groups	0,441	2,2675737	-1,181	0,00086698	0,00329333
		2.5 Metabolism of					
		coenzymes and					
folC	Folylpolyglutamate synthase (bifunctionnal enzyme)	prosthetic groups	0,656	1,52439024	-0,608	0,01364176	0,03553355
PFCIRM1		2.6 Metabolism of					
38_06220	NUDIX hydrolase	phosphate	0,392	2,55102041	-1,353	1,5664E-06	1,434E-05
PFCIRM1		2.6 Metabolism of					
38 08215	Inorganic pyrophosphatase	phosphate	0,478	2,09205021	-1,064	0,00076125	0,0029376
		2.7 Metabolism of					
cysW	Sulfate transport system permease protein CysW	sulfur	0,081	12,345679	-3,62	2,4676E-07	2,6862E-06
	Sulphate transport system permease protein 2:Sulfate	2.7 Metabolism of					
cysT	ABC transporter, permease protein CysT	sulfur	0,097	10,3092784	-3,368	6,5968E-07	6,4725E-06
PFCIRM1		2.7 Metabolism of					
38 00925	Sulfate-binding protein precursor	sulfur	0,118	8,47457627	-3,078	4,7385E-05	0,00027212
		2.7 Metabolism of	,	,	,	,	,
nirA2/sir2	Sulfite reductase [ferredoxin]	sulfur	0.076	13,1578947	-3,717	1,1963E-09	2,1456E-08
		2.7 Metabolism of	- ,	-,	- ,	,	,
cvsH	Putative phosphoadenosine phosphosulfate reductase	sulfur	0.051	19.6078431	-4.3	2.4806E-17	1.7508E-15
	Sulfate adenvivitransferase subunit 2 (Sulfate adenviate			,	.,_		.,
mmcV/cvs	transferase) (SAT) (ATP-sulfurvlase small subunit)	2.7 Metabolism of					
D	(Mitomycin biosynthesis protein V)	sulfur	0.035	28.5714286	-4.827	7.1865E-27	2.2463E-24
	Sulfate adenvlvltransferase, large subunit/ ATP-	2.7 Metabolism of	- ,	-,-	,	,	,
cvsN	sulfurvlase, subunit 1 (ATP:sulfate adenvlvltransferase)	sulfur	0.033	30.3030303	-4.932	1.7165E-32	9.3895E-30
		2.8 Metabolism of	-,		.,	, <b></b>	.,
norP	Nitric-oxide reductase subunit B (nitric-oxide reductase)	nitrogen/nitrate and	0 475	2 10526316	-1 073	0 00068892	0 00271004

		nitrite					
		2.8 Metabolism of					
		nitrogen/nitrate and					
glnA1	Glutamine synthetase	nitrite	0,543	1,84162063	-0,882	0,00408826	0,01261651
dnaN	DNA polymerase III, beta chain	3.1 DNA replication	0,493	2,02839757	-1,019	0,0005583	0,00225798
PFCIRM1							
38_07260	Exonuclease	3.1 DNA replication	0,393	2,54452926	-1,346	5,3832E-06	4,2603E-05
PFCIRM1							
38_09150	DeaD/DeaH box helicase	3.1 DNA replication	0,583	1,71526587	-0,779	0,00197229	0,00675333
		3.10 Protein					
pepP	Xaa-Pro aminopeptidase I	degradation	0,583	1,71526587	-0,78	0,00361727	0,01130657
PFCIRM1		3.10 Protein					
38_02300	Zinc metallopeptidase	degradation	0,562	1,77935943	-0,831	0,00608512	0,01791958
PFCIRM1	Amidohydrolase (Peptidase M20D) (Putative metal-	3.10 Protein					
38_03755	dependent amidase/aminoacylase/carboxypeptidase)	degradation	0,418	2,3923445	-1,26	0,00551416	0,01648223
		3.10 Protein					
map	Methionine aminopeptidase (MAP) (Peptidase M)	degradation	0,315	3,17460317	-1,669	1,4872E-06	1,3673E-05
PFCIRM1		3.10 Protein					
38_10230	Secreted protease with a PDZ domain	degradation	0,312	3,20512821	-1,682	1,7693E-11	4,3992E-10
		3.10 Protein					
pepN	Aminopeptidase N, Lysyl aminopeptidase	degradation	0,521	1,9193858	-0,94	0,0039231	0,01217343
	ATP-dependent Clp protease proteolytic subunit 2	3.10 Protein					
clpP2	(Endopeptidase Clp 2)	degradation	0,583	1,71526587	-0,777	0,0181493	0,04471922
		3.2 DNA restriction					
PFCIRM1		and modification					
38_05540	nuclease (RecB family)	(and repair)	0,639	1,56494523	-0,646	0,01158533	0,030892
		3.2 DNA restriction					
		and modification					
nth	Putative endonuclease III	(and repair)	0,536	1,86567164	-0,9	0,00227571	0,0076968
		3.2 DNA restriction					
PFCIRM1		and modification					
38_06795	Hypothetical protein	(and repair)	0,466	2,14592275	-1,1	0,00888308	0,02460276
		3.3 DNA					
		recombination, and					
recB	Exodeoxyribonuclease V beta chain RecB	repair	0,657	1,52207002	-0,607	0,01511099	0,03871528

PFCIRM1		3.5.1 Transcription			1		
38_11040	transcription initiation factor	initiation	0,631	1,58478605	-0,664	0,00899062	0,02486914
		3.5.2 Transcription					
rnc	Ribonuclease III	regulation	0,615	1,62601626	-0,701	0,00989615	0,02699846
		3.5.2 Transcription					
dskA	Transcriptional regulators, TraR/DksA family	regulation	0,433	2,30946882	-1,206	7,5969E-06	5,7716E-05
PFCIRM1		3.5.2 Transcription					
38_01955	Response regulator receiver protein	regulation	0,417	2,39808153	-1,262	1,9882E-06	1,7755E-05
		3.5.2 Transcription					
merR1	Transcriptional regulator, MerR	regulation	0,587	1,70357751	-0,767	0,00113456	0,00419328
PFCIRM1		3.5.2 Transcription					
38_02925	Transcriptional regulator, MerR	regulation	0,479	2,08768267	-1,062	0,00026844	0,0012012
		3.5.2 Transcription					
deoR3	Putative regulatory protein, DeoR family	regulation	0,503	1,98807157	-0,991	0,00029069	0,00129528
PFCIRM1		3.5.2 Transcription					
38_04635	Transcriptional Regulator, GntR family protein	regulation	0,39	2,56410256	-1,359	5,9982E-06	4,7209E-05
PFCIRM1		3.5.2 Transcription					
38_06690	two component system response regulator	regulation	0,27	3,7037037	-1,888	5,6001E-10	1,0748E-08
PFCIRM1		3.5.2 Transcription					
38_08370	Two-component system response regulator	regulation	0,447	2,23713647	-1,162	6,7331E-06	5,1873E-05
		3.5.2 Transcription					
sirR	Iron dependent repressor	regulation	0,232	4,31034483	-2,106	2,1627E-10	4,5941E-09
PFCIRM1		3.5.2 Transcription					
38_09335	Excisionase	regulation	0,375	2,66666667	-1,415	1,4199E-06	1,3109E-05
PFCIRM1		3.5.2 Transcription					
38_10835	two-component response regulator LuxR	regulation	0,544	1,83823529	-0,878	0,0085986	0,0240893
PFCIRM1		3.5.2 Transcription					
38_10890	Cell envelope-related transcriptional attenuator	regulation	0,652	1,53374233	-0,618	0,00875209	0,02439059
	DNA-directed RNA polymerase beta chain (RNAP beta						
	subunit) (Transcriptase beta chain) (RNA polymerase	3.5.3 Transcription					
rpoC	beta subunit)	elongation	0,502	1,99203187	-0,994	0,00938291	0,02575884
	DNA-directed RNA polymerase alpha chain (RNAP alpha						
	subunit) (Transcriptase alpha chain) (RNA polymerase	3.5.3 Transcription					
rpoA	subunit alpha)	elongation	0,253	3,95256917	-1,983	1,0341E-09	1,9174E-08
_		3.5.4 Transcription					
nusG	Transcription antitermination protein NusG	termination	0,57	1,75438596	-0,81	0,00287161	0,00940582

		3.6 RNA					
spoU1	rRNA methylase	modification	0,392	2,55102041	-1,35	6,7802E-05	0,0003718
PFCIRM1		3.6 RNA					
38_05905	tRNA/rRNA methyltransferase	modification	0,481	2,07900208	-1,057	0,00020819	0,00096917
	·	3.6 RNA					
rsmC	rRNA (guanine-N2-)-methyltransferase	modification	0,589	1,69779287	-0,764	0,01885313	0,04588504
PFCIRM1		3.6 RNA					
38_10045	GatB_Yqey, GatB/Yqey	modification	0,651	1,53609831	-0,62	0,01167347	0,0310347
		3.6 RNA					
orn	Oligoribonuclease	modification	0,33	3,03030303	-1,6	4,508E-10	8,8068E-09
	Glutamyl-tRNA(Gln) amidotransferase subunit C						
	(Aspartyl/glutamyl-tRNA(Asn/Gln) amidotransferase	3.7 Protein					
gatC	subunit C)	synthesis	0,437	2,28832952	-1,194	8,5304E-05	0,00045193
	Aspartyl/glutamyl-tRNA(Asn/Gln) amidotransferase	3.7 Protein					
gatB	subunit B (Asp/Glu-ADT subunit B)	synthesis	0,343	2,9154519	-1,544	3,9131E-08	5,1268E-07
		3.7.1 Ribosomal					
rplY	Ribosomal protein L25	proteins	0,308	3,24675325	-1,701	1,4762E-09	2,5634E-08
		3.7.1 Ribosomal					
rpIT	50S ribosomal protein L20	proteins	0,276	3,62318841	-1,858	5,1494E-08	6,5548E-07
		3.7.1 Ribosomal					
rpml	50S ribosomal protein L35	proteins	0,331	3,02114804	-1,593	2,713E-09	4,497E-08
		3.7.1 Ribosomal					
rpsA	30S ribosomal protein S1	proteins	0,503	1,98807157	-0,992	0,00452294	0,01384083
		3.7.1 Ribosomal					
rpsR	30S ribosomal protein S18	proteins	0,487	2,05338809	-1,038	0,0007434	0,00288761
		3.7.1 Ribosomal					
rpll	50S ribosomal protein L9	proteins	0,46	2,17391304	-1,12	0,00066158	0,00261288
		3.7.1 Ribosomal					
rpsK	30S ribosomal protein S11	proteins	0,493	2,02839757	-1,019	0,00059883	0,00240409
		3.7.1 Ribosomal					
rplA	50S ribosomal protein L1	proteins	0,536	1,86567164	-0,899	0,0063055	0,01849387
		3.7.1 Ribosomal					
rplJ	50S ribosomal protein L10	proteins	0,064	15,625	-3,963	2,5466E-21	4,509E-19
		3.7.1 Ribosomal					
rplL	50S ribosomal protein L7/L12	proteins	0,063	15,8730159	-3,991	2,679E-21	4,509E-19

		3.7.1 Ribosomal					
rpsG	30S ribosomal protein S7	proteins	0,625	1,6	-0,678	0,01500913	0,03849939
		3.7.1 Ribosomal					
rpsJ	30S ribosomal protein S10	proteins	0,36	2,77777778	-1,474	3,5353E-08	4,7166E-07
		3.7.1 Ribosomal					
rplC	50S ribosomal protein L3	proteins	0,339	2,94985251	-1,561	1,1246E-09	2,0486E-08
		3.7.1 Ribosomal					
rpID	50S ribosomal protein L4	proteins	0,358	2,79329609	-1,481	7,9489E-09	1,1913E-07
		3.7.1 Ribosomal					
rplW	50S ribosomal protein L23	proteins	0,265	3,77358491	-1,914	1,0791E-14	4,5404E-13
		3.7.1 Ribosomal					
rplB	50S ribosomal protein L2	proteins	0,193	5,18134715	-2,37	2,124E-15	1,0562E-13
		3.7.1 Ribosomal					
rpsS	30S ribosomal protein S19	proteins	0,178	5,61797753	-2,493	2,4001E-18	2,0198E-16
		3.7.1 Ribosomal					
rplV	50S ribosomal protein L22	proteins	0,2	5	-2,325	4,933E-11	1,1606E-09
		3.7.1 Ribosomal					
rpsC	30S ribosomal protein S3	proteins	0,18	5,55555556	-2,47	8,7151E-12	2,2974E-10
		3.7.1 Ribosomal					
rplP	50S ribosomal protein L16	proteins	0,135	7,40740741	-2,892	7,5514E-14	2,8487E-12
		3.7.1 Ribosomal					
rpmC	50S ribosomal protein L29	proteins	0,131	7,63358779	-2,929	2,4445E-13	8,7682E-12
		3.7.1 Ribosomal					
rpsQ	30S ribosomal protein S17	proteins	0,165	6,06060606	-2,601	1,9039E-10	4,1245E-09
		3.7.1 Ribosomal					
rplN	50S ribosomal protein L14	proteins	0,203	4,92610837	-2,303	3,4743E-10	7,0388E-09
		3.7.1 Ribosomal					
rplX	50S ribosomal protein L24	proteins	0,19	5,26315789	-2,393	4,2376E-10	8,4289E-09
	·	3.7.1 Ribosomal					
rplE	50S ribosomal protein L5	proteins	0,183	5,46448087	-2,454	2,2501E-09	3,8165E-08
rpsN1,	•	3.7.1 Ribosomal					
rpsZ	30S ribosomal protein S14 type Z	proteins	0,145	6,89655172	-2,783	1,3472E-08	1,9393E-07
		3.7.1 Ribosomal					
rpsH	30S ribosomal protein S8	proteins	0,097	10,3092784	-3,358	1,557E-15	7,9444E-14
-		3.7.1 Ribosomal					
rplF	50S ribosomal protein L6	proteins	0,122	8,19672131	-3,032	1,5666E-12	5,0407E-11

		3.7.1 Ribosomal					
rplR	ribosomal protein L18	proteins	0,113	8,84955752	-3,141	7,6417E-12	2,0642E-10
_		3.7.1 Ribosomal					
rpsE	30S ribosomal protein S5	proteins	0,164	6,09756098	-2,608	5,3134E-08	6,7201E-07
_		3.7.1 Ribosomal					
rpmD	50S ribosomal protein L30	proteins	0,132	7,57575758	-2,926	2,1034E-09	3,5955E-08
		3.7.1 Ribosomal					
rplO	50S ribosomal protein L15	proteins	0,101	9,9009901	-3,305	9,1233E-12	2,3764E-10
_		3.7.1 Ribosomal					
rpsD	30S ribosomal protein S4	proteins	0,346	2,89017341	-1,531	3,8986E-06	3,2189E-05
		3.7.1 Ribosomal					
rplQ	50S ribosomal protein L17	proteins	0,24	4,16666667	-2,057	8,8665E-11	2,0208E-09
		3.7.1 Ribosomal					
rspl	30S ribosomal protein S9	proteins	0,629	1,58982512	-0,668	0,01958507	0,04740248
	Tyrosyl-tRNA synthetase (TyrosinetRNA ligase)	3.7.2 Aminoacyl-					
tyrS	(TyrRS)	tRNA synthetases	0,551	1,81488203	-0,86	0,00078642	0,00301876
	Phenylalanyl-tRNA synthetase beta chain	3.7.2 Aminoacyl-					
pheT	(Phenylalanine tRNA ligase beta chain) (PheRS)	tRNA synthetases	0,572	1,74825175	-0,805	0,00659853	0,01919891
		3.7.2 Aminoacyl-					
proS	Prolyl-tRNA synthetase (ProlinetRNA ligase) (ProRS)	tRNA synthetases	0,622	1,60771704	-0,684	0,01208965	0,03202441
	Threonyl-tRNA synthetase (ThreoninetRNA ligase)	3.7.2 Aminoacyl-					
thrS	(ThrRS)	tRNA synthetases	0,303	3,30033003	-1,723	1,5531E-11	3,9978E-10
		3.7.2 Aminoacyl-					
glnS	Glutaminyl-tRNA synthetase	tRNA synthetases	0,421	2,37529691	-1,247	3,9158E-07	4,0799E-06
	Histidyl-tRNA synthetase (HistidinetRNA ligase)	3.7.2 Aminoacyl-					
hisS	(HisRS)	tRNA synthetases	0,499	2,00400802	-1,004	3,7977E-05	0,00022158
	Aspartyl-tRNA synthetase (AspartatetRNA ligase)	3.7.2 Aminoacyl-					
aspS	(AspRS)	tRNA synthetases	0,469	2,13219616	-1,093	1,3341E-05	9,2323E-05
		3.7.2 Aminoacyl-					
serS1	Seryl-tRNA synthetase	tRNA synthetases	0,398	2,51256281	-1,331	1,3601E-06	1,261E-05
		3.7.2 Aminoacyl-					
ileS	Isoleucyl-tRNA synthetase (IsoleucinetRNA ligase)	tRNA synthetases	0,327	3,05810398	-1,613	5,0045E-09	7,8775E-08
		3.7.2 Aminoacyl-					
argS	Arginyl-tRNA synthetase (ArgininetRNA ligase) (ArgRS)	tRNA synthetases	0,614	1,6286645	-0,705	0,01027066	0,02781214
	Glutamyl-tRNA synthetase (GlutamatetRNA ligase)	3.7.2 Aminoacyl-					
gltX	(GluRS)	tRNA synthetases	0,543	1,84162063	-0,881	0,00044697	0,00187712

		3.7.2 Aminoacyl-					
glyQS	Glycyl-tRNA synthetase (GlycinetRNA ligase) (GlyRS)	tRNA synthetases	0,263	3,80228137	-1,926	1,9314E-09	3,3275E-08
		3.7.2 Aminoacyl-					
trpS	Tryptophanyl-tRNA synthetase	tRNA synthetases	0,601	1,66389351	-0,733	0,01114231	0,02995009
_	Methionyl-tRNA synthetase (MethioninetRNA ligase)	3.7.2 Aminoacyl-					
metG	(MetRS)	tRNA synthetases	0,508	1,96850394	-0,977	0,00013296	0,0006642
		3.7.2 Aminoacyl-					
valS	ValyI-tRNA synthetase (ValinetRNA ligase) (ValRS)	tRNA synthetases	0,466	2,14592275	-1,101	4,3426E-05	0,00025136
		3.7.3 Translation					
infC	Translation initiation factor IF-3	initiation	0,505	1,98019802	-0,985	0,00225997	0,00766639
		3.7.4 Translation					
fusA	Elongation factor G (EF-G)	elongation	0,271	3,6900369	-1,886	4,9822E-10	9,6469E-09
		3.7.4 Translation					
tuf	Elongation factor Tu	elongation	0,547	1,82815356	-0,87	0,00731936	0,02098919
	Ribosome recycling factor (Ribosome-releasing factor)	3.7.5 Translation					
frr	(RRF)	termination	0,514	1,94552529	-0,961	0,00035543	0,00155682
		3.8 Protein					
hflC1	Stomatin/prohibitin homolog	modification	0,437	2,28832952	-1,193	6,5556E-07	6,4611E-06
PFCIRM1	dolichyl-phosphate-mannose-protein	3.8 Protein					
38_00200	mannosyltransferase (Glycosyl transferase, family 39)	modification	0,455	2,1978022	-1,135	8,3235E-06	6,2799E-05
		3.8 Protein					
pknB	Serine/threonine protein kinase	modification	0,369	2,7100271	-1,44	2,0423E-07	2,3202E-06
		3.8 Protein					
trxA2	Thioredoxin	modification	0,165	6,06060606	-2,602	2,3871E-06	2,0726E-05
	FK506-binding protein (peptidyl-prolyl cis-trans	3.8 Protein					
fkbP	isomerase)	modification	0,457	2,18818381	-1,129	2,269E-05	0,00014348
ffh	GTP binding signal recognition particle protein	3.9 Protein folding	0,677	1,47710487	-0,564	0,01935583	0,04689984
groL2							
(groEL2)	60 kDa chaperonin 2 (Protein Cpn60 2) (groEL protein 2)						
(hsp60)	(Heat shock protein 60 2)	3.9 Protein folding	0,486	2,05761317	-1,041	0,00348033	0,01090968
groS1(gro							
ES1)	10 kDa chaperonin 1 (Protein Cpn10 1) (groES protein 1)						
(hsp10)	(Heat shock 10 1)	3.9 Protein folding	0,394	2,53807107	-1,342	0,00049046	0,00202096
groL1	60 kDa chaperonin 1 (Protein Cpn60 1) (groEL protein 1)						
(groEL1)	(Heat shock protein 60 1)	3.9 Protein folding	0,304	3,28947368	-1,72	2,8762E-06	2,4504E-05

groS2							
(groES2)	10 kDa chaperonin 2 (Protein Cpn10 2) (groES protein 2)						
(hsp10)	(Heat shock protein 10 2)	3.9 Protein folding	0,603	1,65837479	-0,73	0,00968977	0,02650153
tig	Trigger factor (TF)	3.9 Protein folding	0,527	1,89753321	-0,924	0,00197876	0,0067649
PFCIRM1	Beta-lactamase-like:RNA-metabolising metallo-beta-						
38_01290	lactamase	4.2 Detoxification	0,39	2,56410256	-1,36	3,1801E-07	3,3614E-06
PFCIRM1							
38_07575	beta-lactamase	4.2 Detoxification	0,53	1,88679245	-0,916	0,00488089	0,01476462
pcaC2	4-carboxymuconolactone decarboxylase	4.2 Detoxification	0,613	1,63132137	-0,706	0,0153359	0,03924556
		4.4 Phage-related					
casC	CRISPR system CASCADE complex protein CasC	function	0,573	1,7452007	-0,803	0,0184017	0,04523923
		4.4 Phage-related					
casD	CRISPR system CASCADE complex protein CasD	function	0,526	1,90114068	-0,926	0,00280899	0,00922833
		4.4 Phage-related					
casE	CRISPR system CASCADE complex protein CasE	function	0,504	1,98412698	-0,989	0,00098685	0,00370365
		4.4 Phage-related					
cas1	CRISPR-associated endonuclease Cas1, ECOLI subtype	function	0,469	2,13219616	-1,092	0,00134929	0,00485328
PFCIRM1							
38_02315	Kinase	4.6 Miscellaneous	0,383	2,61096606	-1,385	0,00139952	0,00500353
PFCIRM1							
38_02865	HIT family protein	4.6 Miscellaneous	0,415	2,40963855	-1,27	0,00143203	0,00511138
PFCIRM1							
38_03220	GCN5-related N-acetyltransferase	4.6 Miscellaneous	0,54	1,85185185	-0,889	0,00174795	0,00611924
	S-ribosylhomocysteine lyase (Autoinducer-2 production			4 0000045	0 705	0.00050004	0.04004000
luxS	protein luxS) (AI-2 synthesis protein)	4.6 Miscellaneous	0,614	1,6286645	-0,705	0,02050961	0,04931322
oxaA	Membrane protein insertase	4.6 Miscellaneous	0,374	2,67379679	-1,418	2,1836E-07	2,4253E-06
	Silent information regulator protein Sir2 /NAD-dependent						
npdA	deacetylase	4.6 Miscellaneous	0,446	2,24215247	-1,166	4,0683E-05	0,00023611
PFCIRM1							
38_05780	Fe-S oxidoreductase	4.6 Miscellaneous	0,127	7,87401575	-2,974	3,6151E-16	2,0281E-14
PFCIRM1							
38_05925	Oxidoreductase	4.6 Miscellaneous	0,153	6,53594771	-2,708	1,3825E-15	7,3778E-14
mviN	Conserved membrane protein, MviN-like protein	4.6 Miscellaneous	0,461	2,1691974	-1,118	0,0018332	0,00634657
lipA	Lipoic acid synthetase	4.6 Miscellaneous	0,564	1,77304965	-0,826	0,0006516	0,00258279

PFCIRM1							
38_09940	Von Willebrand factor, type A	4.6 Miscellaneous	0,325	3,07692308	-1,621	4,4755E-06	3,6268E-05
PFCIRM1							
38_10155	PP2Cc, Serine/threonine phosphatases,	4.6 Miscellaneous	0,627	1,59489633	-0,674	0,0196066	0,04740248
PFCIRM1							
38_10615	dehydrogenase/reductase	4.6 Miscellaneous	0,661	1,5128593	-0,597	0,01863681	0,04558667
PFCIRM1							
38_11500	Glycosyltransferase	4.6 Miscellaneous	0,52	1,92307692	-0,944	0,00117522	0,00431439
PFCIRM1							
38_11855	oxidoreductase	4.6 Miscellaneous	0,43	2,3255814	-1,218	3,2795E-05	0,00019605
		5.1 Protein of					
		unknown function					
		similar to other					
PFCIRM1		proteins from the					
38_12335	protein of unknown function	same organism	0,402	2,48756219	-1,315	9,4569E-06	6,9669E-05
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_00015	Hypothetical protein	organisms	0,591	1,69204738	-0,759	0,00762589	0,0217451
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_00185	Hypothetical protein	organisms	0,355	2,81690141	-1,494	3,2154E-09	5,173E-08
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_00660	Hypothetical membrane protein	organisms	0,492	2,03252033	-1,024	0,0004143	0,00176018
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_01150	Hypothetical protein	organisms	0,559	1,78890877	-0,839	0,01546312	0,03952488
PFCIRM1	Hypothetical protein	5.2 Protein of	0,494	2,0242915	-1,018	0,00017924	0,00084337

38_01300		unknown function					
		similar to proteins					
		from other					
		organisms					
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_01745	Hypothetical protein	organisms	0,35	2,85714286	-1,517	2,9818E-06	2,5287E-05
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_01750	Hypothetical protein	organisms	0,309	3,23624595	-1,692	3,4592E-05	0,00020456
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_01755	Hypothetical protein	organisms	0,446	2,24215247	-1,164	1,472E-05	0,00010002
		5.2 Protein of				·	
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 01760	Hypothetical protein	organisms	0,523	1,91204589	-0,935	0,01895944	0,04609251
		5.2 Protein of	,	,	,	,	,
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 02105	Hypothetical protein	organisms	0.618	1.61812298	-0.695	0.01661365	0.04187864
00_02.00		5.2 Protein of	0,010	.,	0,000	0,01001000	0,01101001
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 02305	Hypothetical protein	organisms	0.35	2 85714286	-1 515	0 0028286	0 00927883
PFCIRM1		5.2 Protein of	0,00	2,007 1 1200	.,010	0,0020200	5,00021000
38 02320	LIPE0210 protein	unknown function	0 4 1 5	2 40963855	-1 27	2 0854E-07	2.3399E-06
00_02020			0,410	2,4000000	1,41	2,00040 01	2,00000 00

		similar to proteins					
		organisms					
		5 2 Protein of					
		unknown function					
		similar to proteins					
PECIRM1		from other					
38 02810	Hypothetical protein	organisms	0 484	2 0661157	-1 047	0 00058972	0 0023719
00_02010		5 2 Protein of	0,404	2,0001107	1,047	0,0000012	0,0020710
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 02830	Hypothetical protein	organisms	0.493	2.02839757	-1.021	0.00122682	0.00446635
		5.2 Protein of	-,	_,	.,		-,
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 02950	Hypothetical protein	organisms	0,52	1,92307692	-0,942	0,00133948	0,00482829
		5.2 Protein of			,		
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_03420	Hypothetical protein	organisms	0,5	2	-1,001	0,00015646	0,00075568
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_03690	Hypothetical transmembrane protein	organisms	0,553	1,80831826	-0,854	0,00030891	0,00137099
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_03855	Hypothetical membrane protein	organisms	0,621	1,61030596	-0,686	0,01606597	0,04077997
		5.2 Protein of					
PFCIRM1		unknown function					
38_05525	Hypothetical membrane protein	similar to proteins	0,419	2,38663484	-1,256	1,5751E-06	1,436E-05

		from other					
		organisms					
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_05935	Hypothetical protein	organisms	0,179	5,58659218	-2,484	2,1105E-11	5,1884E-10
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_06015	Hypothetical protein	organisms	0,458	2,18340611	-1,126	0,00014176	0,000697
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_06085	Putative uncharacterized protein	organisms	0,584	1,71232877	-0,776	0,00930584	0,02561156
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 07050	Hypothetical membrane protein	organisms	0,595	1,68067227	-0,748	0,00507038	0,01525997
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 07405	Hypothetical protein	organisms	0.284	3.52112676	-1.818	0.00650729	0.01900928
		5.2 Protein of	-, -	- )	,		-,
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 07760	Hypothetical membrane protein	organisms	0.458	2.18340611	-1.127	0.00012091	0.00061098
		5.2 Protein of	,	,	,	,	,
		unknown function					
PFCIRM1		similar to proteins					
38_08100	Hypothetical protein	from other	0,55	1,81818182	-0,862	0,00227597	0,0076968

		organisms					
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 08555	Hypothetical protein	organisms	0,255	3,92156863	-1,972	0,00065493	0,00259129
		5.2 Protein of				,	
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 08630	Hypothetical protein	organisms	0,566	1,76678445	-0,82	0,01379083	0,0358365
		5.2 Protein of				,	
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 08980	Hypothetical protein	organisms	0,362	2,76243094	-1,466	7,0988E-08	8,629E-07
		5.2 Protein of	,	,	,	,	
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 10000	Hypothetical protein	organisms	0,55	1,81818182	-0,863	0,00155047	0,00548049
		5.2 Protein of				,	
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38 10235	Hypothetical protein	organisms	0,253	3,95256917	-1,98	1,8996E-07	2,1827E-06
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_10240	Hypothetical secreted and membrane protein	organisms	0,244	4,09836066	-2,037	7,8508E-12	2,0948E-10
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_10400	Hypothetical protein	organisms	0,499	2,00400802	-1,004	0,00175393	0,00613034

		5.2 Protein of unknown function					
5501514		similar to proteins					
PFCIRM1		from other					
38_10520	Hypothetical secreted protein	organisms	0,445	2,24719101	-1,167	0,00024499	0,00110979
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_10550	Hypothetical protein	organisms	0,592	1,68918919	-0,757	0,00596512	0,01758988
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_10765	Hypothetical protein	organisms	0,614	1,6286645	-0,703	0,01133687	0,03036117
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_11190	Hypothetical membrane protein	organisms	0,554	1,80505415	-0,851	0,00092164	0,00347082
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_11325	Hypothetical secreted protein	organisms	0,599	1,66944908	-0,74	0,00274834	0,00909738
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_11530	Hypothetical protein	organisms	0,45	2,22222222	-1,152	0,00015461	0,00075007
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_11550	Hypothetical protein	organisms	0,602	1,66112957	-0,731	0,00647959	0,01895366
PFCIRM1	Hypothetical protein	5.2 Protein of	0,484	2,0661157	-1,046	0,00181393	0,0062998
38_11810		unknown function					
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		similar to proteins					
		from other					
		organisms					
		5.2 Protein of					
		unknown function					
		similar to proteins					
PFCIRM1		from other					
38_12275	Hypothetical secreted protein	organisms	0,603	1,65837479	-0,73	0,00454104	0,01387682
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_00435	Hypothetical protein	other proteins	0,429	2,33100233	-1,22	0,00023681	0,0010776
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_01880	Hypothetical protein	other proteins	0,646	1,54798762	-0,631	0,01081575	0,02914391
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_02445	Hypothetical protein	other proteins	0,57	1,75438596	-0,811	0,00335633	0,01059689
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_02765	Protein of unknown function	other proteins	0,286	3,4965035	-1,807	0,00293814	0,00953805
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_04485	Hypothetical protein	other proteins	0,498	2,00803213	-1,006	0,01671466	0,04208477
	, ,	6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38 04545	Hypothetical protein	other proteins	0,445	2,24719101	-1,168	0,00026335	0,00118804
		6 Protein of			•		-
PFCIRM1		unknown function.					
38 05150	Hypothetical protein	without similarity to	0,594	1,68350168	-0,753	0,00693117	0,01995447

		other proteins					
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_05205	Hypothetical protein	other proteins	0,606	1,65016502	-0,721	0,01284854	0,03378918
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_05510	Protein of unknown function	other proteins	0,21	4,76190476	-2,252	0,00157402	0,00555478
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_06570	Hypothetical protein	other proteins	0,338	2,95857988	-1,567	1,0521E-05	7,6224E-05
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_06675	Hypothetical protein	other proteins	0,635	1,57480315	-0,656	0,00665084	0,01928747
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_06720	Hypothetical protein	other proteins	0,576	1,73611111	-0,797	0,00346043	0,01089446
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_07920	protein of unknown function	other proteins	0,579	1,72711572	-0,789	0,01375217	0,03577853
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_08640	Hypothetical protein	other proteins	0,564	1,77304965	-0,826	0,00444145	0,01364873
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_09065	Hypothetical protein	other proteins	0,318	3,14465409	-1,654	1,1699E-08	1,7179E-07
		6 Protein of					
PFCIRM1		unknown function,					
38_10065	Hypothetical protein	without similarity to	0,345	2,89855072	-1,533	3,8435E-06	3,1855E-05

		other proteins					
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_11045	Hypothetical protein	other proteins	0,502	1,99203187	-0,994	5,059E-05	0,00028751
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_11050	Hypothetical protein	other proteins	0,422	2,36966825	-1,244	0,00038207	0,00165211
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_11490	Hypothetical membrane protein	other proteins	0,559	1,78890877	-0,839	0,00296424	0,00958036
		6 Protein of					
		unknown function,					
PFCIRM1		without similarity to					
38_12015	Hypothetical protein	other proteins	0,524	1,90839695	-0,933	0,00112543	0,00416657

 Table S3: RT-qPCR results for validation of RNAseq results.

Rnaseq						RT-qPCR vali							
Induction (I) Repression (R)	Fold	Target	Sample	Mean Ca	Mean Efficiency Corrected Ca	Normalized Expression	Relative Normalized Expression	Fold	Pvalue	Significativity P value <0.05	Forward primer	Reverse primer	Reference
		8	1d post-	-1	-1		<b>F</b> = 0.000				<b>F</b>	<b>P</b>	
			inoculati										
		5930	on	17.1	16.71	23.43029	1	1	N/A			<b>T</b> 0010100	
R	6	5930	inoculat	22.4 1	21.91	5.22024	0.2228	- 4.48 836	0.0210 89	Yes	TGCCGA CCTATGA	GAGTTGTA CGG	this paper
		5930	9d post- inoculati on	23.3 8	22.85	4.28595	0.18292	- 5.46 676	0.0212 04	Yes			
		7015	1d post- inoculati on	24.8	24.9	0.08008	1	1	N/A				
1	49.4	7015	3d post- inoculat ion	23.2 7	23.37	1.89254	23.63262	23.6 3262	0.0001 42	Yes	ACCCCTA CGGAAC GCAAG	GAGCAATC TGGTTGAC GTTGTT	this paper
		7015	9d post- inoculati on	23.7 5	23.86	2.13738	26.69002	26.6 9002	0.0274 67	Yes			
		argG	1d post- inoculati on	22.0 4	21.57	0.80639	1	1	N/A				
R	3.1	argG	3d post- inoculat ion	26.4 4	25.88	0.33294	0.41287	- 2.42 205	0.0076 66	Yes	GACCATC AACGGC AAGAAGT	TCGTAGG CGATGAAC AACAG	Dalmasso 2012 PlosOne
	-		9d post-	27.6				-	0.0403				
		argG	inoculati	7	27.09	0.22701	0.28151	3.55	64	Yes			

			on					23					
		<i></i>	1d post- inoculati	19.7	10.50								
		fabF	on	8	19.59	3.18311	1	1	N/A			<u></u>	
R	6.3	fabF	3d post- inoculat ion	25.9	25.66	0.38881	0.12215	- 8.18 678	0.0254 84	Yes	CTCACAC CTCTCTC	GATGACG CCGATTCC ACT	this paper
		fabF	9d post- inoculati on	26.2 5	26.01	0.48128	0.1512	- 6.61 386	0.0353 1	Yes			
		gadB	1d post- inoculati on	27.5 5	28.26	0.00785	1	1	N/A				
I	5	gadB	3d post- inoculat ion	28.9 6	29.71	0.02349	2.9926	2.99 26	0.0105 01	Yes	ATCTCGG AGGACC ACAAGG	GTACATGC CGGTGTA GGTGA	this paper
		gadB	9d post- inoculati on	24.4 8	25.11	0.89795	114.41965	114. 4196 5	0.0377 62	Yes			
		gtfF	1d post- inoculati on	30.1	27	0.01875	1	1	N/A				
I	6.1	gtfF	3d post- inoculat ion	31.5 7	28.31	0.06173	3.29223	3.29 223	0.0113 46	Yes	CGACGAT TCCGACA AGCCCG AATAC	TGATCTCG TCGCTGTC GAGGATC AC	Deutsch et al. AEM 2010
		gtfF	9d post- inoculati on	35.0 6	31.45	0.01109	0.59131	- 1.69 116	0.4866 05	Not			
		iolT1	1d post- inoculati on	27.3 3	27.76	0.01108	1	1	N/A				
I	11	iolT1	3d post- inoculat ion	29.0 5	29.5	0.02699	2.43566	2.43 566	0.0145 06	Yes	GCTCGAT CCCATTC CCACC	TGTTGGGT GTCTCGG GGT	this paper

			9d post- inoculati	25.1				58.3	0.0192				
		iol 1	on	8	25.58	0.64701	58.38609	8609	36	Yes			
		narH	1d post- inoculati	30.4	28.61	0.00613	1	1	N/A				
	22	narH	3d post- inoculat ion	31.7 1	29.84	0.02142	3.49599	3.49 599	0.0065	Yes	GTGGGA CCTCATC TCCACCT	CGTACCA GACCATC GGCATC	this paper
		narH	9d post- inoculati on	28.1 5	26.49	0.344	56.13897	56.1 3897	0.0758 88	Not			
		sdhC1	1d post- inoculati on	19.1 7	19.49	3.42533	1	1	N/A				
R	8.6	sdhC1	3d post- inoculat ion	25.2 6	25.68	0.38255	0.11168	- 8.95 383	0.0003 78	Yes	CACATGG TCGGAAA TCTCAA	AGATCCAC AGACCAC CCTCA	this paper
		sdhC1	9d post- inoculati	32.3 8	32 92	0.004	0.00117	- 856. 9302 6	N/A	Not			
		slgT	1d post- inoculati on	22.8	21.9	0.64339	1	1	N/A				
R	4	slgT	3d post- inoculat ion	28.7 6	27.62	0.09946	0.15458	- 6.46 897	0.0000 94	Yes	TCGGATA CTGGAC CACCAA	ACGATGAA GGGGATG AACAG	this paper
		slgT	9d post- inoculati on	33.3 8	32.06	0.00723	0.01124	- 88.9 9659	0.0000 1	Yes			
		slpE F2R2	1d post- inoculati on	27.7 1	26.9	0.02012	1	1	N/A				
1	12.9	slpE F2R2	3d post- inoculat	28.3	27.47	0.1102	5.47841	5.47 841	0.0177 65	Yes	ATCGTGA TGCGTTG	CATCCGA GAACGTCT	this paper

			ion								ATTGCCT	GACCC	
		slpE F2R2	9d post- inoculati on	32.7 6	31.8	0.00867	0.43103	- 2.32 005	0.4845	Not			
		mmda	1d post- inoculati on	17.6 7	16.84	21.49449	1	1	N/A				
R	2.4	mmda	3d post- inoculat ion	21.9 3	20.91	10.46725	0.48697	- 2.05 35	0.0050	Yes	CGCGTC GAGAAG CAACATT C	AACGAATG GGGATCG AGCAG	this paper
		mmda	9d post- inoculati on	22.4 1	21.36	12.07668	0.56185	- 1.77 983	0.4506 15	Not			
		dadA2	1d post- inoculati on	28.2	27.77	0.01098	1	1	N/A				
1	13	dadA2	3d post- inoculat ion	30.5	30.03	0.0187	1.70259	1.70 259	0.1238 29	Not	CAAGTCG GAGTTCG TCAAGG	GCCCTGTT CGTGCATC TC	this paper
		dadA2	9d post- inoculati on	28.5 7	28.14	0.10988	10.00451	10.0 0451	0.0864 16	Not			
normalizati											CGCGGA ACTGCTC	CACCTCGT CCTTCGAG	Dalmasso 2012
on genes		sdaA								stable	AATATCT TACGAGT TCGAGG	TAGC AAACCAGC GGGTAGT	PlosOne Dalmasso 2012
		eno1								stable	GCAAGTC CCTACAC	CATTG	PlosOne
		cstA								stable	ACCGGA GGAAGA G	GGACCGT CCATTTGT TATGC	Dalmasso 2012 PlosOne

# CAPÍTULO 3: Discussão geral, conclusão e perspectivas

### **CHAPITRE 3 : Discussion générale,** conclusion et perspectives

#### Discussão geral

O objetivo geral desta tese foi de melhorar a nossa compreensão dos mecanismos envolvidos no fenômeno sobrevivência a longo prazo em *P. freudenreichii*. Uma das principais características da espécie é, de fato, uma boa sobrevivência a longo prazo, e este é um critério favorável à sua utilização em vários setores da indústria, tais como: a produção de queijos PPC, especialmente o Emmental, um dos produtos mais consumidos na França (Dalmasso *et al.* 2012a), e o desenvolvimento de produtos probióticos com efeitos benéficos para a saúde humana (Cousin *et al.* 2012). A síntese de biomoléculas, tais como ácido propiônico e vitamina B12 (Van Wyk, Witthuhn and Britz 2011; Wang and Yang 2013) é uma outra utilização industrial importante para esta espécie. Mas esta última é realizada sob condições de cultura altamente controlados para manter uma atividade metabólica ótima e a problemática de sobrevivência a longo prazo não está no escopo destas aplicações.

Neste trabalho, mostramos, através da análise do perfil de expressão de genes e da produção e consumação de metabólitos, que *P. freudenreichii* é capaz de reduzir o seu metabolismo, colocar-se em um estado de dormência e utilizar vias alternativas de energia. Estes resultados serão discutidos nos parágrafos seguintes.

### > Caracterização fenotípica de *P. freudenreichii*:

Estudos anteriores relatam que *P. freudenreichii* é uma bactéria versátil que sobrevive a vários estresses ambientais (Jan, Rouault and Maubois 2000; Dalmasso *et al.* 2012b; Saraoui *et al.* 2013b). Além disso, *P. freudenreichii* é capaz de utilizar diferentes fontes de carbono como substrato e pode armazenar certos compostos para a sobrevivência a longo prazo (Thierry *et al.* 2011). Para uma melhor compreensão desta bactéria, nós realizamos a caracterização fenotípica de 8 linhagens de *P. freudenreichii*, durante um período de carência nutricional. Sabendp-se que, em condições ótimas, esta bactéria tem um tempo de geração de 5h (Falentin *et al.* 2010a), assumiu-se que um período de 11 dias (com uma 1 amostragem por dia) durante o qual a bactéria está sujeita ao estresse nutricional permitiu separar e caracterizar bem as diferentes fases de crescimento, até uma fase de sobrevivência a longo prazo. Testes de sobrevivência a longo prazo já foram realizados em outras bactérias indicando uma caracterização num tempo, significativamente, mais longo, como por exemplo, Finkel e Kolter que consideraram uma duração de um ano para a avaliação da sobrevivência em *Escherichia coli* (Finkel 2006b). Com certeza seria interessante fazer análises mais longas, mas um

dos objetivos desta tese foi de tentar elucidar os mecanismos que permitiram a adaptação de *P. freudenreichii* à esta fase de crescimento.

A capacidade de sobrevivência a longo prazo varia em função do género, da espécie ou mesmo da linhagem estudada. Os estudos fenotípicos de 8 linhagens demonstraram que o conjunto de todas as linhagens de *P. freudenreichii* apresentaram uma boa sobrevivência a longo prazo, no entanto, com algumas variações entre as linhagens. Estudar este fenômeno durante um período de 6 ou 12 meses, nos teria permitido de determinar durante quanto tempo era possível a esta bactéria de sobreviver em carência nutricional e, sobretudo, poderia ter revelado e ou enfatizado as diferenças entre as linhagens durante a sobrevivência a longo prazo. Esta capacidade poderia de fato tornar-se um critério para a seleção de linhagens para aplicações em queijos ou probióticas de *P. freudenreichii*.

### Dormência em P. freudenreichii:

Muitas bactérias são capazes de entrar em um estado de dormência permitindo uma maior resistência e uma melhor sobrevivência a ambientes desfavoráveis (Rittershaus, Baek and Sassetti 2013b). Esta adaptação é caracterizada pela manutenção da viabilidade, no entanto, associada a uma perda de cultivabilidade em condições padrão laboratoriais. Este estado também é descrito pelo termo de estado VBNC (Viable But Non Culturable). O estado VBNC constitui uma estratégia muito importante para algumas espécies bacterianas, Gram positivas ou negativas, para a sobrevivência durante períodos prolongados de carência. Muitos estudos científicos têm sido dedicados a este tema. Uma busca PubMed mostra que o termo "viable but nonculturable bacteria" totaliza em 385 resultados. Dentre eles, 64 artigos sobre bactérias patogénicas ("VBNC" AND "pathogen") e 78 artigos sobre bactérias em estado VBNC em alimentos ou na água ("VBNC" AND "food"). Nesta última pesquisa, a maioria dos artigos tratavam-se patógenos alimentares (Campylobacter sp., E. coli, Vibrio sp., Salmonella sp.) e alguns poucos, eventualmente, sobre as bactérias de interesse alimentar (food grade). Três artigos relacionados com probióticos. Esta diferença de tratamento ou de interesse entre estas áreas é em parte explicado pelo fato de que muitas bactérias patogênicas são capazes de induzir a mesma infecção ou intoxicação mesmo estando em um estado VBNC, e geralmente, essas bactérias (não cultiváveis) não são detectadas por técnicas microbiológicas convencionais, como a contagem das UFC. Os estudos sobre probióticos são realizados para desenvolver a sua

importância na formulação destes produtos, uma vez que o número de bactérias viáveis no produto final é um critério levado em consideração para os efeitos probióticos sobre o hospedeiro. Em um estudo realizado por Davis e colaboradores, demonstrou-se que as bactérias que haviam propriedades probióticas entravam em estado VBNC e que elas só podiam ser quantificadas corretamente através de métodos alternativos, tais como hibridação *in situ* fluorescente, qPCR, RT-PCR, propídio monoazida-PCR e/ou a separação de células ativadas por fluorescência (Davis 2014). Desta mesma maneira, confirmamos o estado VBNC em *P. freudenreichii* CIRM-BIA138, linhagem mais resistente a carência nutricional, combinando diferentes metodologias, tais como a contagem de UFC, a microscopia de epi-fluorescência e qPCR.

# Genes diferencialmente expressos em fases de crescimento exponencial e estacionária de *P. freudenreichii*:

Para aprofundar os estudos relacionados com a sobrevivência a longo prazo desta bactéria, foi realizada a análise do perfil transcricional da linhagem P. freudenreichii CIRM- BIA138 utilizando a técnica de RNA-Seq, durante um período de carência nutricional. Inicialmente, escolhemos três pontos de crescimento bacteriano para realizar esta análise: exponencial, estacionária e estacionária tardia. Para obter estas amostras, realizamos a cultura bacteriana, a extração de RNA total e a depleção de RNAr (RNA ribossômico). Em seguida, sequenciamos o cDNA destas amostras pela plataforma Illumina HiSeq 2000, uma tecnologia de sequenciamento de próxima geração. Infelizmente, observa-se que não foi possível obter bons resultados de sequenciamento nas amostras da fase estacionária tardia. Isto, porque os resultados das 3 replicatas desta última fase apresentaram um perfil de contaminação por DNA genômico, uma vez que as reads estavam alinhadas de maneira uniforme e nãoorientada, impedindo assim a interpretação dos dados de sequenciamento. No entanto, a análise de RNA-Seq entre as fases de crescimento bacteriano exponencial e estacionária nos permitiu identificar 912 genes diferencialmente expressos e potencialmente envolvidos na preparação e adaptação da linhagem à carência nutricional.

Atualmente, um dos poucos estudos de transcriptoma sobre o tema de sobrevivência a longo prazo é o realizado sobre o patógeno *Listeria monocytogenes* que provoca infecções de origem alimentar. A cultura de *L. monocytogenes* foi amostrada em diferentes fases do desenvolvimento (13h, 17h, 24h e 168h, 336h), num meio de cultura de triptona de soja ágar com extrato de levedura (Becton Dickinson, MD). Este

estudo foi realizado por meio de técnicas de microarranjo e RT-qPCR que revelaram genes induzidos envolvidos em: (i) síntese de aminoácidos, de ácidos graxos, de proteínas, de fosfolípidos, de envelope celular e de ribonucleótideos ; (ii) na transcrição; (iii) na desintoxicação; (iv) no transporte de proteínas; e (v) na divisão celular. Estes metabolismos altamente expressos durante a fase logarítmica são reprimidos durante o período de sobrevivência a longo prazo (Wen *et al.* 2011).

Observou-se ainda que os genes envolvidos na síntese de proteínas foram reprimidos, o que indica também, que esta bactéria, entra em estado de dormência após períodos prolongados (Wen *et al.* 2011). Apesar de *L. monocytogenes* ser patogênica, a estratégia de sobrevivência adotada é semelhante a adotada por *P. freudenreichii*. Os resultados do transcriptoma de *P. freudenreichii* CIRM-BIA138 também revelaram que, na fase estacionária, a bactérias começa a se preparar para um estado de dormência que lhe permite sobreviver e de utilizar fontes alternativas para obter e armazenar a energia.

Além disso, os nossos resultados demonstram uma redução no metabolismo de *P. freudenreichii*, uma vez que os genes envolvidos na fosforilação oxidativa e no ciclo de Wood-Werkman estão reprimidos durante a fase estacionária. Estes resultados estão de acordo com o trabalho de Saraoui e colaboradores que mostram que, em *P. freudenreichii* CIRM-BIA1, as vias da glicólise, do ciclo de Wood-Werkman e fosforilação oxidativa foram reprimidos em fase estacionária no cólon de suínos (Saraoui *et al.* 2013a).

### Mecanismos moleculares envolvidos na sobrevivência a longo prazo de P. freudenreichii:

Através de dados transcriptômicos e bioquímicos de *P. freudenreichii* CIRM-BIA138, obtivemos pistas que indicam quais genes estão ativos nesta bactéria e quais são os fatores ambientais envolvidos no desencadeamento de mecanismos moleculares que preparam a sobrevivência bacteriana a longo prazo, durante um período de carência nutricional.

Durante este período, a linhagem *P. freudenreichii* CIRM-BIA138 é também capaz de explorar novas fontes de energia, tais como aminoácidos (como o glutamato, glicina e alanina), a arabinose e o inositol que são fontes de carbono alternativas. Para equilibrar o seu metabolismo, na ausência de oxigénio, esta bactéria utiliza provavelmente o polifosfato e o pirofosfato para armazenar a energia, os quais os genes estão presentes no genoma desta espécie (Falentin *et al.* 2010a). Assim, neste trabalho,

os genes envolvidos na síntese de polifosfatos são induzidos durante a fase estacionária para a utilização de ATP. Além disso, nesta última fase, uma diminuição na atividade de transcrição, tradução e secreção de proteínas foi observada. Outros parâmetros experimentais também foram analisados neste estudo, como a dosagem de compostos bioquímicos, demonstrando a influência destes últimos sobre o metabolismo de *P. freudenreichii* CIRM-BIA138, durante a carência nutricional. Alterações na síntese e no consumo de ácidos orgânicos, de açúcares e de aminoácidos também foram observados. Isto demonstra a capacidade de *P. freudenreichii* CIRM-BIA138 se adaptar ao ambiente, e contribui para uma melhor compreensão do conhecimento biológico desta espécie.

Todos estes dados fenotípicos, transcriptômicos e metabolômicos de P. freudenreichii CIRM-BIA138 são de grande importância para a compreensão desta bactéria, especialmente, durante as fases importantes encontradas, como por exemplo, durante a maturação de queijos PPC. No entanto, nas condições de maturação, P. freudenreichii atinge rapidamente uma fase estacionária de crescimento, mas raramente associada a uma carência de carbono. Este tipo de trabalho já havia sido começado na equipe, portanto com análise transcricional por microarranjo para identificar estratégias de adaptação de P. freudentreichii nas condições de fabricação de queijos (Dalmasso et al. 2012a). Nossos resultados de RNA-Seq vieram para complementar estes dados. Além disso, nosso estudo metabolômico foi muito direcionado a determinados compostos. Seria interessante fazer um estudo metabolômico mais abrangentes como o trabalho recente de Le Boucher (Le Boucher et al. 2013). Este último trabalho demonstrou a capacidade da espectrometria de massa – metabolic fingerprinting – a detectar alterações induzidas pelo metabolismo bacteriano no queijo durante 0, 8 e 48 h. Este estudo encontrou variações em 45 metabólitos, tais como aminoácidos, vitaminas e metabólitos voláteis (Le Boucher et al. 2013).

A interdisciplinaridade e os diferentes métodos aplicados neste trabalho de tese permitiram de identificar mecanismos que poderiam explicar o fenômeno de sobrevivência a longo prazo nesta bactéria. Uma destas estratégias de sobrevivência parece ser o uso de vias alternativas de energia para manter a atividade metabólica. Nestas etapas, a bactéria se prepara a entrar em um estado de dormência para enfrentar um período de estresse nutricional e sobreviver a longo prazo, enquanto aguarda a ocorrência de condições mais favoráveis.

#### **Discussion générale**

L'objectif général de cette thèse était d'améliorer notre compréhension des mécanismes impliqués dans le phénomène de survie à long terme chez *P. freudenreichii*. Une des principales caractéristiques de l'espèce est, en effet, sa bonne survie à long terme et ceci est un critère favorable à son utilisation dans divers secteurs de l'industrie tels que : la production des fromages PPC, en particulier l'Emmental, l'un des produits les plus consommés en France (Dalmasso *et al.* 2012a) et le développement de produits probiotiques ayant des effets bénéfiques sur la santé humaine (Cousin *et al.* 2012). La synthèse de biomolécules, comme l'acide propionique et la vitamine B12 (Van Wyk, Witthuhn and Britz 2011; Wang and Yang 2013) est une autre utilisation industrielle d'importance pour cette espèce. Mais celle-ci s'effectue dans des conditions de culture hautement contrôlées permettant de maintenir une activité métabolique optimale et la problématique de survie à long terme est de fait hors champs pour ces applications.

Dans ce travail, nous avons montré, à travers l'analyse du profil d'expression des gènes et de la production et consommation de métabolites, que *P. freudenreichii* est capable de réduire son métabolisme, de se mettre dans un état de dormance et d'utiliser des voies d'énergie alternatives. Ces résultats seront discutés dans les paragraphes suivants.

### > Caractérisation phénotypique de P. freudenreichii:

Des études antérieures rapportent que *P. freudenreichii* est une bactérie versatile qui survit à divers stress environnementaux (Jan, Rouault and Maubois 2000; Dalmasso *et al.* 2012a; Saraoui *et al.* 2013a). En outre, *P. freudenreichii* est capable d'utiliser différentes sources de carbone comme substrat et peut stocker certains composés pour sa survie à long terme (Thierry *et al.* 2011). Aussi, pour une meilleure compréhension de cette bactérie, nous avons effectué la caractérisation phénotypique de 8 souches de *P. freudenreichii*, au cours d'une période de carence nutritionnelle. Sachant que, dans des conditions optimales, cette bactérie a un temps de génération de 5h (Falentin *et al.* 2010b), nous avons supposé qu'une période de 11 jours (avec un 1 échantillonnage par jour) durant laquelle la bactérie est soumise au stress nutritionnel permettait de séparer et bien caractériser les différentes phases de croissance, jusqu'à une phase de survie à long terme. Des essais de survie à long terme avaient déjà été réalisés chez d'autres bactéries indiquant une caractérisation sur un temps nettement plus long, comme par example, Finkel et Kolter qui ont consideré une durée d'un an pour l'évaluation de la survie chez *Escherichia coli* (Finkel 2006b). Il serait sûrement intéressant de disposer d'essais plus longs, mais un des buts de cette thèse était de tenter d'élucider les mécanismes qui permettait à l'adaptation de *P. freudenreichii* à cette phase de croissance.

La capacité de survie à long terme varie en fonction du genre, de l'espèce, voire de la souche étudiée. Les études phénotypiques sur 8 souches ont montré que l'ensemble des souches de *P. freudenreichii* présentait une bonne survie à long terme avec cependant quelques variations inter-souches. Etudier ce phénomène sur une durée de 6 ou 12 mois, nous aurait permis de déterminer pendant combien de temps encore il était possible à cette bactérie de survivre en carence nutritionnelle et, surtout, aurait peut-être permis de révéler et ou accentuer des différences inter-souches en matière de survive à long terme. Cette capacité pourrait en effet devenir un critère de sélection de souches pour les applications fromagères ou probiotiques de *P. freudenreichii*.

#### ➢ La dormance chez P. freudenreichii :

De nombreuses bactéries sont capables d'entrer dans un état de dormance permettant une plus grande résistance et une meilleure survie à des environnements défavorables (Rittershaus, Baek and Sassetti 2013b). Cette adaptation se caractérise par le maintien de la viabilité associée cependant à une perte de cultivabilité en conditions de laboratoire classiques. Cet état est également décrit sous le terme d'état VBNC (pour Viable But Non Culturable). L'état VBNC constitue une stratégie très importante chez certaines espèces bactériennes, à Gram positif ou négatif, pour la survie pendant de longues périodes de carence. De nombreuses études scientifiques lui ont été consacrées. Une recherche PubMed montre que le terme « viable but nonculturable bacteria » totalise 385 résultats. Parmi eux, 64 articles concernent les bactéries pathogènes (« VBNC » AND « pathogen ») et 78 articles portent sur les bactéries en état VBNC dans l'aliment ou l'eau (« VBNC » AND « food »). Dans cette dernière recherche, la majorité des articles ont trait aux pathogènes alimentaires (Campylobacter sp., E. coli, Vibrio sp., Salmonella sp.) et très peu, finalement, portent sur des bactéries d'intérêt alimentaire (food grade). Trois articles ont trait aux probiotiques. Cette différence de traitement ou d'intérêt entre ces divers domaines s'explique entre autres par le fait que beaucoup de bactéries pathogènes sont capables d'induire une infection ou une intoxication même à partir d'un état VBNC, et généralement ces bactéries (non cultivables) ne sont pas détectées par des techniques de microbiologie classique, telles que le comptage des UFC. Les études portant sur les probiotiques sont amenées à se développer de par leur importance pour la formulation de ces produits, car le nombre de bactéries viables dans le produit final est un critère pris en compte pour l'effet probiotique sur l'hôte. Dans une étude de Davis et collaborateurs, il a été démontré que les bactéries ayant des propriétés probiotiques se mettaient en état VBNC et qu'elles ne pouvaient être quantifiées correctement qu'à travers des méthodes alternatives comme l'hybridation *in situ* en fluorescence, la PCRq, la RT-PCR, le propidium monoazide-PCR et/ou la séparation des cellules activées par fluorescence (Davis 2014). De même, nous avons confirmé l'état VBNC chez *P. freudenreichii* CIRM-BIA138, la souche la plus résistante à la carence nutritionnelle, en combinant différentes méthodologies telles que la numération des UFC, la microscopie à épi-fluorescence et la PCRq.

### Gènes différentiellement exprimés dans les phases de croissance exponentielle et stationnaire de *P. freudenreichii* :

Pour approfondir les études liées à la survie à long terme de cette bactérie, nous avons effectué l'analyse du profil transcriptionnel de la souche P. freudenreichii CIRM-BIA138 en utilisant la technique de RNA-Seq, au cours d'une période de carence nutritionnelle. Tout d'abord, nous avons choisi trois points de la croissance bactérienne pour effectuer cette analyse : exponentiel, stationnaire et stationnaire tardif. Pour obtenir ces échantillons, nous avons réalisé une culture bactérienne, une extraction des ARN totaux et une déplétion des ARNr (ARN ribosomiques). Ensuite, nous avons séquencé le cDNA de ces échantillons par la plateforme Illumina HiSeq 2000, une technologie de séquençage de nouvelle génération. Malheureusement, nous avons observé qu'il n'était pas possible d'obtenir de bons résultats de séquençage sur les échantillons de la phase stationnaire tardive. En effet, les résultats des 3 replicates de cette dernière phase ont présenté un profil de contamination par de l'ADN génomique, car les reads se sont alignés de manière uniforme et non orientée, empêchant ainsi l'interprétation des résultats de séquençage. Cependant, l'analyse du RNA-Seq entre les phases de croissance bactérienne exponentielle et stationnaire nous a permis d'identifier 912 gènes différentiellement exprimés et potentiellement associés à la préparation et l'adaptation de la souche à la carence nutritionnelle.

Actuellement, l'une des rares études de transcriptome sur le thème de la survie à long terme est celle menée sur le pathogène *Listeria monocytogenes* qui provoque des

infections d'origine alimentaire. La culture de *L. monocytogenes* a été échantillonnée à différentes phases de développement (13h, 17h, 24h et 168h, 336h), dans un milieu de culture gélose tryptone soja avec de l'extrait de levure (Becton Dickinson, MD). Cette étude a été réalisée à travers des techniques utilisant des puces à ADN et la RT-qPCR qui ont révélé des gènes inductibles impliqués dans : (i) la synthèse d'acides aminés, d'acides gras, de protéines, de phospholipides, de l'enveloppe cellulaire et de ribonucléotides; (ii) la transcription ; (iii) la détoxification ; (iv) le transport de protéines ; et (v) la division cellulaire. Ces métabolismes fortement exprimés pendant la phase logarithmique sont réprirmés pendant la période de survie à long terme (Wen *et al.* 2011).

Il a également été observé que les gènes impliqués dans la synthèse des protéines ont été réprimés, ce qui indique aussi, chez cette bactérie, l'entrée en état de dormance après des périodes prolongées (Wen *et al.* 2011). Bien que *L. monocytogenes* soit pathogène, la stratégie de survie qu'elle adopte est similaire à celle adoptée par *P. freudenreichii*. Les résultats du transcriptome de *P. freudenreichii* CIRM-BIA138 ont également révélé que, dans la phase stationnaire, la bactérie commence à se préparer pour un état de dormance qui lui permet de survivre et d'utiliser des sources alternatives pour obtenir et stocker de l'énergie.

En outre, nos résultats démontrent une réduction du métabolisme de *P. freudenreichii*, puisque les gènes impliqués dans la phosphorylation oxydative et le cycle Wood-Werkman sont réprimés pendant la phase stationnaire. Ces résultats sont en accord avec le travail de Saraoui et collaborateurs qui montrent que, chez *P. freudenreichii* CIRM-BIA1, les voies de glycolyse, du cycle de Wood-Werkman et de la phosphorylation oxydative étaient régulé négativement en phase stationnaire dans un côlon de cochons (Saraoui *et al.* 2013a).

### Mécanismes moléculaires impliqués dans la survie à long terme de P. freudenreichii :

À travers des données transcriptomiques et biochimiques de *P. freudenreichii* CIRM-BIA138, nous avons obtenu des pistes indiquant quels gènes sont actifs dans cette bactérie et quels sont les facteurs environmentaux impliqués dans le déclenchement des mécanismes moléculaires qui préparent la survie bactérienne à long terme, au cours d'une période de carence nutritionnelle.

Au cours de cette période, la souche P. freudenreichii CIRM-BIA138 est aussi capable d'explorer de nouvelles sources d'énergie, telles que des acides aminés (comme le glutamate, la glycine et l'alanine), l'arabinose et l'inositol qui sont des sources de carbone alternatives. Pour équilibrer son métabolisme, en l'absence d'oxygène, cette bactérie utilise probablement le polyphosphate et pyrophosphate pour stockér l'énergie et qui les gènes sont présentes dans le génome de cette espèce (Falentin et al. 2010b). Ainsi dans ce travail, les gènes impliqués dans la synthèse de polyphosphates sont induits au cours de la phase stationnaire pour utiliser l'ATP. En outre, dans cette dernière phase, une diminution de l'activité de la transcription, de la traduction et de la sécrétion des protéines a été observée. D'autres paramètres expérimentaux ont aussi été analysés dans ce travail, comme le dosage des composés biochimiques, montrant l'influence de ces derniers sur le métabolisme de P. freudenreichii CIRM-BIA138, au cours de la carence nutritionnelle. Des altérations dans la synthèse et la consommation d'acides organiques, de sucres et d'acides aminés ont également été observés. Cela démontre la capacité de P. freudenreichii CIRM-BIA138 à s'adapter à l'environnement, et contribue à une meilleure compréhension de la connaissance biologique de cette espèce.

Toutes ces données phénotypiques, transcriptomiques et métabolomiques de P. freudenreichii CIRM-BIA138 sont d'une grande importance pour la compréhension de cette bactérie notamment pendant des étapes importantes rencontrées, par exemple, au cours de l'affinage des fromages PPC. Toutefois, dans les conditions d'affinage, P. freudenreichii atteint rapidement une phase stationnaire de croissance mais rarement liée à une carence carbone. Ce type de travail avait déjà commencé dans l'équipe lors de l'analyse transcriptionnelle par puce à ADN pour identifier les stratégies d'adaptation de P. freudentreichii dans les conditions de fabrication fromagère (Dalmasso et al. 2012a). Nos résultats RNA-Seq viennent complétés ces données. En outre, notre étude métabolomique était très ciblée sur certains composés. Il serait intéressant de faire une étude métabolomique plus exhaustive à l'instar des travaux récemment menés par Le Boucher (Le Boucher et al. 2013). Ce dernier travail a montré la capacité de la spectrométrie de masse - *metabolic fingerprinting* – à détecter les modifications induites par le métabolisme bactérien dans le fromage au cours de 0, 8 et 48 h. Cette étude a révélé des variations sur 45 métabolites, tels que les acides aminés, les métabolites volatils et les vitamines (Le Boucher et al. 2013).

L'interdisciplinarité et les différentes méthodes appliquées dans ce travail de thèse ont permis d'identifier des mécanismes qui pourraient expliquer le phénomène de survie à long terme de cette bactérie. Une de ces stratégies de survie semble être l'utilisation de voies énergétiques alternatives permettant le maintien de l'activité métabolique. Dans ces étapes, la bactérie se prépare à entrer en dormance pour affronter une période de stress nutritionnel et survivre à long terme, en attendant la survenue de conditions plus favorables.

**Conclusão/ Conclusion** 

### Conclusão

Neste presente trabalho de tese, apresentamos, pela primeira vez, a identificação de estratégias de sobrevivência a longo-prazo em *P. freudenreichii*, combinando as abordagens de microbiologia, genômica e transcriptômica. Desde a caracterização fenotípica à analise funcional do genoma de *P. freudenreichii*, nossos estudos demonstraram que esta bactéria diversifica suas fontes de energia, em seguida, diminuem o seu metabolismo energético para entrar em um estado de dormência, o qual elas são capazes de sobreviver por períodos prolongados. Além disso, este trabalho de tese fornece informações sobre a biologia do grupo Propionibacteria lactéos.

### Conclusion

Dans ce travail de thèse, nous avons présenté, pour la première fois, l'identification de stratégies de survie à long terme chez *P. freudenreichii*, en combinant des approches microbiologique, génomique et transcriptiomique. De la caractérisation phénotypique à l'analyse fonctionnelle du génome de *P. freudenreichii*, nos études ont permis de démontrer que cette bactérie diversifie ses sources d'énergie, puis diminue son métabolisme énergétique pour entrer dans un état de dormance pendant lequel elle est capable de survivre durant de longues périodes. En outre, ce travail de thèse fournit des informations sur la biologie du groupe de Propionibacteries laitières.

**Perspectivas/ Perspectives** 

### Perspectivas

Os resultados obtidos ao longo deste trabalho de tese nos permitiram a melhor compreender o fenômeno de sobrevivência a longo prazo em *P. freudenreichii*. Eles abrem várias perspectivas. De um modo, os dados gerados podem ser completados e explorados para melhorar o nosso conhecimento da genômica de *P. freudenreichii* (tópicos 1 e 2 abaixo). E de outro modo, ainda há muito a ser feito na análise de sobrevivência a longo prazo e diversas pistas poderiam ser utilizadas para completar este trabalho (tópicos 3-7).

### (I) Sequência completa do genoma de P. freudenreichii CIRM-BIA138:

Como uma primeira abordagem, podemos considerar que um genoma completo (isto é, sem *gaps*) não é essencial, se podemos extrair as informações necessárias das análises de transcriptômica e metabolômica desta bactéria. No entanto, um genoma completamente montado permite uma melhor anotação e, assim, nos dará uma melhor qualidade de informação genética sobre o organismo.

Para obter o genoma completo de *P. freudenreichii* CIRM-BIA138, uma opção seria realizar um mapeamento óptico do genoma desta linhagem. Isto permitiria a superação de dificuldades encontradas na etapa de montagem deste genoma, como por exemplo, a presença de uma grande quantidade de transposases. O mapeamento óptico de fato dá uma visão geral do genoma, auxiliando a reorganização de *contigs* e ao fechamento de *gaps*. Além disso, é possível combinar diferentes programas de montagem para melhorar o processo como um todo. De outra maneira, como já foi demonstrado em estudos precedentes (Garber *et al.* 2011), e para complementar as abordagens anteriores, é possível utilizar os dados do transcriptoma provenientes do RNA-Seq para alinhar os transcritos no genoma e fechar os *gaps*. Os dados de RNA-Seq também podem auxiliar no processo de anotação do genoma, utilizando programas como o ReadXplorer, que permite o alinhamento dos dados de expressão de cada gene sob o genoma para refinar a anotação (Hilker *et al.* 2014).

### (II) Identificação de novos elementos no genoma de *P. freudenreichii* CIRM-BIA 138:

Os dados de RNA-Seq permitem a descoberta de novos transcritos que não foram identificados e anotados em versões anteriores do genoma. Uma análise detalhada

permitirá também de identificar pequenos RNAs não-codificantes em *P. freudenreichii* CIRM-BIA138. A tecnologia de RNA-Seq revelou-se eficaz para a detecção de pequenos RNAs em *Pseudomonas aeruginosa* (Gómez-Lozano *et al.* 2012), e para a descoberta de novos transcritos em várias espécies bacterianas (McClure *et al.* 2013). Programas de bioinformática, como Rockhopper, através de uma interface gráfica amigável, permite que o usuário entre com genomas de referência e modifique as configurações, tais como: leituras *paired-end* ou *mate-pair*; alinhamento de leituras de várias replicatas; número de replicatas e condições experimentais diferentes utilizadas para a análise de RNA-Seq (McClure *et al.* 2013).

# (III) Complemetação dos dados de RNA-Seq na fase estacionária tardia de *P. freudenreichii* CIRM-BIA138:

Durante a análise de RNA-Seq da linhagem *P. freudenreichii* CIRM-BIA138, a quantidade de RNA extraída de amostras associadas com a fase de sobrevivência a longo prazo foi muito baixa para permitir um sequenciamento de qualidade. Além disso, as amostras da fase estacionária tardia foram contaminadas com DNA genómico. Isto limitou severamente os resultados obtidos. Por conseguinte, seria vantajoso obter uma maior quantidade de RNA e a melhorar a sua qualidade, aumentando o volume da cultura bacteriana amostrada (por exemplo 25 mL) visando melhorar, a partir disso, as próximas etapas de extração e de depleção dos rRNA certificando-se de remover o DNA genômico por tratamentos com DNase.

### (IV) Mecanismo de dormência na espécie P. freudenreichii:

Muitas bactérias Gram-positivas e Gram-negativas entram em dormência na fase de sobrevivência a longo prazo. Como vimos, dentre as 8 linhagens de *P. freudenreichii* testadas, todas sobrevivem bem e suas populações permanecem em níveis elevados durante o período de carência nutricional. No entanto, observamos que a linhagem CIRM-BIA138 apresenta um nível de sobrevivência maior do que outras linhagens. Esta linhagem foi objeto de uma análise mais aprofundada (demonstração do estado VBNC e RNA-Seq). Neste contexto, seria interessante complementar os resultados obtidos nesta tese, explorando a capacidade de sobrevivência a longo prazo e o estado VBNC em outras linhagens. Isso talvez permita identificar linhagens de *P. freudenreichii* com baixa sobrevivência a longo prazo, e em seguida, por genômica comparativa os genes envolvidos (por exemplo, pela sua ausência ou por serem pseudogenes) nesta sobrevivência inferior. Do mesmo modo, outras formas de stress (por exemplo, calor, ácido ou osmótico) podem ser aplicados a culturas em estado de sobrevivência a longo prazo e carência nutricional. Isto permitiria a identificar os fenômenos de adaptação e resistência cruzada que poderiam nos esclarecer sobre os mecanismos implementados e nos fornecer uma melhor explicação do comportamento da espécie em condições de fabricação de queijos.

Além disso, este estudo pode ser complementado pela construção de mutantes visando a interromper os genes envolvidos no desencadeamento da dormência em *P. freudenreichii*. Trabalho abordando esta metodologia (construção de mutantes) já foi realizado para mostrar o envolvimento do gene *clpX* em *S. typhimurium* (Kusumoto, Miyashita and Kawamoto 2013) e *rpo*S em *E. coli* (Boaretti *et al.* 2003) na sobrevivência a longo prazo nestas espécies.

### (V) Reanimação na espécie P. freudenreichii:

Ao sair do estado VBNC, muitas espécies bacterianas passam por um estado chamado de ressuscitação permitindo-lhes retomar o crescimento normal. A reanimação (ou ressuscitação), por vezes, requer condições especiais de crescimento. Da mesma forma, alguns genes parecem estar diretamente envolvidos neste fenômeno. Assim, em *M. luteus*, tem sido mostrado que as células são reanimadas depois da produção do fator Rpf (*Ressuscitation Promoting Factor*) (Greenblatt *et al.* 2004a). A investigação deste estado de reanimação nas células de *P. freudenreichii* seria de grande interesse para a conclusão deste trabalho. No entanto, é necessário realizar experimentos adicionais que comprovem que as células são ressuscitadas após o estado VBNC. Isto requer a identificação de condições de cultura que provocam a liberação do estado VBNC e da retomada do crescimento normal.

# (VI) Sobrevivência a longo prazo em *P. freudenreichii* CIRM-BIA138 na matriz de queijo:

Os nossos resultados permitem uma melhor compreensão dos mecanismos moleculares envolvidos na sobrevivência a longo prazo de *P. freudenreichii* submetida a uma carência nutricional. Esses resultados, obtidos *in vitro* e em condições de laboratório, devem ser complementadas e validadas, estudando o comportamento de *P. freudenreichii* CIRM-BIA138 na matriz de queijo. Esta linhagem foi isolada à partir do queijo Emmental. A matriz de queijo é complexa, tem propriedades físico-químicas

específicas e as bactérias são imóveis, então, seria interessante estudar o comportamento de CIRM-BIA138 neste contexto. Isso iria avaliar, em primeiro lugar, a sua sobrevivência a longo prazo na matriz de queijo e, por outro, verificar se os mecanismos que se suspeita importante na transição para a fase estacionária tardia também estão envolvidos na sobrevivência a longo prazo em queijo.

### (VII) Ativação e regulação de vias metabólicas em P. freudenreichii CIRM-BIA138:

Após o consumo total de lactato, o seu substrato preferencial, P. freudenreichii CIRM-BIA138 ativa várias outras vias metabólicas para garantir a sua sobrevivência. Seria interessante entender como estas vias alternativas são ativadas. Todas elas estão ativas simultaneamente ou são induzidas somente quando o lactato está esgotado? Neste caso, eles são induzidos por concomitante ou sequencial? Seria interessante, neste caso, identificar os estímulos e mecanismos reguladores. Uma opção é a construção de mutantes de genes envolvidos e induzidos no início de cada um dos processos metabólicos específicos. Por exemplo, para ativar o metabolismo de inositol, poderíamos tentar determinar se P. freudenreichii reconhece especificamente, a presença de inositol no meio e, em caso afirmativo, como esta bactéria distingue o estímulo no meio e reorienta o seu metabolismo. Isto poderia envolver a construção de genes mutantes com iolT1 (Major myo-inositol transporter iolT1), iolT2 (Myo-inositol transporter iolT2) e iolT3 (Myo-inositol transporteriol iolT3). O acompanhamento por RT-qPCR, de outros genes envolvidos no metabolismo de inositol, e dosagens bioquímicas, da concentração de inositol nos sobrenadantes de cultura, podem ser realizadas nestes mutantes e podem ser comparadas a linhagem selvagem.

### Perspectives

Les résultats obtenus au cours de ce travail de thèse nous ont permis de mieux cerner le phénomène de survie à long terme chez *P. freudenreichii*. Ils ouvrent aussi plusieurs perspectives. D'une part, les données générées peuvent être complétées et exploitées pour améliorer notre connaissance sur la génomique de *P. freudenreichii* (points 1 et 2 ci-dessous). D'autre part, beaucoup reste encore à faire concernant l'analyse de la survie à long terme et plusieurs pistes pourraient permettre de compléter ce travail (points 3 à 7).

#### (I) Achever la séquence du génome de *P. freudenreichii* CIRM-BIA138 :

En première approche, on peut considérer qu'un génome complet (c'est-à-dire, sans gaps) n'est pas indispensable si l'on peut en extraire les informations nécessaires aux analyses transcriptomique et métabolomique de cette bactérie. Cependant, un génome entièrement assemblé permettrait une meilleure annotation et donnerait ainsi une meilleure qualité d'informations génétiques sur l'organisme.

Pour obtenir le génome complet de *P. freudenreichii* CIRM-BIA138, une option serait d'effectuer une cartographie optique du génome de cette souche. Ceci permettrait de pallier les difficultés rencontrées dans l'étape d'assemblage de ce génome, dues à la présence d'une grande quantité de transposases. La cartographie optique donne en effet un aperçu général du génome, aide à la réorganisation de contigs et à la fermeture des gaps. En outre, il est possible de combiner différents programmes d'assemblage pour améliorer l'ensemble du processus. Par ailleurs, comme cela a déjà été démontré dans de précédentes études (Garber *et al.* 2011), et pour complémenter les précédentes approches, il est possible d'utiliser les données transcriptomiques dérivées du RNA-Seq pour aligner les transcrits sur le génome et pour fermer les gaps. Les données de RNA-Seq peuvent également aider au processus d'annotation du génome, en utilisant des programmes tels que ReadXplorer qui permettent l'alignement des données d'expression de chaque gène sur le génome pour en affiner l'annotation (Hilker *et al.* 2014).

# (II) L'identification de nouveaux éléments dans le génome de *P. freudenreichii* CIRM-BIA 138 :

Les données de RNA-Seq permettent la découverte de nouveaux transcripts qui n'ont pas été détectés et annotés sur les précédentes versions du génome. Une analyse dédiée permettrait également d'identifier de petits ARN non codants chez *P*.

*freudenreichii* CIRM-BIA138. La technique de RNA-Seq s'est révélée efficace pour la détection de petits ARNs chez *Pseudomonas aeruginosa* (Gómez-Lozano *et al.* 2012), ainsi que pour la découverte de nouveaux transcripts dans plusieurs espèces bactériennes (McClure *et al.* 2013). Des programmes bioinformatiques, comme Rockhopper, grâce à une interface graphique conviviale, permettent à l'utilisateur d'y entrer des génomes de référence et de modifier des paramètres, tels que : les lectures *paired-end* ou *mate-pair* ; l'alignement des lectures de plusieurs réplicats ; le nombre des replicats et de conditions expérimentales différentes utilisés pour les analyses de RNA-Seq (McClure *et al.* 2013).

# (III) Compléter les données de RNA-Seq sur la phase stationnaire tardive de *P. freudenreich*ii CIRM-BIA138 :

Au cours de l'analyse RNA-Seq de la souche *P. freudenreichii* CIRM-BIA138, la quantité d'ARN extraite des échantillons liés à la phase de survie à long terme a été trop faible pour permettre un séquençage de qualité. De plus, les échantillons de la phase stationnaire tardive ont été contaminés par de l'ADN génomique. Ceci a fortement limité la portée des résultats obtenus. Il serait donc intéressant d'obtenir une plus grande quantité d'ARN et d'en améliorer la qualité, en augmentant le volume de culture bactérienne échantillonné (par ex. 25 mL) en vue d'améliorer, par la suite, les étapes d'extraction et de déplétion des ARNr en veillant à éliminer l'ADN génomique par des traitements à la DNase.

### (IV) Le mécanisme de dormance dans l'espèce P. freudenreichi :

De nombreuses bactéries à Gram positif et à Gram négatif entrent en dormance dans la phase de survie à long terme. Comme nous l'avons vu, parmi les 8 souches de *P. freudenreichii* testées, toutes survivent bien et leurs populations se maintiennent à des taux élevés au cours de la période de carence nutritionnelle. Nous avons cependant observé que la souche CIRM-BIA138 présentait un niveau de survie supérieur aux autres souches. Cette souche a fait l'objet d'une analyse plus poussée (démonstration de l'état VBNC et RNA-Seq). Dans ce contexte, il serait intéressant de compléter les résultats obtenus dans cette thèse, en explorant la capacité de survie à long terme et l'état VBNC d'autres souches. Ceci permettrait peut-être d'identifier des souches de *P. freudenreichii* présentant une faible survie à long terme puis d'identifier par génomique comparative les gènes impliqués (par exemple, par leur absence ou leur pseudogénisation) dans cette moindre survie. De même, d'autres stress (par exemple, thermique, acide, ou osmotique) pourraient être appliqués aux cultures en état de survie à long terme et carence nutritionnelle. Ceci permettrait d'identifier des phénomènes d'adaptation et de résistance croisées qui pourraient nous éclairer sur les mécanismes mis en œuvre et mieux expliquer le comportement de l'espèce en conditions de fabrication fromagère.

En outre, cette étude pourrait être complétée par la construction de mutants visant à interrompre les gènes impliqués dans le déclenchement de la dormance chez *P. freudenreichii*. Des travaux de ce type (construction de mutants) ont déjà été menés pour montrer l'implication du gène *clpX* chez *S. typhimurium* (Kusumoto, Miyashita and Kawamoto 2013) et *rpoS* chez *E. coli* (Boaretti *et al.* 2003) dans la survie à long terme chez ces espèces.

### (V) La réanimation dans l'espèce P. freudenreichii :

Au sortir de l'état VBNC, de nombreuses espèces bactériennes passent par un état appelé réanimation leur permettant de reprendre une croissance normale. La réanimation (ou ressuscitation) nécessite parfois des conditions de croissance particulières. De même, certains gènes semblent directement impliqués dans ce phénomène. Ainsi, chez *M. luteus*, il a été montré que les cellules sont réanimées après la production du facteur Rpf (*Ressuscitation Promoting Factor*) (Greenblatt *et al.* 2004a). L'investigation de cet état de ressuscitation dans les cellules de *P. freudenreichii* serait d'un grand intérêt pour compléter ce travail. Cependant, il est nécessaire d'effectuer des expériences supplémentaires prouvant que les cellules sont réanimées après l'état VBNC. Cela requiert d'identifier les conditions de culture qui déclenchent la sortie de l'état VBNC ainsi que la reprise d'une croissance normale.

# (VI) La survie à long terme chez *P. freudenreichii* CIRM-BIA138 en matrice fromagère :

Nos résultats permettent une meilleure compréhension des mécanismes moléculaires impliqués dans la survie à long terme de *P. freudenreichii* soumis à une carence nutritionnelle. Ces résultats, obtenus in vitro et en conditions de laboratoire, devraient être complétés et validés en étudiant le comportement de *P. freudenreichii* CIRM-BIA138 en matrice fromagère. Cette souche a été isolée à partir du fromage Emmental. La matrice fromagère est complexe, a des propriétés physico-chimiques particulières et les bactéries y sont immobilisées, il serait donc intéressant d'étudier le comportement de CIRM-BIA138 dans ce contexte. Ceci permettrait d'évaluer, d'une part, sa survie à long terme en matrice fromagère et, d'autre part, de vérifier si les mécanismes que l'on suspecte importants dans la transition vers la phase stationnaire tardive sont aussi impliqués dans la survie à long terme en fromage.

### (VII) L'activation et la régulation des voies métaboliques chez *P. freudenreichii* CIRM-BIA138 :

Après consommation totale du lactate, son substrat préférentiel, P. freudenreichii CIRM-BIA138 active plusieurs autres voies métaboliques pour assurer sa survie. Il serait intéressant de comprendre comment ces voies métaboliques alternatives sont activées. Sont-elles toutes actives en même temps ou sont-elles induites seulement lorsque le lactate est épuisé ? Dans ce cas, sont-elles induites de façon concommitante ou séquentielle ? Il serait intéressant dans ce cas d'en identifier les stimuli et mécanismes régulateurs. Une option consisterait à construire des mutants des gènes impliqués et induits au début de chaque processus métabolique spécifique. Par exemple, pour l'activation du métabolisme de l'inositol, nous pourrions essayer de déterminer si P. freudenreichii arrive à sentir spécifiquement la présence d'inositol dans le milieu et, si oui, comment cette bactérie reconnaît ce stimulus dans le milieu et réoriente son métabolisme. Ceci pourrait passer par la construction de mutants avec les gènes *iolT1* (Major myo-inositol transporter iolT1), iolT2 (Myo-inositol transporter iolT2) et iolT3 (Myo-inositol transporter iolT3). Le suivi, par RT-qPCR, des autres gènes impliqués dans le métabolisme de l'inositol et, par dosage biochimique, de la concentration d'inositol dans les surnageants de culture pourra ensuite être mené sur ces mutants et comparé à la souche sauvage.

**Referências/ Références** 

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**Anexos / Annexes** 

Anexo 1: Capítulo de livro: *Genomics* publicado em *A Text Book of Biotechnology* (disponível em www.smgebooks.com)

Annexe 1 : Chapitre du livre : *Genomics* publié dans *A Text Book of Biotechnology* (disponible en www.smgebooks.com)

# Genomics SMGr@up SMGr@up

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# Abstract

In the last few years, the technologies have revolutionized new areas in science especially those involving genomics, proteomics and transcriptomics. This chapter approach each of these areas showing their concepts, importance and applications.

**Keywords:** genomics, structural genomics, functional genomics, transcriptomics, proteomics.

# Genomics

# **Structural genomics**

# Introduction

Structural genomics is the analysis of sequence and structure of the genome elements such as genes, regulators and mobile elements. Sequencing is necessary to obtain this information. The identification of variants among genomes has applications in evolution studies, health, biotechnology and the comprehension of relation between genotype and phenotype. Knowledge of genome sequence is fundamental to obtain this kind of information, and sequencing is the process of characterization of nucleotide sequence at a region (Targeted Genome Sequencing, TGS) or in the entire genome (Whole Genome Sequencing, WGS).

The first complete genome sequence of an organism was achieved in 1995, from the Gram-negative Haemofilus influenzae. Since then, the number of sequenced genomes is increasing, mainly because of new technologies that reduced the required cost and time, and new bioinformatics tools able to process the volume of data generated. Initiated formally in 1990, the Human Genome Project (HGP) was an international collaborative project, that sequenced the human genome in 13 years at a cost of approximately US \$ 2.7 billion (http://www.genome.gov/12011238).

#### **Genome sequencing**

The procedure for sequencing complete genomes involves fragmentation of the DNA molecule, creation of genomic libraries, reading nucleotide sequence of each fragment and reassembles the genome by alignment of the reads. Since the 1970s, different methods have been developed to determine the nucleotide sequence of a DNA molecule.

#### **Sequencing generations and technologies**

In 1977, Sanger and Nicklen published a DNA sequencing method by chain termination. This method required the creation of libraries by insertion of DNA fragments to be sequenced into cloning vectors (plasmids or artificial chromosomes) and then amplified *in vivo*. Four sequencing reactions are performed for each fragment, each one containing a proportion of one nucleotide without 3' OH group (dideoxy nucleotides), which causes termination of synthesis of a new DNA strands in random positions.

The generated fragments are separated by size on gel electrophoresis and the nucleotide sequence is determined by the last nucleotide of each fragment (Sanger, Nicklen and Coulson, 1977). Over the years, improvements have been incorporated into the method, such as *in vitro* amplification and automation by thermal cyclers, labeling the amplified fragments with fluorescent dideoxynucleotides (ddNTP's), as well as automatic sequencing based on reading fragments by capillary electrophoresis. The sequenced fragments generated by this method have a maximum size between 800 and 1000 bases. According to the National Human Genome Research Institute (NHGRI), in 2001, the cost to sequence 1 Mb (1,000,000 bases) and the complete human genome

were respectively U\$ 5,292 and U\$ 95,293,072. In 2007, the cost was reduced to U\$ 397 and U\$ 7,147 (http://www.genome.gov/sequencingcosts/). The amount of data generated by chain termination technology was inadequate for sequencing projects and to obtain complete genomes, even with the reduction of required costs and time.

In 2005, Next Generation Sequencing (NGS) platforms were launched, belonging to the second generation. These platforms are characterized by (i) the use of genomic libraries independent of cloning and (ii) a higher throughput compared to the first generation, by means of parallel sequencing of thousands or millions of fragments. However, the necessity of amplification in the preparation of the libraries can bias the quantification of fragments, insert sequence errors during the replication and increase the complexity of sample preparation. In addition, the smaller size of the reads and the large amount of generated data make the assembly of a more complex genome and creates a demand for computational structures with greater processing and storage capacity. In 2014, the utilization of these platforms, made the cost for sequencing 1Mb and a human genome decreased to, respectively, U\$ 102 and U\$ 3,063 million in 2007 to \$ 0.05 to U\$ 4905 (http://www.genome.gov/sequencingcosts/). The first released NGS technology was pyrosequencing technology. The sequencing reaction occurs in emulsion oil droplets distributed in wells of a fiber optic slide. Each oil droplet contain one bead linked to a single library fragment and PCR reagents. The reaction receives a flow of each nucleotide at a time followed by a wash step. The incorporation of a nucleotide is detected by the releasing of pyrophosphate. This technology is used in 454 Genome Sequencer platform (Roche), launched in 2005, being able to generate 200.00 reads of 110 base pairs (bp), and generating approximately 20MB of data. In 2007, Illumina/Solexa launched 1G Genetic Analyzer platform. In this technology, the fragments connected to adapters are denatured and its ends hybridized to complementary sequence adapters present on the surface of a solid blade. Complementary strand are synthesized by extension of the adapter attached to a solid plate. Also in 2007, Applied Biosystems launched SOLiD platform, based on ligation sequencing. A new complementary DNA strand is synthesized by ligation of an oligonucleotide to a primer and cleavage of oligonucleotide 3' end, releasing a fluorescent color, which represents the first two nucleotides of the incorporated oligonucleotide. The new DNA strand is removed by cleavage of the last nucleotide primer and a new cycle begins, initiated one position behind in relation to previous cycle. A color code and the sequence of each cycle colors are used to determine the

nucleotide sequence. In 2010, Ion Torrent (Applied Biosystems) released the Personal Genome Machine, which sequencing chemistry is similar to pyrosequencing, however the incorporation of nucleotides is detected by pH variation. This technology uses a semiconductor in place of fluorescence and a scanning camera, resulting in higher speed, lower cost and smaller equipment. Since then, updates of each platform were launched, in order to increase the size of fragment sequences and reduce the required time, cost and labor.

The third generation is characterized by (i) non-interruption of the sequencing reaction to detect each nucleotide incorporated (flow type of each nucleotide), (ii) larger reads and (iii) sequencing of a single molecule. This has reduced the sequencing time and simplified genome assembly in relation to the second generation, as reads sizes exceeds 1000pb. This generation technologies are based on imaging of singles molecules of DNA polymerase as they synthesize a single molecule of DNA; threading DNA polymerase to or next a nanopore and detection of nucleotide passage; or imaging of individual DNA strands by advanced microscopy. One example is the Platform II PacBio RS (Pacific Biosciences), launched in 2013, that generate reads longer than 20kb.

#### **Genomic libraries**

The DNA library preparation is one of the fundamental steps for the success of a genome project. Despite the variation in protocols, the similarities are fragmentation of the sample to sizes between 50 and 500 nucleotides, selection of fragments by size, ligation to adapters. The adapters often contain elements for ligation on a solid surface, primer annealing sites for amplification and sequencing, and a barcode sequence that makes possible to sequence samples of different sources.

In single-end or fragments libraries, adapters are ligated to one fragment, generating one read. In paired-end libraries, the ends of a DNA fragment are read, generating two reads, separated by a gap of known size. In mate-pair libraries, DNA fragments are connected to an adapter, circularized, and their ends are read, generating two reads, separated by a gap of known size. The difference between paired-end and mate-pair library is circularization and larger fragments of the second one. The latter two libraries provide an approximate distance between two reads, and this information facilitates the genome assembly and makes it possible to detect genome deletions, inversions, duplications and nucleotide insertions.

## Applications

In studies with the genomic DNA, NGS technologies have been used in sequencing of complete genomes or fragments, analysis of genetic variation, chromatin immunoprecipitation followed by sequencing (ChIP-Seq), epigenetic markers, identification of genes associated with disease, genetic screening, monitoring of infectious agents, forensic research, metagenomics and agrigenomics. Thousands of and millions of sequencing projects genomes were released (http://www.genomics.cn/en/index). Among the RNA sequencing applications are analysis of gene expression and single cell transcriptome. In gene expression studies, NGS made the prior knowledge of genes unnecessary to the identification and quantification of transcripts, alternative splicing and sequence variations. The various applications of NGS led to the launch of benchtop sequencers, cheaper, faster, easier to use and accessible to small laboratories. These include GS Junior (Roche), Mi Seq (Illumina) and Personal Genome Machine (Life Technologies).

### **Genome Assembly**

After sequencing the genome is reconstructed from fragments reads, in a process called genome assembly. The assembly is performed by means of alignment and overlap of reads to generate contigs (Figure 1), and contigs overlap. It is important that all nucleotides of a genome are represented among the sequence reads. The theoretical (or expected) coverage of a genome refers to the average expected times a nucleotide is sequenced. The calculation is performed by multiplying the size of the reads by the number of reads, and dividing the result by the size of the sequenced genome. The depth of coverage refers to the number of times a particular nucleotide of the genome is represented in the reads. This number varies because the fragment library preparation, sequencing and the process of assembling the genome generate deviations in the distribution of reads. The breadth of coverage is the percentage of a given genome sequenced by a number of reads.



Figure 1 – Assembly of contigs sequences by reads alignment.

## Data analysis

The first assembly step is data analysis. Raw data from sequencer is converted to file formats used by assembly software. The main formats are FASTQ and FASTA. FASTA (Fast Alignment) format stores nucleotide sequences (Figure 2), while FASTQ stores nucleotide sequence and quality scores. Removal of poor quality sequences, and barcode reads is required, usually performed automatically by the assembly software. Reads quality is measured by the Phred index, a measure of error probability in identification of each nucleotide in a read.



Figure 2 – Fasta file containing two sequences (multifasta file).

## Assembly

To assembly a genome is necessary to find overlaps between reads and generate a consensus, or contiguous, sequence. The assembly software uses algorithms that create substrings of each read, called k-mers, and test alignments with k-mers of other reads (Figure 3) to generate contiguous sequences. The use of paired libraries facilitates the assembly process because the known distance among the pairs of readings. For example, if each paired reading belongs to a different contig, it is possible to determine the distance and orientation between these two contigs. A genome can be assembled using de novo or ab initio methods and by reference method.





The de novo assembly is performed only by reads overlapping, not requiring the use of a reference genome mapping (Figure 1). The most commonly used algorithms are the Greedy, OLC (Overlap-consensus) and De Bruijn graph. This method allows identification of absent regions in a reference genome. The difficulty of this method is repetitive regions where reads can be aligned in wrong places.

In the assembly by reference, the reads are mapped in a phylogenetically close genome. This method has lower computational cost and is useful in identifying SNPs, insertions and deletions between the reference and the new genomes (Figure 4). Another advantage is the resolution of repetitive regions, because the reads are distributed at the reference regions. However, absent sequences in the reference genome will not be mapped.

REFERÊNCIA:	ATGTCGTAGTCGATCGATCAGCTAGCTAGCATGCATCAGCTAGCATCGATCG		
LEITURAS:	ATGTCGTAG GTCGTAGTC CGTAGTCGATCG GTCGATCGATCAGO ATCAGO CAGO	GCAAGCATGC/ GCATGC/ C/ CTAGCAAGCATG CTAGCAAGCATG	ATCAGCT ATCAGCTAGC ATCAGCTAGCATCGAT CAGCTAGCATCGATCGATCGATCA CATCGATCGATCGATCA GATCGATCAGCGATGT



#### Scaffolding and closure of gaps

The contigs have to be ordered and oriented by overlaps between their ends (de novo assembly), mapping on reference genome or optical mapping. The latter is done by mapping restriction sites in vitro and comparison to the contigs. A contig is called scaffold after its position and orientation is defined. The not identified overlaps between the contigs and, mainly, uncovered regions cause gaps in the assembly. Gaps can be closed in silico by overlapping scaffold ends, extension of scaffold by paired reads libraries, mapping reads in another reference genome in the region equivalent to the gap. The closing is accomplished by *in vitro* primer design which is long to the edge of two adjacent scaffolds and subsequent amplification of the region of the gap, usually by the Sanger method by generating sequences which can cover or extend the gap ends of the scaffolds. The third-generation sequencing platforms which generate long readings are becoming obsolete closing gaps in vitro, mainly due to the cost required. If a genome is still contains gaps after assembly, it is called a draft. The most commonly used parameters to measure the quality of an assembly is the number of contigs, minimum and maximum contig size and N50. In a good assembly, the number of contigs and its sizes is, respectively, the minimum and the maximum possible. The N50 is the size of the contig which the sum with the others, in descending order of size, is greater than or equal to 50% of the assembled genome.

#### Genome Annotation (automatic and manual steps)

After assembly, the structure and function of genome elements as genes, regulators and other non-coding sequences have to be identified to posterior analysis. This process is called genome annotation. Gene prediction can be made empirically, by similarity with known sequences in databases and proteins, or *ab initio*, by computational techniques, using algorithms to find stop codons, coding sequences (CDS), start codons. Normally, the algorithms to predict coding sequences are based on Markov chain models. In Prokaryotes, gene prediction is based on Open Reading Frames (ORFs) in the six possible frames. An ORF is a sequence that initiates at a start codon and finish at the nearest 3' stop codon. The prediction of false positives, incorrect start codons and overlapping genes are the main problems of this method. This problem can be minimized by searching for ribosome binding sites (RBS) to indicate a true start site and homologies in closely-related organisms.

The occurrence of introns and alternative splicing makes Eukaryotic gene prediction more complex. The prediction is based on recognizing signal functions on DNA strand and posterior accurate prediction of gene structure and organization. Sequences are classified as coding and non-coding and functions are assigned by alignments with functionally-related documented sequences, involved in transcription, translation and splicing.

After prediction, annotation can be done automatically, by align similarity with sequences at databases and transferring the information about their function to the new sequence. Databases can hold information of nucleic acids (e.g. NCBI, EMBL), protein sequences (e.g. UniProtKB), domains (e.g. Pfam) and structures (e.g. Protein Data Bank), metabolic pathways, gene expression (e.g. dbEST), gene ontology and evolutionary relationships. Different databases are integrated by automatically annotation services, as provided by RAST (Rapid Annotation using Subsystem Technology). Automatic annotation can provide a great amount of information about a new genome from its sequence. However, not all databases are fully curated, updated or have standard terminology. Manual curation is done by checking the results for different databases to identify protein domains and functions, correct start codon positions, gene names, gene products and to identification of frameshift.

After annotation, the genome can be submitted to a database, as submission is a pre-requisite to publication in scientific journals. Once this data is available, it can be used researches about genetic diversity, evolution, biotechnology and health.

## **Comparative Genomics**

After sequencing, assembly and annotation of a genome, different analyzes will try to find out to remove as much information as possible from a genome of an organism. In this context, there is the comparative genomics in order to seek from genomic data, which are the between intra and inter-specific relationships. This can be studied using the homology the genes, loci, functions, processes and categories.

Currently, there are a number of tools available for the comparison of genomes at the level of their genes and proteins, among them: BLAST (Altschul et al., 1997), but their visualization is somewhat limited, hindering an overview of the genomes studied. However, emerged over the last decades other tools, which integrate different databases and integrate visualization and their processing, widely used by bioinformatics are they: CGView (Grant JR, Stothard P, 2008) and BLAST Ring Image Generator (BRIG). With these tools you can see the structure and organization of genes in one or more genomes through a graphical interface (Figure 6). The other programs, such as the PIPS, are already able to report data pathogenicity, improving the entire information and providing the user various relevant information to do exploration of the body (Soares S et al., 2012).

All of these tools require a computing power to process a large amount of data, since genomes can be processed genomes from of simple organisms (prokaryotes) to more complex organisms (eukaryotes). Another important detail to highlight is the fact that numerous incomplete genomes are being deposited in the National Center for Biotechnology Information (NCBI). Incomplete genomes are those assigned "draft", which will have a percentage of genetic information lost one time analyzed using these tools, this is because part of the "gap" cannot be correlated to anything since it is said to be unknown. Therefore, we emphasize the importance of finalizing a genome in all steps, assembly and annotation, thus avoiding the loss of biological information (Barbosa EGV, 2014).

## **Functional genomics**

#### Transcriptomics

#### **Concepts and evolution**

Transcriptomics is a global study of gene expression levels of all the genes in an organism (genome-wide expression profiling), in order words, a set of detectably transcripts in a given cell. Transcripts can be modulated diversed factors, such as environment, time, diseases and response to adverse processes.

The first idea of transcriptome emerged in the late 70's, described as the Dot-Blot technique in order to use nucleic acids for the simultaneous analysis of some genes in arrays (Kafatos et al., 1979). In 1995, with the evolution of different technologies, this method has been enhanced by improving their level of sensitivity in optical detection (Brown et. al.). After that, in 2002, the Affymetrix launch the identification of previouslly unknown genes and detection of splice variants. Finally, in 2008, the RNAseq technique revolutionized history with sequencing from new generation technologies.

## **Different RNAs**

Total RNA is the combination of coding and noncoding; being the first comprises the messenger RNA (mRNA), transcript that carries the code information for proteins synthesis. In prokaryotes, the mRNA can be transcribed in operons or single genes and contain the exact transcript, and in eukaryotes, the mRNA has fragments that are translated in proteins (exons) and other regulatory regions (introns) that are removed by a process call splicing.

The noncoding RNAs are a combination of different classes: ribosomal RNA (rRNA), transfer RNA (tRNA), small RNA (sRNA) and long noncoding RNA (lncRNA). This topic will focuses in the non-coding ones, since the coding ones represents less than 2% of the total transcripts of human cells, and to be more precisely the emphases will be given to the regulatory RNAs, which are need to maintain genetic integrity and gene expression in a fine tune. There is still a lot to find out about those types of RNA and as a result of the use of new technologies there set are continually rising.

The tRNAs were known to be responsible for transport specific amino acids for protein synthesis and also recognize their specific codons, but recently another function were attribute to this RNA, when they are exposed to environmental stress in eukaryotes. After cleaved by ribonucleases, tRNAs derived fragments are able to inhibits translation and consequently forms stress granules where the non-translated mRNAs are storages (Kato and Slack, 2013). Therewith, now the tRNAs are also recognized as a new class of regulatory RNAs in response to stress.

The sRNA are responsible for mediate post-transcriptional gene silence (PTGS) and transcriptional gene silence (TGS). In PTGS, the target is cleaved or transcriptionally repress after is bound with the small RNA. TGS is a protective mechanism of genome integrity that keeps a heterochromatic state through DNA methylation or histone modifications (Sabin, Délas and Hannon, 2013a). One of the most known type of sRNA, in eukaryotes, are the miRNA, noncoding RNAs responsible for regulates the expression of genes in a sequencing specific matter. Others types are siRNA, piRNA and sRNAs (prokaryotes). The siRNAs are double-stranded RNA (dsRNA), like miRNA, which directs transcriptional silence of targets, participates in histone modification, heterochromatin formation. Piwi-interacting RNA (piRNA) is a defense system against transposons that acts through deposition of modifications in histones, inducing a heterochromatic state. The piRNAs are present in the genome in clusters and are a germ line specific class from both male and female organisms, and

different from the others classes of sRNAs, piRNA requires the Piwi protein to be generated (Sabin, Délas and Hannon, 2013b).

In prokaryotes, regulatory noncoding RNAs are transcripts of 50 to 250 nucleotides known as sRNAs that can either bound to mRNA and regulates gene expression or bind to proteins and alter their function. sRNAs can act on a target in cis or trans, and are involved in stress response, virulence, quorum sensing, DNA elimination, among others. A recently discover mechanism is the clustered regularly interspaced short palindromic repeat (CRISPR) pathway which are responsible for direct sequence-specific cleavage of foreign DNA (Sabin, Délas and Hannon, 2013b).

Another type of regulatory RNA are the lncRNAs, transcripts with proximally 200 nucleotides or more that is not translate into proteins. They include different classes of RNA like enhancer RNAs (eRNA), small nucleolar RNA (snoRNA), intergenic and overlapping transcripts (Batista and Chang, 2013). Independent of the class, the lncRNA are cell specific with high selective pressure, considering the genetic sequences, the promoters and there function structure, and also can function in cis and in trans, affecting neighbor genes or more distant ones (Fatica and Bozzoni, 2014).

An important aspect of the lncRNAs is that they interact with diverse proteins, forms nucleus domains, interacting two distinct chromosomal regions and modulates the chromatin (Wang and Chang, 2011). The lncRNA can act as a regulatory element, having an influence in histone modification, activating and repressing, even silence alleles of several genes (genomic imprinting) (Lee and Bartolomei, 2013). One example is XIST, a lncRNA involved in the inactivation of the X chromosome in mammals, and aberrant XIST showed to be involved in human cancer and an alteration on a XIST regulator (TSIX – antisense noncoding regulator) result in alteration X inactivation and embryonic lethality (Fatica and Bozzoni, 2014). lncRNA are also involved in the development of central nervous system, heart among other organs and environment responses.

The field of study of the ncRNA is just getting started and there is still a lot to learn about then and what they influence in the eukaryote and prokaryotes life, and because of that the scientific world is focusing all efforts on the study of those transcripts, the real responsible for the complexly of organisms.

Methodology of study Advantages and disadvantages
#### Microarray X RNA-seq

Microarray and RNA-seq are experimental techniques to measure the expression levels of the transcripts. These technologies have driven the functional genomics research in different organisms, prokaryotes and eukaryotes, for studies of physiological or pathological conditions. Microarrays allow a quantification of nucleic acids (mRNA form of genomic DNA or cDNA) by hybridization to complementary array. However, the RNA-seq technique, by sequencing of RNAs, can apply the same protocol for various interests, such as total RNA, mRNA, small RNA, such as miRNA, tRNA and rRNA.

There are a number of advantages and disadvantages about these two techniques. Microarray techniques are limited to detect transcripts, whose genomic sequences are their known; however, the microarrays are excellent for specific identification of known variants, for example in diagnostic testing, such as: cystic fibrosis. Nevertheless, RNAseq is used for detection of known and unknown transcripts sequences and detects splicing and variants, especially you used for further depth studies in any organism, prokaryotes and eukaryotes.

#### **Real Time PCR**

Real time PCR is a golden standard technique of gene expression that detects the product as soon it amplifies. The sample (RNA of study) is first reverse transcribed and then a mix similar with the conversional PCR plus a fluorescent agent is added in the sample for amplification. Real time PCR are applied in a variety of biological fields such as microbiology, veterinary science, pharmacology, biotechnology among others and can be useful in detecting SNP variation, allelic discrimination, forensic studies, quantification of pathogens and genetically modified organisms (GMOs), food quality, besides gene expression and ncRNA analysis (Derveaux, Vandesompele and Hellemans, 2010; Navarro et al., 2015).

This methodology has some advantages over the conversional PCR considering specially the measure of the DNA concentration, and another relevant fact is that the results obtained can be either qualitative or quantitative (qPCR), different from de conventional. Real time PCR is widely used technique and so became necessary an experimental protocol with detail information, being a simple and efficient instrument of research is require a maximal quality control so the results can be more reliable and

possible to compare (Navarro et al., 2015). Real-time PCR is very good for the few genes already known, however when do you want to see all a comprehensive analysis already exist other techniques are more suitable and effective.

#### Applied biotechnology: Looking in to the future

In the past, the focuses of genetics studies were or in the genome or in proteins, but the discovery of new classes of RNA and what they can do opened up a whole world of different approaches to old problems. Besides the knowledge of the biology of the specie at a given moment and situation, obtained especially by the mRNA, now the RNA is also being used in medical treatment, agriculture improvement, and in others fields involving biotechnology.

The development of global transcriptome profile using next generation sequencing as a functional genomics tool allows a better selection of possible target for further studies. With that in mind plus the fact that constant improvement and discoveries are being made in the field of science, the techniques applied now that focused on RNA made incredible breakthrough and are a promise in various fields of research.

#### Ageing

The use of noncoding RNA like miRNA was first tested in the improvement of lifespan by Boehm and Slack in *Caenorhabditis elegans* and from that different authors pursued this topic (Boehm and Slack., 2005). The miRNAs modulates protein homeostasis, mitochondrial metabolism, stress response, including DNA damage (Kato and Slack, 2013). Using new sequencing technologies among other, researchers discovered that many C. elegans miRNA including lin-4 changed their expression during life and in the absence of lin-4 the animals decreased their lifespan (Kato *et al.* 2011). This and other breakthroughs demonstrated that miRNAs as capable of predicting the longevity and remaining lifespan, and that possibly others ncRNAs are also involved in ageing (Kato and Slack, 2013).

#### Gene function and vaccine production

Next generation sequencing became an extremely important tool for the understanding of gene function, since the costs of sequencing for the whole genome and transcriptome is more cheap and fast than previously methodologies. The study of the transcriptome allows us to understand what happens with the microorganism and the host during the infection and select potential virulent factors with more precision. This selection is possible thanks to studies of different expression in environment (e.g. culture medium) that simulates the conditions faced by the pathogen or by a double transcriptome study where both the microorganism and the host are analyzed *in vitro* or *in vivo*.

#### **Insect management**

Insects pests are a major problem in agriculture and the use of pesticides are slowly decreasing since it is toxic for humans, fact that brought the necessity for nontoxic approaches. RNA interference (RNAi), a process of specific gene silence based on dsRNA, was susceptibly tested in deleting essential genes in insects and by doing so caused lethality (Baum et al., 2007). Both miRNA and siRNA are pathways endogenous of insects and plants, and the siRNA are recognizing as a major antivirus defense mechanism of insects. Even so this strategy proved to have potential not only in killing pests but also in the control of diseases transmitted by insects (Burand and Hunter 2013). The problem encounter for this method was the way of delivery, since dsRNA apply do not replicate in the host and because of that need to be constantly supplied. Two popular approaches are the use of RNAi pesticides that are safe for humans and beneficial insects, and use of transgenic plants that already produce dsRNAs (Burand and Hunter, 2013) but in this case brings further problems that will not be discussed here.

#### **Disease therapy**

The RNAi therapy shown to be effective in treating a variety of disease from viral disease to single nucleotide polymorphisms (SNP) and oncogenes, but as mentioned in the topic above the delivery and possible toxicity of the RNAi are still a concern (Hannon and Rossi, 2004).

Human immunodeficiency virus (HIV) was one of the first target by RNAi in the attempt to silence an infectiousness targets, but since it is a highly mutate virus there is still a lot to figure it out until this approach be successful. A second method were then tested to improve the effectiveness of the treatment which instead of focusing on the virus the target were a host receptor require for the infection (Boden et al., 2003). The use of RNAi to treat disease are in progress and apart from HIV others illness are also in experimental testing and among then are hepatitis C virus (HCV), amyotrophic lateral

sclerosis (ALS), Huntington, different types of cancer and more (Hannon and Rossi, 2004; Havens, Duelli and Hastings, 2013).

Approaches that manipulates splicing also function as disease therapy by correcting aberrant splicing or inducing then. Mutation that disrupt or create new splice sites are very common, in both intronic and exonic location, and are known to cause genetic diseases (e.g. Duchenne muscular dystrophy, Miyoshi myopathy, Cancer) (Havens, Duelli and Hastings, 2013).

#### **Proteomics**

The development of new technologies, involved in next-generation sequencing technology and access the complete genomes has promoted an increase of information about genomic data for several organisms. Thus, the functional genomics come with aim of complementing genomic data, where through of transcriptomic and proteomic studies, has promoted the description of the gene and protein functions as well their interactions. However, unlike of genomic and transcriptomic studies, the proteomic try to evaluate the protein expression at a given condition. Thus, a proteomic study provides valuable information about: protein synthesis, qualitative and quantitative information of protein product, post-translational modifications, protein-protein interaction and subcellular localization.

Currently, due the great advances both in classical and high-throughput proteomic technologies, new advances has been observed in the identification of targets for infectious diseases, inflammatory diseases, chronic conditions, cancer, drug discovery. Furthermore, through of comparative proteomic screening, between two conditions, i.e normal and diseased cells, allow the identification of differentially expressed proteins, that can promoted the identification of novel targets and biomarkers (Heffner et al., 2014; Wu et al., 2008).

The proteomic allows the analysis of the gene expression on a large scale; this study is basically divided into two steps: (i) separation of the protein extract through of the intrinsic fundamental properties of each protein and (ii) identification of proteins through of the sequencing of peptides predigested enzymatically. However, the success of a proteomic study is due the complimentarily by bioinformatics analysis; where several softwares are developed to auxiliary in the data analysis, i.e., correlation with genomic data, furthermore, several public data repository for proteomics data were created. Thus, proteomics is presented as a multidisciplinary discipline in the investigation medical, biologic and biochemistry.

#### **Gel-based proteomic**

Gel-based methods like gel electrophoresis are routinely used in proteomic analysis to separation of molecules or peptides. Two-dimensional gel electrophoresis (2-DE) is a classical proteome technique, applied to resolution of protein from complex protein mixture. 2-DE is used to separate proteins in two dimensions: (i) during the process of isoelectric focalization (IEF) the proteins are separated according to the isoelectric point (pI) and (ii) using a SDS-PAGE the proteins are separate by molecular weight. After the electrophoretic resolution the spot/protein are submitted to tryptic digestion and identified by mass spectrometry (MS) (Görg et al., 2004). Thus, the combination of the 2-DE with the technique MALDI-TOF-MS (matrix-assisted laser desorption/ionization – time of flight), has promoted great insight in the study proteomics.

The 2-DE analysis allows the generation of proteins maps, where spots/proteins differentially expressed are visualized, as well the presence of post-translational modifications (PTM) and isoform of proteins. However, a disadvantage of 2-DE classical is gel-to-gel variation, thus to suppress this variation the two-dimensional fluorescence difference gel electrophoresis (2-D DIGE) was developed (Unlü et al., 1997). This technique allows the simultaneous separation of two samples different pre-labeled with cyanine dye (CyDye) fluorophores in the same 2-DE. Furthermore, the great advance of this technique is the realization of a multiplex assay, where an internal standard is used for normalization of data that minimize the experimental variation between gels and favor the quantitative analysis of the proteins (Arentz *et al.* 2014).

#### **Gel-free proteomic**

Due the great advances in the biotechnological, new technologies also has been developed in proteomic area, where several advances have been observed in recent years, in order to address the weaknesses observed in 2-DE. Thus, the development of gel-free methods, such as liquid chromatography (LC) MS/MS promoted a great advance in the proteomic field, where are obtained higher sensitivity, accuracy and resolution in proteomic study of large-scale. In this approach normally utilize a microcapillary reverse phase (RP)  $\mu$ LC system to separate the protein extract previously

cleaved into peptides and *in tandem* analyzed by MS/MS, using electrospray ionization (ESI) coupled the mass analyzer (time-of-flight (TOF), quadrupole and ion trap). Thus, LC-MS/MS analysis associated the database searching has promoted a high-throughput proteomic analysis (Roe and Griffin, 2006).

The abundance changes of proteins present in complex biological mixtures is a major challenge in the proteomic study, thus with the introduction of gel-free methods, several workflows to quantification have been developed such as: (1) Chemical labeling: utilize thiol-specific tags to covalently label in cysteine residues through of the isotope-coded affinity tag (ICAT) reagent approach, in addition other approach used to quantification using the chemical labeling is the technique iTRAQ, where is based in the labeling of an amine reactive, isobaric, isotope tag. (2) Metabolic labeling: metabolic precursors (amino acids, carbohydrates, organic salts) are introduces in the culture media and changed the molecular weight of the proteins, these metabolic precursors are labeling, through of the technique of stable isotope labeling of amino acids in cell culture (SILAC). (3) Label-free quantification: no label type is used to quantify the samples. The quantification is based on (i) spectral counting; where the quantification is achieved by total number of MS/MS spectra for a protein, and (ii) intensity of the chromatographic peak area under the curve or intensity of the precursor ion MS spectra (Otto, Becher and Schmidt, 2014).

#### Proteomic in applied microbiology and biotechnology

The proteomic study offer a global survey of cellular changes in different physiological states at protein level, thus several studies using distinct or combined proteomic approach has been applied both in the applied microbiology and biotechnology.

In the microbiology field several proteomic studies have been performed with lactic bacteria, due the importance of this group of bacteria in the aliment industry, act as components of probiotic products and mainly in the fermentation process of yoghurt and cheese as well in the long-term storage of these products. Thus, proteomic studies related to knowledge of the process adaptive of *Lactobacillus*, *Propionibacterium* and *Bifidobacterium*, to acid condition and bile salts, showed a set of proteins that favor the survival and adapt theses bacteria in theses environmental that are present in the gastro-intestinal tract (Leverrier et al., 2003; Sanchez et al., 2007; Hamon et al., 2012). In addition, the combination of bioinformatics tools and proteomic techniques has been

applied to explore the host-bacteria interaction process, through of the characterization of surface-exposed proteins main protein fraction used by bacteria to promote the interaction with receptors of the host-cell (Barinov et al., 2009; Meyrand et al., 2013). However, these studies favor the selection of strain bacterial that can be used in the alimentary industry.

The genomic data from cancer patients has promoted information about variations, mutations and molecular pathways this disease. However, these alterations reflect in the gene expression of the cell cancer and consequently affected the function and stability of the proteins coding by these genes. Thus, proteomic study allow unveil this alteration and identify proteins involved in this pathway. In addition, currently the proteomic has been applied in the research of potential biomarker to have utilized in diagnostic of various type of cancer. In this type of study the biological samples evaluated from blood specimens like serum or plasma, thus quantitative proteomic both label-based and label-free has promoted the identification of several regulated differentially proteins, when compared the normal cell and tumor cell (Barinov et al., 2009; Kocevar, Hudler and Komel, 2013; Meyrand et al., 2013).

Currently with the technology advances, great efforts have been directed to the study of sustainable energy using mainly the bioenergetics process, which aims at energy production from biological materials and photosynthetic organism. A of the applied bioenergetics process is at production of biofuels. Thus, several proteomic studies are used to characterize the main raw material used to produce biofuels. Proteomic study performed with the plants: Sorghum sugarcane and maize, and algae and cyanobacteria that also are used in biofuels fabrication, have promoted the characterization of proteins present in different subcellular compartments, involved in several biological process as well the identification of proteins associated the resistance and adaptation to several stress conditions. These studies combined different proteomic approach and the results obtained in these studies theses several source used in the bio-ethanol production, promote information about of the physiology of this organism as well the identification of targets that can be used in its improvement main during the cultivation in different climatic conditions, favoring the biofuels production (Ndimba et al., 2013).

## Application to a bacterial protein-protein interaction

With the advent of second and third generation of sequencing technologies of there has a significant reduction of cost and time, and simultaneously happend an amount of sequenced genomes. With that, the genome sequencing became more accessible the genomes sequencing, therefore increasing the amount of biological data such as DNA, RNA and proteins available in public databases about of various organisms (Casals et al., 2012; Koboldt et al., 2013; van Dijk et al., 2014a).

DNA, RNA and proteins do not act individually in a living organism they work together interacting with each other other, physicaly or regulatory, in such a way they can execute their biological processes. Knowing whereby these elements are grouped and how they interact is a relevant area of study and research, allowing the understanding of a given organism, disease or biological process at systemic level. Therefore, the data generated from DNA, RNA, proteins and other molecules, permit researchers to broaden the fields of research to better understand organisms and/or diseases, search for new pharmaceuticals, veterinary, agricultural and biotechnology compounds.

An important tool for understanding how these molecules act in complex biological systems are protein-protein interactions (PPI) that, pair-to-pair, form a complex set of biological interactions called interactome (Ngounou Wetie et al., 2014; Rao et al., 2014). A PPI is composed of nodes that represent proteins (A and B), and edges that connect these proteins (nodes). The edge that connects both proteins do not have source and destination, thus forming a non-directional interaction, being equivalent the interpretation that the protein interacts with protein B or that protein B interacts with the protein A (Figure 5).



Figure 5: Representation of the interaction between two proteins using the graphs theory where the nodes A and B are connected by an edge.

Each interaction has two elements, called binary, and several binary interactions form a complex network of PPI that may represent all interactions of an organism or pathogen-host (Figure 6). The literature describes methods for the identification of PPI, being classified as: (i) experimental - yeast-two-hybrid (Y2H), protein chip, tandem affinity purification followed by mass spectrometry (TAP-MS); - biophysical - atomic force microscopy (AFM), analytical ultracentrifugation (UC), nuclear magnetic resonance (NMR) or (ii) computational - three-dimensional structure, text mining, phylogenetic profile, phylogenetic tree, gene co-localization, interolog mapping-based and machine learning.



Figure 6: Example of a protein-protein interaction network, made up of several pairs of interaction.

In time, the methods can be classified by data generated as low-throughput or high-throughput. High-throughput methods are able to detect greater amount of interactions, but have low specificity, while the low-throughput methods, which test specific interactions, have high specificity. More specific is the method, most sure has that the interaction occurs within the organism and, the more sensitive is the method, the more interactions are known. Experimental and computational methods complement each other, both being necessary for the scientific community to better explore the data (Harrington, Jensen and Bork, 2008; Mulder et al., 2014).

Regardless of the method used, a network interaction can provide researchers with more knowledge about an organism, making possible to think about new biological hypotheses and direct new experiments. In this sense, though indirectly, network interaction can be used for biotechnological purposes. By better knowing the organism, this can be genetically modified and have his behavior changed, aiming at greater efficiency or quality in the production of a product of our interest, as well as, can be genetically modified to produce or metabolize a product that is harmful to human, animal health or the environment, such as bio-remediation of soils. Aided by a network of PPI this is possible.

Assuming that some gene, óperon or protein involved in a biotechnological process of interest be known, with the help of a PPI network, it is possible to identify the proteins that interact with (partner) the protein of interest. By identifying proteins an interaction, may also be known their functions allowing to better understand the biological process or even the biochemical pathway involved in biotechnological process of interest. This information, aggregated to the knowledge about the organism, can help a researcher to understand the organism at systemic level, allowing new hypotheses to be tested in the laboratory. Still, with a network of interaction topological analysis can performed to identify biologically important proteins as proteins Hubs that have a high degree of interaction with other proteins, and proteins Betweenness that make the connection between the functional modules in a network of interaction. These both protein class, Hub and Betweenness, are considered essential proteins (Mulder et al., 2014).

Information of interactions, protein Hubs and betweenness allies proteomic experiments or information to RNA-Seq, which test a condition of biotechnological interest and measure the expression of transcripts or proteins, extend the prospects of identifying targets or metabolic pathways relevant. Besides allowing know which existing proteins and the expression of these, it is possible to meet with who these proteins interact and work together for the realization of biotechnological condition tested. Anexo 2: Capítulo de livro: *RNA-seq: Reveling biological insights in Bacteria* aceito em *Next Generation Sequencing* (disponível em breve em InTech - Open Access Publisher).

Annexe 2 : Chapitre du livre : *RNA-seq: Reveling biological insights in Bacteria* accepté dans *Next Generation Sequencing* (bientôt disponible sur InTech - Open Access Publisher).

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**Abstract:** New technologies are being constantly released and those improvements carry by then bring advance to not only on transcriptome, the focus of this book chapter, but also to diverse areas of biological research. Since the announcement and application of the RNA-seq approach, discoveries are being made in this field and when we consider bacterial species, this progress proceeded a few years behind. However, with the application of RNA-seq derivatives approaches, we can reveal biologicals insights in Bacteria world and thus aspire to uncover the mysteries involving gene expression, organization and others functional genomics features.

Keywords: RNA-seq, bacteria, transcriptomics, bioinformatic analysis workflow.

### 1. Introduction

The RNA-seq technology has driven advances in gene expression analysis, through the new generation sequencing platforms, being versatile, powerful, ensuring quality results with accuracy and reproducibility never reached before. This technology generates information that designates significance to the set of transcripts (transcriptome), opening up possibilities to understand cell behavior in different environments. RNA is a component of excellence within the cell, since it plays different roles as messenger regulatory molecule, carrier and is essential for maintenance of housekeeping genes (Creecy and Conway 2015).

In 2005, the first new generation of sequencing technology was released and has since been evolving rapidly (Margulies et al. 2005). After starting the process of gene expression analysis in bacteria (Passalacqua et al. 2009; Perkins et al. 2009), which a

more accessible cost, shorter time experiment and without probe, the technology took off and today overlaps other tools used for this purpose, such as the microarray technology, till now extremely used for this type of analysis.

### 2. Applications of RNA-seq

The understanding of the transcriptome is essential to the knowledge of functional genomics of an organism. The development of next-generation sequencing (NGS) impact different areas, such as medical and industrial, and have gone through a revolutionary process where different approaches have emerged in the field of microbiology and molecular biology, among them the RNA-seq technique, in order to aid understand and bring answers to the bacterial domain investigations. In this section, we will detail some of these applications that are part of our current context.

## 2.1. The medical field

The implication of these NGS technologies in medicine allowed the expansion in the field of diagnostic, treatment and prevention, especially concerning the understanding of bacterial diseases. One of their major applications have been quantification of expression levels of each transcript under different conditions that simulate the intracellular environment, such as the work of Pinto and collaborates (2014) in order to understand the relationship between host-pathogen (Pinto et al. 2014). The validation of this technique to the transcriptome of the pathogenic bacteria as their host was demonstrated in the work of Westermann and collaborates (2012) through the dual RNA-seq, that simultaneously analysed gene expression of the pathogen and host (Westermann et al. 2012). This allows us to better understand the systems biology between bacteria and their host, helping scientists to develop drugs and vaccines.

Another field that has been much explored is the research involving metatranscriptome, as scientists seek to comprehend the composition and regulation of microbial ecosystems (Leimena et al. 2013; Maklaim et al. 2013) and pursuing this, they use the RNA-seq technique to generate a large volume of data what grants large reliability and allows a broad interpretation of the data. The study Leimena and collaborates (2013) validated RNA-seq technique in the microbiota of human's small intestine with ileostomy, aiming to understand the interactions involved in this microbial ecosystem and how these relationships can be associated with diseases (Leimena et al. 2013). Transcriptome analysis pipelines (see topic 5) can be used for different experimental designs, applied for many bacteria, besides those on the medical field.

## 2.2. The industrial field

Industrial applications have been developed over the past few years, mainly around the probiotic industry, since it brings gains to the world economy. The work Bisanz and collaborates (2014) shows metatranscriptome of probiotic yogurt through RNA-seq technique (Bisanz et al. 2014), seeking the understanding of metabolic activities that allows the survival of this organism in the products. The results show the adaptive capacity of this bacterium as well as the variation in differential gene expression according to the taste or storage life of the product (Bisanz et al. 2014).

Studies such as these are of great importance, because enrich the knowledge of the industrial field, opening new perspectives for an attractive area in the marketplace, which results in an improvement in the quality of the final product to be delivered to the consumer.

In addition to the probiotic market, another major importance sector is the production and synthesis of biomolecules by bacteria. Studies such as Wiegand and collaborates (2013) sought to understand through the RNA-seq technique the regulatory RNAs during fermentation of *Bacillus licheniformis*. This study identified active genomic regions, which in turn contribute to the efficiency and optimization of the fermentation process, which can promote the industrial production of exoenzyme and antibiotics (Wiegand et al. 2013).

Microorganisms produce antioxidant molecules can be used in the pharmaceutical and cosmetic industry, as well as other compounds, such as propionate, applicable in the production of chemicals aids and produced by *Propionibacterium freudenreichii ssp. shermanii*, considered valuable on the Food Industry (Wang and Yang 2013b). In this scope, the RNA-seq technology is very promising and its application is able to bring advances polls in these studies.

### 3. RNA-seq and derivative techniques

#### 3.1. RNA-seq

The RNA-seq technology is able to directly and quantitatively identify all RNAs: coding and non-coding; rare and abundant; smaller and larger. This method provides the transcription start site information (TSS), untranslated region (UTRs), detection of unknown open reading frames (ORFs), improving quality in genomic annotation (Sorek and Cossart 2010), and also allows the distinction between primary transcript and processed (dRNA-seq) (Sharma et al. 2010).

The major constraint is to ensure representatives for rare transcripts, and in this case, the recommendation is to increase the representation of reads per library (Haas et al. 2012) or enhance these transcripts eliminating the ribosomal (rRNA) and transfer (tRNA), that are in abundance in the cell representing about 95% of total RNA (Bischeler et al. 2015).

Despite being considered the gold standard in gene expression analysis, for some researchers is complicated to define this technology as the gold standard, a method that presents itself in different platforms and address different strategies, showing advantages and disadvantages. However, they do not question the superiority of the technology compared to other in the past (Kratz and Carnin 2014).

Even with all superiority, we cannot rule out the need for biological replicates and depth of sequencing, hence the results may achieve greater reliability and reproducibility (Sendler, Johson and Krawetz 2011). Differentially expressed genes are better appraised when we have samples with more biological replicates compared to an enhanced depth with less replicas (Liu, Zhou and White 2014).

Transcriptomics studies have provided a revolution in the bacterial environment. Different bacterial species have been targeted for RNA-seq studies (Sharma et al. 2010; Isabella and Clark 2011; Pinto et al. 2014; Patenge et al. 2015) and gene expressionbased discovery has transformed the scientific paradigm of these organisms. The detection of an unexpected amount of coding genes in *Helicobacter pylori* demonstrated that despite having a small compact genome the transcriptome of this bacteria is extremely complex (Sharma et al. 2010).

A surprising result was the detection of a large number of Transcription Start Sites (TSS) never achieved before by any technology, aside from derivative RNA-seq technology like the differential RNA-seq (dRNA-seq), which was able to differentiate primary transcripts that exhibit triphosphate end from processed transcripts that presents monophosphate end as rRNAs and tRNAs. In this case, to enrich mRNA, the strategy was the treatment of total RNA samples with exonuclease enzymes that degrade nucleotide monophosphate. This strategy identified 5'UTR end, operons, antisense transcription, revealing a new perception in the organization of the bacterial transcriptome and a new vision for analysis of individual genes (Sharma et al. 2010).

These results obtained allows us to infer a role of 5'UTR regions, and a correlation between size and cell function was proposed by the researches, whom found that larger size is related to pathogenicity (Sharma et al. 2010). These results show how little knowledge we have regarding microorganisms, believed to be the simplest form of life, and yet proves to be more complex than previously anticipated which still a lot to discover.

A RNA-seq application that has been widely used in bacterial genomes are studies focused to identify smalls RNAs (sRNA). These elements are regulators of various biological processes and were initially and mostly studied in *Escherichia coli* (Gottesman and Storz 2011). However, with the advances in technology, it has been possible to identify and characterize small RNA in a variety of bacterial species (Papenfort and Vogel 2010; Sharma et al. 2010; Kroger et al. 2012) . Yan and collaborates (2013) identified in *Yersinia pestis* an expression profile of sRNA in vitro and in vivo, which allowed the identification of new sRNAs and the recognition of gene expression modulation during the infection process, improving the understanding of transcription regulation mechanisms of this organism (Yan et al. 2013). The importance of these studies involving sRNA also includes assistance in research related with antibiotics therapies, a study in initial development albeit which a lot of knowledge to be better exploited (Storz, Vogel and Wassarman 2011).

RNA-seq has been used in different areas and situations, advanced studies with the technology can detect details in the cell expression (van Dijk et al. 2014b). Even with the difficulties in separating eukaryotic and prokaryotic material, it was possible to distinguish the simultaneous expression profile between the host-pathogen responses through dual transcriptome studies. This work allowed unknots the host response against the bacterial infection and virulence factors enabled to determine the infectious process (Humphyrs et al. 2013). These studies contribute to the research in the field of biological infection, considering diverse pathogens with different life cycles and methods of infection, granting crucial knowledge for studies of diagnostics and vaccines, as metatranscriptomics study.

After a relatively short time in the market, RNA-seq can accurately reveal structural and functional elements of the bacteria. The mapping of transcripts in the

genome can refine the annotation or even identify new regions, improving the quality of the studied genome, compared to regions previously annotated by predictors or assembled by a ab initio approach (Martin and Wang 2011; Tjaden 2015), and can even check out the abundance of transcript expression.

Data arising from a quality genome tends to collaborate with more promising results, answering the biological question being analysed by researchers. In search of a genome quality, ab initio transcripts assembly or even a hybrid approach, which uses both the reference genome as a reference assembly, become an auspicious endeavour to solve many problems encountered in the genome and complicated to adjust (Martin and Wang 2011).

Pinto e collaborates (2012) conducted a study in *Corynebacterium pseudotuberculosis* adopting ab initio assembly, and therefore was possible to identify differences in the expression of active genes in different environmental conditions, allowing the detection of possible new virulence factors involved in pathogenicity, making it reportedly targets for vaccine development, diagnostic or treatment against caseous lymphadenitis disease caused by this bacterium (Pinto et al. 2012).

These results infer the importance of this technology and the possibility of going further with a tool that aims to improve and probably will expand the field of analysis, making the results increasingly closer to the bacterial molecular reality.

#### 3.2. tagRNA-seq

Bacterial RNA can be divided in two groups: primary or processed transcripts. Primary transcripts are represented by the presence of 5' tri-phosphate (5'PPP) which includes messengers RNA (mRNA) and small RNAs (sRNA), and processed transcripts are those carrying 5' mono-phosphate (5'P) such as mature ribosomal RNA (rRNA) and transfer RNA (tRNA).

Transcriptome studies usually focuses on mRNAs or sRNAs and because of that, depletion or enrichment reactions are needed to eliminate specially the rRNAs, since they represent proximally 95% of the total bacterial transcriptome (Bischler et al. 2015). A recently developed approach called dRNA-seq (Sharma et al. 2010) revolutionized the research of the primary transcripts by consider the 5' difference between the primary and processed groups, as mentioned previously (see topic 3.1).

RNAs are very stable and during the whole preparation, considering the "whetlab" experiments, some transcripts are partially or totally degraded. 5'PPP or 5'P are one of the mechanisms of protection against exonucleases and the first degraded portion. During that process, information are lost and some primary transcripts ended up with 5'P and so being treated akin as a processed transcript and consequently being eliminated during the dRNA-seq technique. Considering this flaw, a new methodology was created to overcame this problem by tagging and clustering the two groups together in a RNA-seq derived approach named tagRNA-seq (Innocenti et al. 2014). This technique also consider the difference between processed and primary transcripts, but instead of degrade the processed ones, two different ligation reactions are implemented with two different markers (PSS-tag and TSS-tag). Figure 1 exhibits briefly the methodology considering the three main steps: (1) the first reaction tags (PSS-tag) on the processed transcripts; (2) treatment with tobacco alkaline phosphatase (TAP), where the 5'PPP loses two phosphate, which allow the third step; (3) the second ligation reaction (TSS-tag) on the primary transcripts. After those steps, the transcripts are sequenced and due to the different markers, they can be distinct and compared (Innocenti et al. 2014).



**Figure 1.** The three main steps of the tagRNA-seq approach. (1) The first ligation reaction, during which the attachment of the PSS-tag (blue) to the processed transcripts (5'P); (2) Treatment with tobacco alkaline phosphatase (TAP), turning triphosphate to monophosphate groups; (3) The second ligation, corresponding to the TSS-tag (yellow) marker on the previously 5'PPP group (primary transcripts). The different markers allow the differentiation of the triphosphate and monophosphate groups after sequencing.

This methodology was first described in *Enterococcus faecalis* (Innocenti et al. 2014) and was based in other technique 5'tagRACE (Fouquier d'Herouel et al. 2011), a 5'RACE derived method. The results provided by tagRNA-seq improved the annotation of the *E. faecalis* genome for having identified or correct several genome portions, including non-coding and coding regions. This study also compared different libraries to prove the effectiveness of this innovative approach and with this provided a new method capable to differentiate primary and processed RNAs, and suited to better comprehend the genetic information of bacteria as other groups (Innocenti et al. 2014).

dRNA-seq and tagRNA-seq are approaches that allow a new view of the transcriptome by selecting the primary transcripts for sequencing or differentiate the primaries from the processed transcripts for a larger insight in the transcriptome. Those state-of-the-art techniques promises a better understanding of RNA structures like TSS, 5'UTR, promoters among others besides the knowledge of non-annotated genes and small RNAs.

### 3.3. FRT-seq (Flowcell Reverse Transcription Sequencing)

Flowcell reverse transcription sequencing (FRT-seq) is a new and improved methodology derived from the RNA-seq technology that was created for Illumina sequencers. Unlike RNA-seq, FRT-seq does not require amplification by PCR, a step that usually introduces bias in the results by displaying an erroneous view of the quantity of some RNA species (Mamanova and Turner 2011). Another important feature of this Illumina sequencing methodology is the ability to generate strand specific information, use of pair-end library and the need of a considerable initial amount of RNA template. The PCR free amplification is a major step forward a more comprehensive library, akin the original one, and also without formation of intermolecular priming artefacts, among others errors, and probably will be a fairly used technique in a near future (Mamanova et al. 2010; Mamanova and Turner 2011). The third generation sequencing platforms like Nanopore and PacBio also uses amplification free approaches but neither are currently being broadly used since they still exhibits sequencing errors.

The FRT-seq technique comprises the fragmentation of the template (e.g. mRNA) followed by ligation of adapters in both 3'end and 5'end, responsible for the hybridization of the template with oligonucleotides on the flowcell surface. The next steps performed are the quantification, the reverse transcription and then the sequence reaction (Mamanova et al. 2010; Mamanova and Turner 2011).

This approach can be applied for booth eukaryotes and prokaryotes although the amount of published papers involving eukaryotes are more substantial. Considering the Bacteria world, we can quote papers involving *Salmonella enterica* (Kroger et al. 2012) and *Shigella fleneri* (Vergara-Irigaray et al. 2014). The FRT-seq methodology was practiced as a complementary approach to describe the transcriptional landscape of the species, and in both cases, FRT-seq showed greater sensitivity and an excellent concordance when compared to others approaches and replicates.

The *S. enterica* paper showed that FRT-seq is as efficient as any other sequencing methodology, such as RNA-seq and dRNA-seq techniques (Figure 2) (Table 1) (Kroger et al. 2012). Figure 2 compares 9 different RNA libraries (TEX (1, 2, 3); RNA-seq (1, 2, 3, \*); FRT-seq (depleted and not depleted). TEX is a dRNA-seq methodology (see topic 3.1 and 3.2) and together with the first three RNA-seq biological replicates were sequenced in a 454 (1 and 2) or a Illumina GAII (3) sequencer. The charts relates the percentages of different RNA species and shows that the FRT-seq libraries show similar results or even better than the others approaches. The data presented on Table 2 also supports this claim, especially considering the total number of reads and the uniquely mapped reads achieved by FRT-seq libraries.



**Figure 2.** Sequencing methodology comparison. Adapted from Kroger et al. 2012. IGR – Intergenic region; TEX – libraries treated with terminator exonuclease; RNA-seq\* – library enriched for small RNA species (sRNA).

Library	Sequencing technology	Description	Total number	Number of reads (not mapped)	Number of reads (uniquel y mapped)	Percent uniquely mapped reads [%]	Minimum fold coverage <sup>#</sup>
TEX_1	454	dRNA-seq library biological replicate 1	161,03 1	72,623	88,408	54.90	1.11
RNA- seq_1	454	RNA-seq library biological replicate 1	248,99 3	83,030	165,963	66.65	2.03
TEX_2	454	dRNA-seq library biological replicate 2	111,462	10,785	100,677	90.32	2.16
RNA- seq_2	454	RNA-seq library biological replicate 2	93,337	38,577	54,760	58.67	0.61
TEX_3	Illumina GAII	dRNA-seq library biological replicate 3	1,738,8 67	122,058	1,211,42 6	69.67	20.99
RNA- seq_3	Illumina GAII	RNA-seq library biological replicate 3	2,148,5 63	136,871	1,360,11 3	63.30	21.16
RNA- seq*	Illumina HiSeq	RNA-seq library biological replicate 4	3,750,7 97	164,658	2,596,01 0	69.21	25.11
FRT- seq	Illumina GAII	FRT-seq library biological replicate 5	18,563, 218	4,203,71 5	2,456,79 2	13.23	16.42
FRT- seq dep	Illumina GAII	FRT-seq library biological replicate 5 rRNA depleted	24,585, 564	9,652,39 7	4,093,74 4	16.65	27.77

**Table 1.** Sequencing statistics. Adapted from Kroger et al 2012.

The *S. fleneri* paper (Vergara-Irigaray *et al.* 2014) also obtained a favorable result concerning FRT-seq, in fact this approach attained a bigger gene repertoire than the RNA-seq (Table 2).

	RNA	A-seq	FRT-seq		
	Condition A	Condition B	Condition A	Condition B	
Total number of mapped reads	20,099,597	22,736,494	49,925,286	47,605,241	
Total number of reads mapping to genes	1,525,782	2,271,423	3,037,954	2,585,600	
Reads mapping genes in sense	1,195,446	1,958,533	2,469,828	2,129,951	
Reads mapping genes in antisense	330,336	312,890	568,126	455,649	

Table 2. Sequencing statistics. Adapted from Innocenti et al 2014.

The data provided in this topic demonstrate the quality of this recent publish methodology and according to the authors (Mamanova et al. 2010; Mamanova and Turner 2011) news updates are still under improvements what will probably set up an even better approach for the users. The fact that this technique is only applicable for Illumina sequencers is a drawback, but since this sequencing platform is present worldwide, this disadvantage can be simply fixed or perhaps in a near future can be settled to work in others sequencing platform. Another particularity of this technique is the efficiency towards AT-rich genomes what do not constrain the application on AT-poor genomes, this fact is due to the PCR-free amplification what compel also a question on others sequencer like Nanopore and PacBio. Despite these issues, this technology has a bright future and is a great advancement for the conventional RNA-seq.

### 3.4. Chromatin immunoprecipitation followed by sequencing (ChIP- seq)

Chromatin immunoprecipitation followed by sequencing (ChIP-Seq) is a technique for genome-wide profiling of DNA-binding proteins, histone modifications, or nucleosomes (Myers et al. 2015). ChIP-Seq has become an essential tool for studying gene regulation and epigenetic mechanisms offering higher resolution, less noise, and greater coverage than its array-based predecessor ChIP-chip, especially after the decreasing cost of sequencing (Park 2009; Myers et al. 2013). This approach has six main steps: (1) it is initiated with cell cultures that are grown in defined conditions. When cultures reached the desired stage of development, they are treated with formaldehyde for crosslinking of proteins and DNA; (2) the chromatin is sheared by sonication into small fragments (200–600 bp); (3) an antibody specific to the protein is used to immunoprecipitate the DNA-protein complex; (4) the crosslinks are reversed by heating; (5) the released DNA are subjected to high-throughput sequencing; (6) *in silico* analysis in which resulting sequencing reads are studied for quality and cropped based

on quality of the reads (Johnson et al. 2007; Robertson et al. 2007; Park 2009). The cropped reads are then aligned to a reference genome. Afterward, areas of enrichment in the ChIP-seq data is identified and represent where the TF binds throughout the genome, usually called as peaks. CisGenome, MOSAiCs and MACS are some known algorithms that have been utilized for bacterial ChIP-seq analysis (Gentleman et al. 2004; Park 2009). After peaks are associated with genes downstream, a number of bioinformatics analyses can be carried out, including identification and analysis of motifs, differential analysis and association with expression data for deep understanding of bacterial regulon as shown in Figure 3 (Myers et al. 2015).



Figure 3. ChIP-seq sample preparation and analysis. Adapted from Myers et al. 2015.

As the whole-genome transcription profiling cannot reveal whether the influence of the Transcription Factors (TF) on RNA levels is direct or indirect, this requires identification of Transcription Factors binding within the appropriate promoter region. The ChIP-seq provide information about where transcription factors are bind, thus, by integrating ChIP methods and transcription profiling, it is possible to identify all direct regulatory targets of a TF for a given condition. For example, work carried out by Stringer and collaborates (2014) on *araC* gene of *Escherichia coli*, and *Salmonella enterica* identified direct regulatory targets of AraC, including 5 novel target genes: *ytfQ*, *ydeN*, *ygeA*, and *polB* (Stringer et al. 2014). Though ChIP-seq has been used only in moderation to study bacterial systems in a few bacterial species, such as, *Vibrio harveyi*, *V. cholerae*, *Rhodobacter sphaeroides*, *Mycobacterium tuberculosis*, *S. enterica*, and *Caulobacter crescentus* (Kahramanoglou et al. 2011; Myers et al. 2013, 2015; Singh et al. 2014; Haycocks et al. 2015) it is used for the identification of novel regulatory interactions, even for well-studied proteins (Grainger et al. 2007).

ChIP-seq could be an efficient tool in combination with RNA-seq to get the detail information of bacterial transcription regulation and how bacteria respond to different external conditions.

### 3.5. RNA Immunoprecipitation Sequencing (RIP-seq)

RNA Immunoprecipitation (RIP) is the study of intracellular RNA and protein binding; it is a tool to understand the dynamic process of post-transcriptional regulatory networks. In this technique, an antibody against a protein of interest is used to recover RNA species bound to the protein and since the sequence information of RNA species bound to a specific protein is often desired, an approach combining RNA immunoprecipitation with sequencing technology (RIP-seq) was created (Chu and Corey 2012). The main challenges of RIP-seq is the cross linking step, which is relatively inefficient and only small amount of RNA is available for the library construction (Hafner et al. 2010; Chu and Corey 2012). After that, treatment with endonuclease elucidates the specific binding sites within the RNA as they will be protected from digestion, followed by the RNA protein complexes purification using electrophoresis and high throughput sequencing (Chi et al. 2009; Chu and Corey 2012), and finally the data obtained from the sequencer are analyzed using bioinformatics tool. The first work using the RIP-seq based technique was carried in Salmonella by Sittka and collaborates (2008) (Sittka et al. 2008). In this analysis, they used RNA-binding property of Hfq protein, as result many new sRNA were discover that were not reported in previous studies (Cho et al. 2013). Thus, RIP-Seq could be an efficient tool for the identification of bacterial non-coding RNAs.

#### 3.6. LEA-seq (Low Error Amplicon Sequencing)

The LEA-seq technique (Low Error Amplicon Sequecing) emerged in 2013 and was developed and patented by Gordon and Faith (2014) (Gordon and Faith 2014). This method was created in order to improve the quality and the depth of sequencing runs, since the mass production data produced by NGS has caused a high rate of errors in the sequencing, due to algorithms or platform reading length problems (Gordon and Faith 2014).

LEA-seq is a nucleic acids sequencing technique that identifies events that occur at low frequency, seeking to understand mutation events. The three basic steps for implementing this technique are: (1) linear PCR, (2) exponential PCR and (3) sequencing. Whereas this technique is performed based on the bacterial 16S sequencing, in which PCR carries numerous times, and each PCR amplified uses specific primers for each linear molecule (Gordon and Faith 2014).

The LEA-seq technique is a quantitative method and presents the advantages of the generation and reading, which permits the formation of a consensus and eliminating errors for each molecule. Currently, the available techniques do not enable error detection in sequencing or whether there is a really variation in the sequence of that microorganism. The multiple sequencing through the LEA-seq technique allows for better quality and precision about the organism.

The study of Faith and collaborates (2013) aimed to identify the composition of the fecal microbiota of adults, the understanding of the role of these bacterial species and theirs therapeutic potential for intestinal diseases. This technique allowed not only work with a large number of samples (over 500 isolates), as well as contribute to a fast and accurate analysis of the data (Faith et al. 2013).

Researchers have a continuous interest in improving this technique, since it can be used for clinical investigation due to their high accuracy in cases, for example, patients with genetic mutations or somatic mutations again. LEA-seq can assist in the search and knowledge of the intestinal microbiota, as it may reveal its composition, opening up prospects for diagnosis, treatment and prevention of gastrointestinal tract diseases.

## 3.7. CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats)

Ishino and collaborates (1987) described CRISPR for the first time (Ishino et al. 1987). This system has been identified in 40% of bacterial genomes so far (Kunin, Sorek and Hugenholtz 2007) and they are defined as short repetitions of grouped bases. Although, the determination of the CRISPR locus and the characterization of adjacent genes, known as *cas* genes, responsible for the function of CRISPR, only happened in 2002 (Jansen et al. 2002). The system CRISPR/Cas uses small non-coding RNAs in association with Cas proteins. The Cas9 is a nuclease, which cleaves DNA in the selected region, so that the CRISPR system/Cas9 can be used for editing genomes.

The activity of the CRISPR/Cas involves three main mechanisms: (1) acquisition, the step which the DNA fragment was inserted into the CRISPR locus in the genome of interest of the study; (2) transcription, which the CRISPR locus will be transcribed and processed; (3) interference, which occur the ejection of nucleic acids; all those mechanisms contributes for bacterial persistence in the environment (Gasiunas et al. 2012; Jinek et al. 2012). Furthermore, the CRISPR provides mechanisms to limit the spread of antibiotic resistance or virulence factors, but the study carried by Gophna and collaborates (2015) demonstrates that even through there is different measurements to evaluate the horizontal gene transfer is not possible to identify a correlation between the CRISPR/Cas system and the evolution of the species, so that changes occur only at the population level (Gophna and Ron 2003).

RNA-seq helped annotation transcription of regions, mainly non-coding, as well as enables the identification of CRISPR elements in prokaryotes (Heidrich et al. 2015).

The CRISPR system can also be used as a tool in studies centred in gene regulation, since this system is able to activate or repress genes.

The working of Zoephel and Randau (2013) discusses how the structure of CRISPR can affect the maturation of the RNA and thus influence the functionally of the CRISPR/Cas system (Zoephel and Randau 2013). The RNA-seq approach was used to evaluate differential gene expression in *S. aureus*, a pathogen of major importance, and was capable to identify the CRISPR in these strains, and investigating their possible role, since these regions shows an adaptive response to infection (Osmundson, Dewell and Darst 2013). Thus, we have seen how important the use of RNA-seq approach to the magnification of knowledge and function in prokaryotes.

## 4. RNA sequencing platforms

The RNA-seq approach can be applied in different next-generation sequencing platforms and the results obtained by then are proportional to the machine capability. In Table 3 a comparison are being made with some of the most currently employed platforms (Li et al. 2014).

**Table 3.** Different Next Generations sequencing platforms in the study of RNA-seq.Adopted and modified from Li et al. 2014.

Company Name	Instrument	Version	Run Time (Hours)	Read Lengths (Mean)	Reads Per Run (Millions)	Applications
Illumina	HiSeq 2000	High Output	132	50	6,000	Gene expression, Splice junction detection, variant calling, fusion
Illumina	HiSeq 2500	High Output	132	50	6,000	Gene expression, Splice junction detection, variant calling, fusion
Illumina	MiSeq	v2 kit	39	250	30	Splice junction detection, variant calling,
Life Technologi es	PGM	318 Chip	7.3	176	6	Splice junction detection, variant calling
Life Technologi es	Proton	Proton I chip	2-4	81	70	Gene expression, Splice junction detection, variant calling
Pacific Biosciences	RS	RS	0.5-2	1,289	0.03	Splice junction detection, variant calling, full-length gene coverage
Roche	454	GS FLX+	20	686	1	Splice junction detection

#### 5. Bioinformatic analysis

Experimental investigations in prokaryotes has been facilitated, extended and complemented by computational approaches (Backofen et al. 2014). Huge amounts of data has been generated by RNA-seq experiments and needs to be stored and analysed using computational techniques and tools (McGettigan 2013). This amount has become a bottleneck to bioinformatic analysis and to biologists, since today's transcriptome analysis consist of experiments and data evaluation (Backofen et al. 2014). Extract biological information from RNA-seq datasets requires bioinformatic knowledge and tools, making the software choice an important issue to successful RNA-seq analysis (Pinto et al. 2011; Backofen et al. 2014).

According to Chierico and collaborates (2015) (Del Chierico et al. 2015) and Pinto and collaborates (2011) (Pinto et al. 2011) RNA-seq process can be understood as a five-step process: (1) isolation of total RNA of the organism; (2) mRNA enrichment; (3) synthesis of cDNA; (4) NGS sequencing, which returns raw data; (5) bioinformatic analysis (Pinto et al. 2011). A flowchart of this process can be seen in Figure 4.



Figure 4. RNA-seq five-step process.

This session focuses on bioinformatic analysis and available computational tools. Based on literature review (Pinto et al. 2011; McClure et al. 2013b; Backofen et al. 2014; Del Chierico et al. 2015; Tjaden 2015), bioinformatic analysis can be comprehended as the extraction of biological information from the sequencing of raw data and classified/divided (Figure 5).



Figure 5. Bioinformatic analysis workflow.

#### 5.1. Bioinformatic workflow

Quality check step aims to increase the accuracy of the results by removing sequences that can contain errors (de Sá et al. 2015), trimming sequences introduced in the library preparation step, as adapters and poly(A)-tails (Forstner, Vogel and Sharma 2014), and removing reads with low phred quality. However, regarding that, the usage of poor-quality bases can lead to less precise results (Ramos et al. 2011), and considering this, quality check can affect drastically the next steps.

Some RNA-Seq pipelines implements quality checking, like ReaDemption (Forstner, Vogel and Sharma 2014), which performs quality trimming, removes adapters

and poly(A) tails, and discards reads shorter than a given cut-off (the default cut-off is 12 nucleotides (nt)). Quality Assessment software (Ramos et al. 2011) evaluates quality based on quality-graph analysis and estimated coverage. According to Backofen and collaborates (2014) (Backofen et al. 2014), FastQC (http://www.bioinformatics.babraham. ac.uk/projects/fastqc/) is a commonly used tool to check reads quality determining the quality profile of the reads. Software suites can be also used to this purpose, FASTX-Toolkit (http://hannonlab.cshl.edu/fastx\_toolkit/) provides tools to remove sequences attached in previous steps and other pre-processing strategies of raw data.

After quality check, if there is a reference genome mapping step will be done, otherwise de novo assembly. Mapping consists of producing the transcriptome map though aligning reads to a reference genome (Pinto et al. 2011) aiming to detect the wright position of reads and distinguish between sequencing errors and genetic variations (Caboche et al. 2014). Lots of mapping softwares have been released differing in their algorithms, memory management, velocity and computational cost back (Backofen et al. 2014) making the choice of a mapping tool a challenge. McClure and collaborates (2013) (McClure et al. 2013b) made a comparison between SOAP2, BWA, Bowtie and Bowtie2 aligners using 75 RNA-seq experiment data, and the comparison of mapping algorithms applied to IonTorrent data can be seen in (Caboche et al. 2014). After mapping quality can be evaluated, ReadXplorer software offers quality classification of read mapping to provide information about quality and quantity to each single read mapping (Hilker et al. 2014). This approach is recommended when a high-quality genome is available as reference, and if it is unavailable, transcripts should be assembled de novo (Tjaden 2015).

De novo assembly can be used when investigating poorly studied organisms (Haas et al. 2012), complex microbial communities or uncultivable organisms (Tjaden 2015). Both DNA and RNA must be assembled, but transcriptome assembly is significantly different than genome assembly (Haas et al. 2013), thus it is important to use RNA assemblers. Tjaden (2015) (Tjaden 2015) affirms that assemblers should be specifically designed to prokaryotes, owing to the different challenges eukaryotic and prokaryotic transcriptome faces. Bacterial genomes are often denser than eukaryotic, considering the proximity of the genes, neighbouring bacterial transcripts can overlap making it difficult to identify appropriately transcript boundaries and non-coding eukaryotic RNAs models are not appropriate to detect bacterial small regulatory RNAs (Tjaden 2015). An assembly comparison of three different softwares (Trinity, SOAPdenovo2 and Rockhooper 2) using data from 9 different bacteria can be seen in (Tjaden 2015).

When reference mapping or de novo assembly is done, data can be analysed structurally and differentially. Differential analyses main purpose is to determine the difference expression between different growth conditions or treatments (Seyednasrollah, Laiho and Elo 2013). Several softwares have been released for this purpose, but there is not a consensus about best practices, what makes difficult to choose a tool or method to use. Aiming to help in this issue Seyednasrollah and collaborates (2013) (Seyednasrollah, Laiho and Elo 2013) made a comparison among

eight differential expression software packages using two real datasets publicly available. Softwares that analyses differential expression can be based on Poisson method (DEGseq and Myrna), negative binomial method (edgeR and DEseq) or in other methods (Pinto et al. 2011; Seyednasrollah, Laiho and Elo 2013). Pinto and collaborates (2011) (Pinto et al. 2011) recommends using DEseq or edgeR when analysing replicates.

Transcriptome annotation and classification can be accentuated in structural analysis, evaluating transcripts regarding the genomic region they have been associated to and classified in: protein-coding, non-coding and intergenic regions (Backofen et al. 2014). Aiming to predict ncRNA transcripts several computational methods has been developed, Herbig and Nieselt (2011) (Herbig and Nieselt 2011) highlights SIPHT, sRNAFinder, sRNAscanner, NOCORNAr and sRNAPredict softwares. NOCORNAr distinguish itself by prediction and characterization of ncRNAs in bacteria (Herbig and Nieselt 2011).

Assess transcripts concerning genomic regions relies on transcript annotation. Computational approach is convenient to be used due its velocity and precision compared to manual annotation; however, human supervision of the results is considered important to avoid false-positives or missing features (Creecy and Conway 2015). In this technique some main structures must be detected: 5' transcript ends, 3' transcript ends, TSS and operon (Backofen et al. 2014; Creecy and Conway 2015).

## a. Transcript boundaries identification

Annotation of transcript boundaries is important to operon identification and regulatory analyses (Creecy and Conway 2015). Identifying 5' UTR is not always possible, a significant number of transcripts lacking 5' UTR were funded in bacteria and called leaderless transcripts, in this, transcripts translation start site and transcription start site remains in almost the same position (Backofen et al. 2014). Annotation of 3' UTR is important to get the whole analytical value of RNA-seq data, Creecy and Conway (2014) (Creecy and Conway 2015) affirm that current best 3' ends detection method is to search correlation between replicates data, highlighting that, the software package TransTermHP can find intrinsic terminators successfully.

## b. TSS identification

TSS annotation can assist ncRNA annotation and polycistronic transcripts (Backofen et al. 2014); according to Creecy and Conway (2013) (Creecy and Conway 2015), it is essential to discover unknown transcripts and to analyse operon, 5' UTR and promoters architecture. Although, there is not a well-established strategies to TSS identification, owing scarce knowledge about transcription start sites in bacteria Due to computational developments in both computational analyses and "wet-lab" experiments, TSS annotation becomes feasible (Backofen et al. 2014). TSSAR is a dRNA-seq data-based tool for rapid annotation of TSS that considers dRNA-seq library statistics (Amman et al. 2014), and according to Backofen and collaborates (2014) (Backofen et al. 2014), the main advantage remains in statistical analysis presented as an easy-to-use web service. TSSpredator tool presents an automated TSS detection and

classification from RNA-seq data performing a genome wide comparative prediction of TSS (Dugar et al. 2013. A comparison among manual annotation, TSSpredator and TSSAR annotation can be seen in (Amman et al. 2014).

## c. Operon identification

The operon represents clusters of co-transcribed genes regulated by the same regulatory sequence and co-transcribed in a single mRNA. This structure has a huge biological importance improving functional gene annotation and giving important information to studies in drug targeting, functional analyses and antibiotic resistance (Chuang et al. 2012). To handle operon occurrence complexity, it should be detected by operon architecture (i.e. 5' ends and 3' ends) and have a sufficient read coverage to connect promoters and terminators. A strong indication that an operon is real is if at least 90% bases of the reads is covered (Creecy and Conway 2015). Chuang and collaborates (2012) (Chuang et al. 2012) classify computational methods to predict operons and evaluates 15 algorithms in respect to accuracy, specificity and sensitivity.

# 5.2. RNA-seq Pipeline tools

Not all pipeline tools contemplate hole RNA-seq workflow described earlier, to help in the tool choice a software functionalities comparison were developed and can be seen in Table 4. To additional support, important issues about each softwares are described forward.

Tool	Quality Check	Mapping	<i>De novo</i> assembly	Differential analyses	
Rockhopper					
(McClure et al.	-	Х	Х	Х	
2013)					
Rockhopper 2				Х	
(Tjaden B. 2015)	-	-	X		
RNA-Rocket					
(Warren et al.	Х	Х	-	Х	
2015)					
READemption					
(Förstner, Vogel,	Х	х	-	Х	
Sharma 2014)					
ReadXplorer					
(Hilker et al.	Х	-	-	Х	
2014)					

 Table 4. Software comparison.

Rockhopper is a system designed specifically to bacterial transcriptome RNAseq data analyses. A novel approach to map transcripts is implemented in this software (similar to Bowtie2 approach). Mapping normalization is performed followed by transcripts assembly, identification of transcript boundaries, quantification of transcript abundance, testing for differential gene expression and operon prediction. Analysis results are presented using Integrative Genome Viewer, allowing different experiments to be viewed simultaneously (McClure et al. 2013b).

Rockhopper 2 is a comprehensive system focused in *de novo* assembly that supports differential analyses and transcripts abundance quantification, according to Tjaden (2015) (Tjaden 2015) it does not require high-performance computers and can run on personal computers. Rockhopper 2 implements a novel *de novo* assembly algorithm to bacterial transcriptomes, the algorithm works in two stages: (1) candidate transcripts are assembled using a found k-mer; (2) sequencing reads are mapped to candidate transcripts aiming to filter candidate transcripts to high-quality final transcripts. Concerning differential analyses, Rockhopper 2 first normalize each RNA-seq dataset enabling to compare different experiments or samples (Tjaden 2015).

RNA-Rocket aims to simplify the process of aligning RNA-seq data to a reference genome and generate quantitative transcript profiles. It is built on Galaxy to provide the necessary tools and services to process RNA-seq data, some benefits of its usage are the possibility of sharing results across research groups, the support of batch analyses for multiple samples and the integration of tools and projects integrating data from the PATRIC platform (Warren et al. 2015).

READemption pipeline aims to integrate individual RNA-seq analyses tasks and putting it into a friendly tool with a command line interface, this tool was primarily developed to analyse bacterial transcriptome. In order to use full modern computers capacity and reduces run time READemption offers parallel data processing. First performs quality polyA and adapters trimming followed by mapping, coverage calculation, gene expression quantification, differential gene expression analyses and plotting. The software is able to analyse RNA-seq data from Illumina and 454 platforms.

ReadXplorer offers straightforward visualizations and analysis functions built around its unique read mapping classification. Analysis as TSS and operon detection, differential expression, RPKM value and read count calculations are available in ReadXplorer and can be exported to Microsoft Excel files. Read mapping classification sorts read mappings into three different classes: perfect match, best match and common match. This classification is incorporated in all analyses functions.

### 5.3. Bioinformatic Challenges

Through bibliographic research (Dai et al. 2012; Mc Clure et al. 2013b; McGetttigan 2013; Van Verk et al. 2013; Forstner, Vogel and Sharma 2014; Tjaden 2015), it has been concluded that bioinformatics has many challenges related to computational issues. RNA-seq experiments generate large amounts of data that must be computationally processed, analyzed, stored and retrieved using a great deal of computational power. In addition to the computational issues, it is important to take into account that not all bioinformatic researchers have extensive computational experience: this makes the lack of user-friendly tools a problem for some users and an import issue for developers. However, great computers, excellent bioinformatic researchers and user friendly tools do not guarantee successful analysis. The software selected must be appropriate to each biological question and to the organisms studied. Even with all

questions presented here, RNA-seq analysis has been very successful in recent years. This success can lead us to imagine the wonderful possibilities for RNA-seq bioinformatic analyses in the future.

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### Anexo 3: Artigo: Value of a newly sequenced bacterial genome

### Annexe 3 : Article : Value of a newly sequenced bacterial genome

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### REVIEW

## Value of a newly sequenced bacterial genome

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# Abstract

Next-generation sequencing (NGS) technologies have made high-throughput sequencing available to mediumand small-size laboratories, culminating in a tidal wave of genomic information. The quantity of sequenced bacterial genomes has not only brought excitement to the field of genomics but also heightened expectations that NGS would boost antibacterial discovery and vaccine development. Although many possible drug and vaccine targets have been discovered, the success rate of genome-based analysis has remained below expectations. Furthermore, NGS has had consequences for genome quality, resulting in an exponential increase in draft (partial data) genome deposits in public databases. If no further interests are expressed for a particular bacterial genome, it is more likely that the sequencing of its genome will be limited to a draft stage, and the painstaking tasks of completing the sequencing of its genome and annotation will not be undertaken. It is important to know what is lost when we settle for a draft genome and to determine the "scientific value" of a newly sequenced genome. This review addresses the expected impact of newly sequenced genomes on antibacterial discovery and vaccinology. Also, it discusses the factors that could be leading to the increase in the number of draft deposits and the consequent loss of relevant biological information.

**Keywords:** Next-generation sequencing; Drafts; Prokaryotic genomes; Computational tools; Omics.

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**Core tip:** Next-generation sequencing (NGS) technologies have made high-throughput sequencing available to medium-and small-size laboratories, culminating in a tidal wave of genomic information. The quantity of bacterial genomes has not only brought excitement to the field of genomics, it has also heightened expectations that NGS would boost antibacterial discovery and vaccine development. Although many possible drug and vaccine targets have been discovered, the success rate of genome-based analysis has remained below expectations. Furthermore, NGS has consequences for genome quality, resulting in an exponential increase in draft genome deposits in public databases. This review will address the expected impact of newly sequenced genomes on antibacterial discovery and vaccinology, as well as the impact of NGS on draft bacterial genomes.

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# **INTRODUCTION**

Since its release in 2005, next-generation sequencing (NGS) has been responsible for a drastic reduction in the price of genome sequencing and for a tidal wave of genetic information [1]. NGS technologies have made highthroughput sequencing available to medium- and smallsize laboratories. The new possibility of generating a large number of sequenced bacterial genomes not only brought excitement to the field of genomics but also heightened expectations that the development of vaccines and the search for new antibacterial targets would be boosted. Nevertheless, these expectations were shown to be naïve. The complexity of host-bacteria interactions and the large diversity of bacterial genetic products have been shown to play greater roles in vaccine development and antibacterial discovery [2-4].

Additionally, as with any methodology, NGS presents its own drawbacks. Among the new sequencing technologies the most consolidated in the market are the 454 GS FLX platform (Roche), Illumina (Genome Analyzer) and SOLiD (Life Technologies)[5,6]. These devices are capable of generating millions of reads, providing high coverage genomic but with a drawback, reads are considerably smaller than the ones produced by Sanger methodology[7,8]. While Sanger methodology produces reads ranging from 800 to 1000 bases, NGS platforms produces reads ranging from 50 (SOLiD V3) to  $2 \times 150$  bases (Illumina)[9]. The small amount of information contained in each read makes it difficult to completely assemble a genome using exclusively computational tools [10, 11]. Therefore small reads made the genome assembly process a quite more laborious task.

In recent years, approaches that use hybrid assemblies were developed to facilitate the assembly process. They take advantage of high read quality of second generation sequencers, i.e., Illumina (Genome Analyzer), and longer read lengths from third generation sequencers, i.e., SMRT sequencers (Pacific Biosciences) and Ion Torrent PGM[12,13]. Although empirically logical, this kind of approach wasn't facilitated due to the lack of integration between sequencers.

In order to improving and verifying quality genome is essential to know which combination of sequencing data, computer algorithms, and parameters can produce the highest quality assembly[14,15]. Also, it is necessary to know the more likely type of error data a sequencer platform will present. For instance, Illumina and SOLiD are more likely to present nucleotide substitution, while 454 GS FLX and Ion Torrent are more likely to present indels [16]. Nearly none bioinformatic system has been developed to integrate reads from different sequencers into a single assembly [12, 17]. This new developed approaches aim to reduce the manual intervention in finishing genomes, since repetitive regions may be solved using an hybrid approach.

Although NGS is directly responsible for considerable growth in the size of genomic databases, it has also been indirectly responsible for a decrease in genome quality [1, 10]. The number of draft genome (partial data) deposits in public databases has grown exponentially since 2005 (Figure 1). In general, if no further studies will be developed using a particular organism's genome, it is more likely to be deposited as a draft genome. Otherwise, the painstaking tasks of improving and finishing the genome (complete data) must be undertaken [18].



Figure 1. Number of complete genome and draft genome (partial data) deposits in public databases.

This review will address the "scientific value" of a newly sequenced genome and the amount of insight it can provide. We will address the factors that could be leading to the increase in the number of draft deposits and the consequent loss of relevant biological information. Additionally, we will summarize the expectations created by NGS technologies regarding vaccine development and antibacterial discovery.

#### **Overview of Sequencing and assembly**

For 30 years, sequencing technologies based on Sanger chemistry dominated the market. Although sequencing had undergone numerous improvements over the years, gene cloning techniques were still necessary to obtain genomic DNA sequences. Therefore, the time and cost required to obtain a complete genome sequence remained high. Moreover, the capacity of parallel sequencing was quite limited [19-21]. NGS platforms made it possible to sequence complete prokaryotic genomes using massively parallel sequencing more rapidly and at a lower cost [20, 22].

Although NGS has facilitated sequencing processes, its relatively smaller reads make the assembly process a computational challenge [10, 11]. The main limitation of short-read assembly methods is their inability to resolve repetitive regions of the genome without paired libraries [11]. The assembly of repetitive regions was an important issue even before the introduction of NGS platforms; shorter reads only made the problem worse.

In 2001, Kececioglu et al [23] argued about the impossibility of correctly assembling regions of the genome that contain identical copies of a sequence. Usually, long DNA repeats are not exact copies. They contain small differences that could, in principle, permit their correct assembly. Nevertheless, a major difficulty arises from sequencing errors. Assembly software must accept imperfect sequencing alignments to avoid missing genuine connections between sequences [22]. With the small amount of information within each read adding to the inherent sequencing error, it is difficult to separate true differences within repeated sequences from sequencing errors.

A study by Phillippy et al [24] revealed that the majority of contig ends in draft genomes were associated with repeated regions. They concluded that it was possible to categorize the majority of mis-assembly events into two general classes: (1) repeat collapse or expansion; and (2) sequence rearrangement and inversion. Each of these classes exhibits specific mis-assembly signatures: the first class is the result of incorrect assembly in repetitive regions, including fewer or additional copies; the second class is the result of the rearrangement of multiple repeated copies, which is caused by the insertion of a read between them. The second class may be considered more influential because, if not fixed, it might be interpreted as a real biological rearrangement event [25, 26]. If the assembler cannot resolve the region between two genomic fragments, a gap is formed. Gaps may occur due to: (1) an intrinsic characteristic of the sequencing platform that leads to incomplete or incorrect information; or (2) the inability of an assembly algorithm to handle regions of low complexity or repeated DNA [18, 27, 28]. The process of identifying and closing these gaps is quite laborious and requires additional manual intervention.

Gap closure processes usually involve the design of primers flanking the gap region to perform semiautomated sequencing of the unrepresented parts of the genome[28]. Several bioinformatics methodologies have been developed to facilitate gap closure. IMAGE is a tool that uses de Bruijn methodology to fill gaps with short reads that are aligned with flanking regions of the gap and were not used in the assembly[28]. In 2011, Cerdeira et al[29] generated a similar strategy by using CLC Genomics Workbench for the recursive alignment of unused short reads from the SOLiD platform. GapFiller is another tool that uses local alignment; its main advantage is the use of paired reads to estimate gap size and allows define the type of paired library: reverse-reverse, forward-forward, reverse-forward and forward-reverse [30]. From a purely practical standpoint, assembly tools are not required to produce a perfectly finished genome as an output. Their main function is to reduce the sequencing reads to a manageable number of contigs [26]. The process of finishing a genome, ensuring that gaps are closed and the gene order is correct, requires human decisionmaking. Therefore, the lack of fully automated processes constitutes a bottleneck in generating complete genomes.

#### "Scientific Value" of a Newly sequenced genome

The value of a newly sequenced genome can be assessed using many different metrics. If publications are considered the main "currency" within the scientific community, there has been a considerable decrease in the value of new sequences over the last four decades.

The introduction of Sanger methodology in 1977 was one of the main landmarks in the early stages of the genomic era[31]. During the first years of using Sanger sequencing, a sequence of no more than 1000 nucleotides was sufficient for a work to be accepted in a journal such as Cell (current impact factor: 32.40) or Nature (current impact factor: 36.28)[32-34]. In 1980, the shotgun DNA sequencing methodology was introduced, enabling the sequencing of longer DNA fragments [35]. Complete bacterial operons were sequenced and published in journals such as Molecular Microbiology (current impact factor: 5.01) and Proceedings of the National Academy of Sciences (PNAS -current impact factor: 9.68) [36-38].

A combination of DNA sequencing improvements and the newly developed TIGR Assembler [39] culminated in the publication of the first complete bacterial genomes in 1995. Papers containing the complete nucleotide sequences of Haemophilus influenzae Rd (1830137 base pairs) and Mycoplasma genitalium (580070 base pairs) were both published in Science (current impact factor: 31.20)[40,41].

Almost 20 years later, a paper containing the sequence of a prokaryotic genome alone may be published in the Genome Announcement section of the Journal of Bacteriology (current impact factor: 3.82) or in Standards in Genomic Sciences (SIGS has not been published sufficiently long to receive an impact factor). A recent article by Smith even refers to the not-so-distant "death" of the "genome paper", noting that the space for genome publication may come to an end soon [42].

The publication impact of newly sequenced genomes decreased following DNA sequencing improvements, and the reason is no mystery. High-impact journals only

publish groundbreaking original scientific research or results of outstanding scientific importance. To produce a higher-impact publication, more information must be extracted from genomes. For instance, several genomes may be examined in a comparative genomic analysis or pangenomic study[43,44], or an analysis may focus on the presence or absence of specific markers or on small differences between DNA sequences[26,45]. In this context, the genome becomes a stepping stone to the main goal, the comparative analysis. As the basis of the analysis, the genome sequence remains important. Nevertheless, it may not be of sufficient importance for one to undertake the painstaking task of completing the genome sequence.

#### What Is Lost When we Opt for a Draft Ganome?

Over the years, arguments have been presented in favor both of complete genomes[41,46] and of the superior "tradeoff" that a draft genome represents[47]. The discussion has been centered around two main points: (1) to provide the greatest amount of useful data, sequences must be as complete as possible; and (2) draft genomes (partial data) are sufficient for most scientific contexts.

The issue at stake is the extra money and manpower necessary to finish a genome. Is the additional information contained in a finished genome worth the investment? To answer this question, one must identify the information that is lost from a draft and analyze the quality of data that is generated using drafts. Furthermore, it is necessary to understand the limits of draft genome use.

The first issue to consider is whether it is possible to properly identify all of an organism's genes in a draft genome. Gene characterization consists of the following: (1) gene prediction with the identification of an open reading frame (ORF); and (2) the functional annotation of the gene product. The main gene identification problems in drafts are associated with the partial or complete loss of ORFs [10]. Such errors may lead either to over-annotation, due to the annotation of multiple fragments originating from the same ORF, or to under-annotation, possibly due to the absence of partial or entire domains from the ORF [10]. These problems affect genomic analyses, causing errors due to missing ORFs that are not annotated or due to multiple fragments that belong to the same ORF but are annotated separately. In other words, the mere absence of a gene from a draft cannot be considered definitive proof of its absence from the organism's genome[10,41].

The pangenomic approach is one type of analysis that may be impaired by reliance on draft genomes, because many genes in a draft may be misidentified due to fragmentation.

Pangenomic projects attempt to characterize the gene pool of a bacterial species as the genes that are present in all strains (the "core genome") and the genes that are present in only a few species (the "dispensable genome")[43].

Horizontal gene transfer (HGT) analysis is another approach that cannot be performed using drafts. HGT is one of the main sources of variability among bacteria because it allows the acquisition of several new genes [36,37]. There is evidence that most gaps in genomic sequences are associated with transposases, insertion sequences and integrases, structures that usually flank a genomic island [48]. Another approach that may be impaired by reliance on drafts is phylogenomics, which aims to reconstruct both the vertical and lateral gene transfer processes of a bacterial species using a whole-genome analysis [49].

Although not strictly related to drafts, the functional annotation of genes is another feature that is usually neglected when we opt for a draft genome (Figure 2). Complete genomes may also present this problem because the quality of functional annotation is related to the amount of effort dedicated to a genome. DNA sequence is being generated much more rapidly than it can be analyzed; thus, a large proportion of the sequence information in databases has been annotated solely by automatic algorithms [50]. It is disturbing that although automatic annotation algorithms have improved over the years, misannotation has increased over time [50]. The misannotation of a reference strain is particularly harmful because the error will likely be propagated to other genomes. In our attempts to exploit the full potential of NGS, we risk having databases filled with incomplete and/or incorrect genomic data.

Structural genomics



Figure 2. General workflow during sequencing process a bacterial genome.

Because the purpose of many sequencing projects is to identify a small number of differences between a newly sequenced genome and the sequence of a closely related species, a large number of genomes are left as drafts [26]. Considering the constant evolution of organisms, a sequenced genome represents a snapshot in the biological history of a species. Therefore, a single finished genome might be useful for decades of future studies. By opting for draft genomes, we may be shutting down the full gamut of future scientific analysis.

## Vaccine Development

Genomic information was expected to boost vaccine discovery. In an attempt to measure the impact of genomic information on this field, Prachi et al[2] analyzed all the patent applications that contained genomic information.

They observed that there was an enormous increase in such applications shortly after the first complete genomes were released, but since 2002, there has been a continuous decrease. The authors attributed this decrease to more stringent legal requirements, which call for empirical evidence to complement in silico data. The initial increase in patent applications containing genomic information was related to the development of a new paradigm in vaccine development. In 2000, Rappuoli [51] described the "reverse vaccinology" (RV) concept, in which he proposed inverting the traditional process of antigen identification. Instead of identifying the antigenic components of a pathogenic organism using serological or biochemical methods, RV uses the organism's genome to predict all of its protein antigens. RV approaches mainly focus on secreted proteins because they are more likely to induce immune responses. Secreted proteins are involved in several processes that modulate the host-pathogen relationship, such as cell adhesion and

invasion, as well as resistance to stress conditions[52-54].

Over the years, several methodologies have been developed to predict secreted proteins and to evaluate their potential immunological properties.

In 2010, Vaxign was released as the first vaccine design tool with a web interface (http://www.violinet.org/vaxign/). Vaxign allows users to submit their own sequences to perform vaccine target predictions. The Vaxign predictions have been consistent with existing reports for organisms such as Mycobacterium tuberculosis and Neisseria meningitides [55]. Another vaccine design tool is MED (Mature Epitope Density -http://med.mmci.uni-saarland. de/). MED attempts to select the more promising vaccine targets by identifying proteins with higher concentrations of epitopes [56]. There are also tools exclusively for protein epitope prediction, such as Immune (http://tools.immuneepitope.org/main/) and Epitope Analysis Vaxitope (http://www.violinet.org/vaxign/vaxitop/index.php). Because a large number of bacterial genomes are already available, reverse vaccinology is quite accessible and inexpensive. Nevertheless, as has been previously discussed [57,58], the expectations for reverse vaccinology techniques do not correspond to reality, given the small number of vaccines have been developed using the bacterial genome sequences available[59]. This occurs because there are also several factors that are involved in the host response during infection, for example, the production of antibodies by the immune system.

#### **Antibacterial Discovery**

The period between the 1930s and the 1960s is known as the "golden age" of antibiotic discovery [11, 60]. During this period, most of the known classes of antibiotics were discovered. These discoveries involved screening natural products regardless of their mechanisms of action. After most of the low-hanging fruits were harvested, the rate of antibacterial discovery decreased, culminating in a slowdown beginning in the 1990s [61].

Hopes for turning this void into a rapid acceleration accompanied the completion of the first bacterial genome sequences. The goal was to use comparative genomic analysis to identify potential targets present in a desirable spectrum (e.g., the bacteria responsible for upper respiratory tract infections) [3, 4, 62]. It was naive to assume that having the genome sequences would be sufficient for this level of discovery; a possible drug target must undergo numerous stages, from discovery through human clinical tests, and it is not possible to develop drugs for all potential targets [3,62]. Nevertheless, the prospect of exploring hundreds of potential targets revived the interest of pharmaceutical companies.

After some years of trials, several companies ended their target-based programs because of a lack of productivity. Despite reports of multi-resistant bacterial strains, the efforts to discover new antibacterial targets were again reduced [63, 64]. Although genomics has not been able to reverse the lack of new antibiotic development, it has significantly improved screening methodologies. Genomics has facilitated high-throughput drug campaigns, which are being used to determine the mechanisms of action of antibacterial compounds and bacterial resistance mechanisms [4].

## Conclusion

Several next-generation platforms have been developed in recent decades, as well as bioinformatics programs to an enhancement of performance and optimization omics techniques. Is not yet possible to integrate reads from different sequencers into a single assembly [17,23]. This newly developed approach aims to reduce the amount of manual intervention needed to complete a genome sequence by using a hybrid approach to resolve repetitive regions.

Improvements are expected not only in sequencing platforms but also in assemblers. Recently, two groups assessed the quality of the currently available assemblers. The 2011 Assemblathon was the first competition among assemblers [65]. For this competition, simulated data were generated and groups of assemblers were asked to blindly assemble it. The use of simulated data poses a problem in determining the applicability of the results to other data sets. The 2012 GAGE (Genome Assembly Gold-Standard Evaluations) competition for assembling real data resulted in the following conclusions: (1) the data quality has a greater influence on the final outcome

than the assembler itself; and (2) the results do not support the current measures of correctness (related to contiguity)[26].

There is a large gap between the availability of genomic sequences in databases and the commercial production of vaccines and antibiotics in recent years, especially in the fields of investment and success ("expected return"). Drug development for all potential targets and effective vaccines has produced limited success. In contrast, there has been an acceleration in the discovery of new targets due to the refinement of bioinformatics tools for this purpose, such as epitope mapping and searching for secreted proteins. However, the major problems facing vaccine and antibiotic development, such as resistance mechanisms and host immune responses, remain unsolved.

Genome analysis constitutes a strategy for the expansion and diversification of the pharmacology and vaccinology sectors. This methodology can be used to explore a large number of targets and to reduce the costs of molecular and immunological tests. Finally, to improve the production of antibiotics and vaccines, it is necessary to know more about bacterial regulatory pathways. New interactome and microbiome studies must be implemented to assist this search.

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# Anexo 4: Nova montagem e anotação do genoma de *Propionibacterium* freudenreichii CIRM-BIA138

Com o intuito de finalizar o genoma alvo da tese, referente a linhagem de *Propionibacterium freudenreichii* CIRM 138, foi proposto como parte deste trabalho uma nova montagem e anotação deste genoma. Para isso, diferentes abordagens foram realizadas na etapa de montagem: a primeira tentativa foi realizada com a associação do programa *CLC Workbench* 7.0 com *CONTIGuator* 2.7 (*scaffolding* por referência com a CIRM-BIA1). Posteriormente ao alinhamento dos dados brutos gerou-se um *scaffold* e e estes foram alinhados contra o genoma de referência (CIRM-BIA1). O fechamento manual de *gaps* foi realizado com o auxílio do programa *CLC Workbench* 7.0 e pelo banco de dados do *Basic Local Alignment Search Tool (BLAST*), e, finalmente, 28 contigs foram obtidos como resultado final deste genoma. Outras abordagens de montagem de genomas, tais que o SPAdes 3.5 e *scaffolding de novo* com o SSPACE 2, também foram testadas com o objetivo de finalizá-lo, porém não apresentaram resultados promissores.

A anotação deste genoma foi realizada em duas etapas: automática e manual. A anotação automática foi executada pelo *RAST Server (Rapid Annotations using Subsystems Technology* - http://rast.nmpdr.org/rast.cgi) e a anotação manual foi realizada através do programa *Artemis - Genome Browser and Annotation Tool*, a partir do arquivo .embl. Neste arquivo, foram verificadas as posições corretas dos códons iniciais e de parada, além da curadoria de todas as sequências codificantes: CDS (*Coding Sequences*), por meio da combinação de diversos bancos de dados, como: *BLAST* (http://blast.ncbi.nlm.nih.gov/Blast.cgi), *Uniprot* (http://www.uniprot.org), *PFAM* (http://pfam.xfam.org/), *InterProScan* 5 (http://www.tcdb.org/).

# Annexe 4 : Nouvel assemblage et annotation du génome de *Propionibacterium* freudenreichii CIRM-BIA138

Afin de finaliser le génome cible de la thèse relative à la souche de *Propionibacterium freudenreichii* CIRM-BIA138, ont été proposé dans le cadre de ce travail, une nouvelle assemblage et annotation de ce génome. A cet effet, différentes approches ont été réalisés dans l'étape de l'assemblage: la première tentative a été réalisée avec l'association du logiciel *CLC Workbench* 7.0 com *CONTIGuator* 2.7 (*scaffolding* par référence avec la souche de *Propionibacterium freudenreichii* CIRM-BIA1), ensuite l'alignement des données brutes généré un *scaffold* et aussi ils ont été obtenus contre le génome de référence (CIRM-BIA1). La clôture manuelle des *gaps* a été effectuée avec l'aide d'un logiciel *CLC Workbench* 7.0 et avec la base de données du *Basic Local Alignment Search Tool (BLAST)*, et enfin 28 contigs ont été obtenues comme le résultat final de ce génome. D'autres approches ont été testés avec l'objectif de leur finaliser, comme le SPAdes 3.5 et *scaffolding de novo* avec le SSPACE 2, mais ne montrent pas des résultats prometteurs.

L'annotation de ce génome a été réalisée en deux étapes : automatique et manuelle. L'annotation automatique a été effectué par RAST Server (Rapid Annotations using Subsystems Technology - http://rast.nmpdr.org/rast.cgi) et l'annotation manuelle a été réalisée en utilisant le programme Artemis - Genome Browser and Annotation Tool, à partir de l'archive .embl. Dans ce fichier, les positions correctes des codons initiales et des codons d'arrêt ont été trouvés, ainsi que le vérification du génome des toutes les séquences codantes : CDS (Coding Sequences) en combinant plusieurs bases de (http://pfam.xfam.org/), 5 données. tels que PFAM *InterProScan* (http://www.ebi.ac.uk/Tools/pfa/iprscan5/) et transporter classification database (http://www.tcdb.org/http://www.tcdb.org/).



Anexo 5: Dados genômicos do Genomes Online Database (GOLD)



Projetos distribuídos de acordo com o seu domínio

Figura 13: Número total de projetos por ano depositados no GOLD (https://gold.jgi.doe.gov/statistics).

#### Distribuição de projetos genômicos em bactérias



Figura 14: Distribuição de projetos genômicos em bactérias de acordo com sua classificação (https://gold.jgi-psf.org/distribution).





Figura 15: Relevância dos projetos bacterianos divididos de acordo com sua categoria (disponível em https://gold.jgi-psf.org/statistics acesso em 11/06/15).



Figura 16: Projetos bacterianos organizados de acordo com sua filogenia (disponível em https://gold.jgi-psf.org/statistics acesso em 11/06/15).



Depósito de genomas no GOLD segundo o genêro bacteriano

Figura 17: Porcentagem de projetos depositados no GOLD de acordo com seu gênero bacteriano (disponível em https://gold.jgi-psf.org/distribution acesso em 11/06/15).



Annexe 5 : Données génomiques du Genomes Online Database (GOLD)

Figure 13 : Nombre total de projets par an déposés dans le GOLD (https://gold.jgi-psf.org/distribution).

#### Distribution des projets génomiques dans les bactéries



```
FRAGMENTS DU GÉNOME (2,5%)
RE-SEQUENÇAGE (0,8%)
TRANSCRIPTOME (0,2%)
GENOME COMPLETEMENT SEQUENCE (96,3%)
AUTRES (0,2%)
```

Figure 14 : Distribution des projets génomiques dans les bactéries selon leur classement (https://gold.jgi-psf.org/distribution).



Figure 15: La pertinence des projets bactériens présentés selon leur catégorie (disponible en https://gold.jgi-psf.org/statistics accès 06.11.15).

Phylogénie des projets bactériens



Figure 16 : Projets bactériens organisé selon leur phylogénie (disponible en https://gold.jgi-psf.org/statistics accès 06.11.15).



Projets déposés dans GOLD selon leur genre bactérien

Figure 17 : Pourcentage des projets déposés dans GOLD selon leur genre bactérien (disponible en https://gold.jgi-psf.org/distribution accès en 11/6/15).

#### Anexo 6: Manuscrito em redação - Survival strategies of Gram-positive bacteria

# Annexe 6 : Manuscrit en cours - Survival strategies of Gram-positive bacteria

#### Survival strategies of Gram-positive bacteria (Review)

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#### Abstract

Bacteria have been subject of studies, since antiquity, can cause disease to humans, animals and plants, and they can be used for therapeutic and biotechnology applications due to their beneficial properties, such as probiotics. Due to a multitude of information that covers this topic, text mining methods that allow automated extraction of information for the investigation of these organisms have become essential. In this review the text-mining with the LitProf program allowed the selection of a well labored content by combining biological concepts and growth phases and strategies of grampositive bacteria that allow their persistence and viability for longer periods. Further we will discuss the criteria for the classification of bacterial growth phases and the expression of various phenotypes, while passing through a series of physiological and morphological cellular adjustments aimed at maintaining the survival of these populations. Therefore, regulation of bacterial metabolism is needed, for example, nutrient storage. Furthermore, strategies that allow bacterial survival to various stresses, such as programmed cell death and dormancy will be addressed in this discussion. Bacteria that utilize dormancy are also capable of maintaining viable but not culturable in unfavorable situations can be revived and set their conditions for bacterial growth when exposed to favorable conditions again. In the future, the manipulation and regulation of bacterial strategies will allow technological advances in diverse industrial sectors such as agrifood, pharmaceutical, cosmetic, biomedical and forestry.

Keywords: bacteria, Gram-positive, long-term survival, survival strategies.

# Introduction

Bacteria are ubiquitous microorganisms, known for their ability to multiply and adapt in many environments and growth conditions (Roszak, DB, e Colwell, RR, de 1987; Tempest e Wouters, 1981; M. Neijssel, e W. Zevenboom, 1983). For decades, bacterial survival has been a subject of great interest in various fields of microbiology and these studies crosslinked with many other disciplines such as ecology, genetics, bioinformatics, biotechnology, food, epidemiology or infectiology. A better understanding of bacterial survival can be helpful in human, animal or plant health issues as well as in the biotechnology or food industry. With regard to survival capacities, bacteria can indeed be divided into two groups: (1) pathogenic bacteria, that are able to cause infections, and whose threat prompt the need for decontamination procedures, vaccines and drugs and (2) non-pathogenic bacteria, some of which are useful for industrial purpose like the production of biomolecules or the development of probiotic products, which in some applications require a good survival to maintain activity.

Bacterial growth comprises different phases (lag, exponential, stationary, and death) along which the bacterial population copes with continuously changing conditions with, among others, high and low availability of nutrients (Helmus et al., 2012). Bacteria are thus able to switch on or off or to modulate the expression of many mechanisms to ensure their homeostasis.

To overcome harsh conditions, some bacterial species sporulate. This strategy has been widely documented and reviewed in detailed elsewhere (Palop, MañAs and CondóN 1999). Besides, some non-sporulating species are nevertheless able to survive for long periods of time and their growth enters a phase called "long-term survival" (LTS). In 1998, Watson and collaborators defined 'long-term survival' as a period of survival in which bacteria can respond to the stimulus of hunger and remain alive and viable. For other authors, this phase of survival is characterized, in addition to starvation, by a resistance to various stresses (Wen et al. 2011). According to Finkel (Finkel 2006a), this is a period in which the bacteria undergo competition for nutrients and most likely have reduced metabolic capacity and exhibit a particlar phenotype. During the LTS phase, many phenotypic changes can indeed be observed: a reduced metabolism, biofilm formation, changes in shape and size of bacterial cells, changes in size and color of colonies, and or viable but nonculturable phenotype. These changes are part of various strategies bacterial species have evolved to ensure a longer viability.

Depending on the study or the species considered, LTS spans a range of time that goes from hours to years (Zambrano and Kolter 1993; Finkel 2006a).

Several strategies bacteria can be used to survive for long periods, for example by using different substrates available in the medium, followed by the ability to enter a resting state for long periods (with low metabolic activity) or by entering a reversible cell stasis, after which these bacteria can resume cellular multiplication when conditions are favorable (Greenblatt et al. 2004b).

In this review article, we will address the strategies Gram-positive bacteria exert during LTS periods, aiming at their metabolic maintenance and proliferation capacity. Sporulation will not be considered here.

#### Evolution of research dealing with long-term survival

Research works dated from 1967 mark the beginning of the scientific community's interest for bacterial survival in extended periods. Since then, a gradual increase in publications can be observed with this issue in PubMed (http://www.ncbi.nlm.nih.gov/pubmed) (Figure 1).



Figure 1: The data in PubMed results by year in 01/09/2015 (http://www.ncbi.nlm.nih.gov/pubmed/?term=long-term+survival+bacteria).

Among the hundreds of references found in databases, the terminology used to describe LTS is quite variable. A short text mining study applied to the terms associated to LTS was carried out using the LitProf program (Chassaubel and Sher, 2002). The literature profiles were generated by extracting the frequencies of Gram positive- and

LTS-related terms from hundreds of abstracts stored in the PubMed database. The terms were filtered on the basis of both repetitive occurrence and co-occurrence and clustered with regard to their frequency values. This resulted in a picture of the functional relationship among the lists of LTS-related terms (Figure 2). Data analysis revealed strong relationship between the LTS-related terms and the terms associated to the description of growth phases, dormancy, resuscitation, etc. It also clearly shows a link with the ability of bacteria to trigger a series of mechanisms involved in the adaptation in response to adverse environmental factors. Interestingly, the results show associations between bacterial virulence and persistence, which is relevant, e.g. in nosocomial infections, and between cell-lysis products and food, which is relevant, e.g. the involvement of lactic acid bacteria in chesse ripening. Most the associations revealed by this data analysis will be presented in the following paragraphs.



Figure 2: Bow's default parameters, except for F = 0.5, and hierarchical clustering with Pearson correlation and UPGMA to group X (93terms) and Y (60 queries)

axes.

## The phases of bacterial growth

The interest in the investigation and characterization of bacterial growth phases began for almost 100 years. Since 1917, in accordance with fundamental theories in physics and mathematical theories the researchers suggested that bacteria have a growth cycle composed of 7 steps: initial stationary phase, lag phase (or positive growth acceleration phase), logarithmic growth phase, negative growth acceleration, maximum stationary phase, phase of accelerated death and logarithm death phase (Buchanan 1918). Later on, Lane-Claypon simplified the description of bacterial phases to four divisions: initial period of slow growth (or even no growth), period of regular growth, period during which numbers remain stable and period during which the numbers of living bacteria are decreasing (Buchanan 1918). In 2006, Finkel divided the bacterial growth into five phases: lag, exponential (or logarithmic), stationary, death and long-term stationary phase (Finkel 2006a). These final steps will be presented in more detail in the Table 1.

Phases of bacterial growth	Description				
Lag	First stage, which consists in the adaptation of the bacteria to the environment in which it is exposed. In this phase, bacteria adopt a high metabolic rate, and consequently a high protein synthesis for cell maintenance.				
Exponential	Second stage. Micro-organisms tend to consume all the nutrients in the medium to generate an energy reserve, with high activity of the DNA polymerase III, involved in DNA replication.				
Stationary	Third stage, the processes that lead to cell death will begin to appear in some bacterial species. The oxidative stress induces the bacteria to activate a number of mechanisms to maintain viability.				
Death	Stimuli such as a lack of available nutrients or the presence of toxic metabolites in the medium can induce metabolic activities causing cell death in bacteria (unable to multiply).				
Long-Term-Survival	The long-term survival phase presents characteristics that associated with bacterial persistence, including morphological alterations. Bacteria adapt to the environment, and trigger mechanisms to ensure their survival.				



In Figure 3, we briefly present the five important bacterial phases: (1) lag phase, (2) exponential phase, (3) stationary phase, (4) death phase and (5) long-term survival.

Figure 3: Bacterial growth shows about the same phases but the time scale ranges from hoursto years depending on the species.

#### Bacterial phenotypes in long term survival

## Cellular and colony morphological changes

During the LTS, Gram positive bacteria can undergo morphological changes, at the cellular or the colony level (Table 2).

In bacteria, nutrient availability has a great impact on bacterial cell size (Chien et al., 2012). Under stress conditions such as starvation encountered in LTS phase, some Gram positive bacteria can enter into a cell-wall deficient state also known as L-form conversion. This phenomenon is well-described in pathogenic bacteria of Mycobacterium genus. In *M. bovis* (Slavchev; Michailoca; Markova, 2013), or *M. tuberculosis* (Markova et al., 2012), L-form conversion was shown to be an adaptive strategy to survive and reproduce under unfavorable conditions. In other Gram positive species, a reduction in cell size is observed when the population enters the LTS phase. This was shown in *S. aureus* which indeed harbours a decrease in cell size in starvation survival. This phenotype is accompanied by an increase in resistance to acid shock and oxidative stress (Watson et al., 1998). Similarly, in *Listeria monocytogenes* cellular

morphology can change from bacilli to cocci during the transition to LTS phase, leading to an increased baro- and thermotolerance (Wen et al., 2011). Morphological changes were also observed during acid adaptation in the food grade bacterium Propionibacterium freudenreichii, with a shift from pleomorphic rods arranged in Chinese characters to bacteria with a segmented aspect and shorter cells (Jan et al., 2001).

Alterations of phenotype at the cell level can also have an impact at the colony level, namely in the size and color of the colonies (Helmus et al. 2012). A change in the size of colonies and a decline of the growth rate can be observed in *Bacillus subtilis*, during nutrient shortage (Chien; Hill; Levin, 2012). In *Streptococcus pyogenes*, after a long period (4 to 14 weeks) in stationary phase and nutrient shortage, culturable cells formed two colony phenotypes: atypical large colonies and microcolonies, which showed extensive changes at the proteome level compared to the parental strain (Wood et al., 2005).

Bacteria	Morphological alterations	Reference	
Listeria monocytogenes	The structure and the form of the cells change during the period of transition from early stationary phase to long-term survival	(Wen <i>et al.</i> 2011)	
Mycobacterium bovis	In stressful conditions, <i>Mycobacterium bovis</i> displays different morphologies, such as L- forms (cell wall-deficient cells), as a way to survive in unfavorable environments	(Slavchev, Michailova and Markova 2013)	
Staphylococcus aureus	Changes in morphology with a decrease in cell size between the hunger and survival phases	(Watson, Clements and Foster 1998)	
Streptococcus pyogenes	After four weeks in stressful conditions, two different colony phenotypes are observed, most likely due to some selective pressure of the environment in two phases of life cycle: atypical large colonies and small colonies	(Wood 1981)	

Table 2:	Morphological	alterations in	bacterial	cells or	colonies.
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#### **Growth rate changes**

Bacteria were early shown to be able to sense and recognize unfavorable conditions in the environment and thus to enter into a state where bacterial replication is inhibited until favorable conditions are re-established (in Wayne and Sohaskey, 2001: HJ Corper, Cohn ML, 1933). This capacity is an important strategy that allows bacterial survival. In Bacillus subtilis, alterations in cell division are directly correlated with nutrient availability and growth rate (Chien, Hill and Levin, 2012). To store energy, which correspond to a reduction in some metabolic pathways, such as the glycolytic pathway, in turn of decreasing the production of ATP (Savic and McShan 2012). As we can see other species, after a stress period, still require a period of latency for the morphological changes to occur, even after reestablishing favorable conditions (Roszak and Colwell 1987). This is because a part of the bacterial population needs more time to recover and return to being competitive after being exposed to the period of stress. The study of Geisel and colleagues shows the relationship between stress resistance and growth rate of various microorganisms. This study revealed that the cells remain in a vegetative state or enter into the dormant state, to decrease bacterial metabolism under unfavorable conditions, which allows these bacteria to adapt and survive in the environment (Geisel, Vilar and Rubi 2011).

#### **Genetic adaptation**

In some bacterial species, LTS phase induces a growth advantage in stationary phase (GASP) phenomenon. GASP is defined as fluctuations in viable cell numbers during stationary phase growth, with a rapid loss of viability followed by an increase in viable counts (see above, Figure 3). In the stationary phase, the bacterial cells that undergo the GASP phenomenon are able to grow displace, the parent cells and become the dominant population in the culture. It is triggered by intense competition for nutrients and actually reflects the prevalent state of bacteria in their natural environment. GASP phenomenon is due to a series of genetic changes, occurring while the bacterial population enters the stationary phase. In parallel, physiological and structural alterations occur and induce a higher tolerance to diverse environmental stresses (Sewell et al., 2011). It was first reported and is well-documented in *E. coli*, the Gram negative model bacterium (Zinser, Erik R, Kolter, 2004; Zinster and Kolter, 1999; Bacun-Druzina et al. 2011). The GASP cells are able to transfer their advantage to

daughter cells, providing a competitive advantage (Finkel 2006a), in low concentrations of nutrients or in physical and chemical conditions that are unfavorable for growth. In contrast, only a few studies report on GASP phenomenon in Gram positive bacteria. One of the GASP associated phenotypes, namely the drop in cell counts at the beginning of stationary phases followed by an increase in cell counts, was shown in *Enterococcus feacium*. GASP however seems to be species-specific since it was not observed in *S. aureus* cultures (Sewell et al., 2011). In *Bacillus subtilis*, a phenomenon comparable to GASP was observed and linked to a hypermutable subpopulation that emerge during the stationary phase. This phenomenon, also called stationary–phase mutagenesis involves *comA* and *comK* genes (associated to prokaryotic differentiation) and is *recA*-independent. It leads to mutations that arise during the stationary-phase, in bacterial cells that do not divide and that are subjected to non lethal stresses, such as nutrient shortage (Sung and Yasbin, 2002).

#### **Regulation of bacterial metabolism**

The maintenance and regulation of bacterial metabolism requires the action of several proteins involved in functions, such as transport and synthesis. In this section, we will highlight the factors that allow the control of these activities for the use of alternative energy sources, and the storage of nutrients and for the maintenance of intracellular homeostasis.

#### Use of atypical nitrogen and carbon source

When the entry in LTS is due to nutritional shortage, some species adapt their metabolism to use other carbon and nitrogen sources. Such adaptation can be operated through the use of acetate or citrate as an alternative electron donor to produce energy, as shown in the Gram negative bacterium *Geobacter sulfurreducens* (Helmus et al. 2012). In Gram positive bacteria, *M. luteus* can use succinic acid as a substrate to produce energy. *Lactobacillus sakei* survival is linked to the degradation of arginine (Champomier Vergès et al. 1999), while *L. monocytogenes* can consume products from cell lysis to remain viable (Wen et al. 2013).

## **Nutrient Storage**

Some species of Gram-positive bacteria store various compounds, such as a polysaccharide, poly- -hydroxybutyrate, lipids, polyphosphate or other elements, which

can be used to maintain their viability during prolonged periods of nutrient shortage (Wilkinson 1959).

In general, when bacterial cells are subjected to nutrient shortage in carbon, nitrogen and phosphorus, "survival" responses are induced. In such conditions, bacteria can use cell reserves like glycogen or polyphosphate (Carvalheira et al. 2014) and/or cell lysis products in the LTS population (Roszak and Colwell 1987). Inorganic polyphosphate (polyP) is critical to bacterial survival in adverse conditions (Rao, Gómez-García and Kornberg 2009) as shown in *Mycobacterium sp*. During phosphate shortage, *Mycobacterium tuberculosis* induces the *ppk*1 gene, which encodes a polyphosphate kinase and *rel*A, which boosts the response to these stress so that the bacterial cells remain viable (Rifat, Bishai and Karakousis 2009).

In environments that do not present a broad range of available nutrients, bacteria are able to store some elements to prepare their survival phase, as shown in *Vibrio cholera* and *Mycobacterium tuberculosis*, which can store glucose in the form of glycogen (Bourassa and Camilli 2009; Amit Singh 2014). They can also do so when they are subjected to various stresses (nitrogen intermediates, reactive oxygen, low pH, and vital element starvation). When the carbon substrate is exhausted in the medium, these reserve substrates are utilized as energy source (Amit Singh 2014).

In LTS transition, *L. monocytogenes* upregulates the expression of genes related to energy metabolism, cell envelope structure, and transport. The upregulation of compatible solute transporters is thought to be involved in the accumulation of cellular solutes, which in turn lowers intracellular water activity and thus increases stress resistance (Greenblatt et al. 2004b; Wen et al. 2011).

# Maintenance of pH and osmoprotection

Bacterial adaptation to nutrient shortage results in the increase of some substrates, which can also be harmful to the cell. An increase in intracellular glucose can decrease bacterial survival as this will directly lower the pH of the medium because the bacteria cannot metabolize the large amounts of lactic acid produced. Some *S. pyogenes* strains are able to survive for long periods most likely thanks to mechanisms that allow them to escape these unfavorable conditions and remain viable, either by buffering systems or a similar mechanism for the maintenance of pH level (Savic and McShan 2012).

In *L. monocytogenes*, storage of osmoprotectants like glycine, betaine and trehalose allows viability. Also, upregulation of the transporter genes and the expression of genes involved in energy metabolic pathways is crucial for survival (Greenblatt et al. 2004a; Wen et al. 2011).

## **Different strategies for survival**

In this section, we discuss the best known mechanisms involved in LTS in Gram-positive bacteria, such as programmed cell death (PCD) and dormancy, which may help us understand this puzzle.

#### **Programmed cell death**

In Gram-positive bacteria, concepts such as programmed cell death have begun to emerge as a mechanism that allow bacterial populations to survive. Some authors suggest this as an evolutionary process that has allowed bacteria to adapt. Dynamic cellular stress responses cause destabilization bacterial population in varying conditions by requiring activation of quorum-sensing systems to trigger programmed cell death (PCD) (Finkel 2006c).

Quorum-sensing is observed in Gram-positive bacteria as a facilitator in response to stress conditions. The perception of stimuli in the medium activates specific signaling pathways, which initiates the process (Bharati et al. 2012). A large number of quiescent cells are responsible for the storage of carbon. In *L. monocytogenes*, this mechanism of quorum-sensing has been proposed as a trigger for activities that balance growth and death in the population by regulating gene expression between periods of long-term survival. This is possible because these bacteria are able to perceive the conditions to initiate programmed cell death, increasing the availability of nutrients to the surviving cells (Wen et al. 2013).

In addition to this mechanism, in this review, we will also focus on another very important phenomenon: dormancy.

#### Dormancy

Bacteria can to regulate their metabolic activity in order to maintain the viability of the population in periods without or without low cell multiplication and in periods of competition in extreme environments with low availability of nutrients (Ratcliff et al.
2013). This step of dormancy or quiescence corresponds to a "resting period" or a "period of reversible phenotypic change" and is triggered in response to extreme conditions of growth (Roszak and Colwell, 1987). It has been well-studied in pathogenic bacteria where, during an infection, the resulting slowly growing bacteria are able to tolerate both the host immune response and prolonged antibiotic exposure. This is typically the case in chronic infections such as tuberculosis caused by the Gram positive pathogen *M. tuberculosis*.

Dormancy (or quiescence) may be defined as a state of resistance to nonfavorable conditions, or stresses where the viable population size is maintained throughout the period of stress, and where bacteria maintain a metabolic capacity, their membrane potential, and do not undergo obvious morphological differentiation (Rittershaus, Baek and Sassetti 2013a). During the dormancy period, cell replication and DNA synthesis are blocked, while the synthesis of RNA and proteins continues, so that bacterial metabolic activity remains viable until the stress is over (Roszak and Colwell, 1987). Besides M. tuberculosis, *Micrococcus luteus* was shown to enter in dormancy over long periods when subjected to starvation (Greenblatt et al. 2004b).

#### Viable but nonculturable state

The entry in a Viable but nonculturable (VBNC) state is a strategy in response to adverse conditions described in various bacteria in situations of nutrient stress or atypical temperatures (Mulyukin et al. 2003). This concept emerged a few decades ago as an explanation for how bacteria can survive for long periods in adverse conditions (Roszak and Colwell, 1987). The VBNC state can be either a state in which bacteria await death or where they wait for a stimulus so that cellular metabolic activities can return to normal (Mulyukin et al. 2003).

It was found that in many species of pathogenic bacteria of VBNC state is present, so that they are able to express their virulence and remain viable. Furthermore, it was shown that bacteria have been VBNC the resuscitation promoting factor (Ramamurthy et al 2014).

#### Resuscitation

Different methodologies that have been carried out in order to observe and characterize the phenomenon of "resuscitation bacterial", seeks to restore conditions to stabilize the population using suitable media, pH and temperature (Roszak and Colwell, 1987).

To get out of the state of dormancy, bacteria can initiate a form of recovery after a period of latency as reported in *M. luteus* (Greenblat et al. 2004b). This phase is named resuscitation and is also observed in *S. aureus*, which however requires a large amount and variety of nutrients to resume its metabolic activities (Diaper and Edwards 1994).

Resuscitation Promoting Factor (Rpf) is present in most bacteria with high G + C content. The Rpf are a group of secreted proteins, that belong to 29 families in genomes of actinomycetes (Dewi Puspita et al. 2013). In *M. tuberculosis*, it has been reported that anti-rpf antibodies are able to inhibit bacterial growth. Similarly, exogenous rpf added to old cultures leads to a reactivation of bacterial growth. Other models of infection *in vitro* were made and guaranteed performance of this group of proteins for resuscitation of cells (Kaprelysants et al. 2012).

Studies show that the analysis of the transcriptome of "resuscitated" cells of *L. monocytogenes* suggests an alteration in the metabolism to bacteria in their exponential phase, clearly showing that the bacterium is able to go through different conditions and remain viable (Wen et al. 2011). For this, it is necessary that genes associated with the synthesis of cell envelope components, proteins, amino acids and ribonucleotides are rapidly expressed (Wen et al. 2011).

#### What is to come? (Future directions)

To date, most studies on long-term survival were carried out using classical microbiology techniques and mostly described the consequences of their phenotypes (size, color, persistence). However, is it possible, at a genomic level, to predict the ability of a bacterium to survive based on its gene content or based on the bacterial metabolic pathways?

New sequencing technologies and the advent of omic technologies, such as genomics, transcriptomics, proteomics and metagenomics, open avenues for research on the survival and adaptability of Gram-positive bacteria. Integrating information from different fields will allow to better understand how these mechanisms are regulated. This is of major interest to develop new products such as drugs, vaccines, or select more robust probiotics.

New research in bacterial survival is of great interest e.g. for the design of therapies that interfere with pathogenesis and persistence of bacteria, which are major concerns in clinical microbiology and food safety. A better understanding of LTS mechanisms could lead to the development of drugs that act even on dormant bacterial cells and impair the establishment of LTS in a pathological context, or in indwelling devices in hospitalized patients. Another application is to enhance LTS to optimize the efficiency of probiotic bacteria since some probiotic species are reportedly fragile (e.g. Bifodobacterium sp.) although their application requires a good survival during manufacturing and storage of products until the end consumer.

#### **Author and Contributors**

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Zinser, Erik R; Kolter, Roberto. Mutations enhancing amino acid catabolism confer a growth advantage in stationary phase. Journal of Bacteriology, PMID: 10482523PMCID: PMC94102, v. 181, n. 18, p. 5800–5807, set. 1999.

Anexo 7: Curriculum vitae

Annexe 7 : Curriculum vitae

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## Personal Information

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# Formal Education

2011 - 2015	Doctorate in Bioinformatics.
	Universidade Federal de Minas Gerais, UFMG, Belo Horizonte,
	Brazil
	(Co-guardianship) Institut National de la Recherche Agronomique
	(Advisor: Yves Le Loir)
	Title: Mecanismos moleculares de sobrevivência em longo prazo em
	Propionibacterium freudenreichii
	Advisor: Vasco Ariston de Carvalho Azevedo
	Scholarship from: Coordenação de Aperfeiçoamento de Pessoal de
	Nível Superior/COFECUB
	Knowledge areas: Bioinformatics and Molecular Genetics in
	Microorganisms.
2007 - 2011	Bachelor's in Biomedicine.
	Universidade Fundação Mineira de Educação e Cultura, FUMEC,
	Belo Horizonte, Brasil
	Study abroad sandwich option: Université de Provence Aix Marseille I
	(Advisor : Jean-Pierre Roll)
	Title: Teste do efeito dos probióticos frente à Pseudomonas
	aeruginosa
	Advisor: Adriana dos Santos e Flávio Henrique Barbosa

# Complementary Education

2014 - 2014	Short Course in Advanced seq annotation and use of pipelines EGene2. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2014 - 2014	Short Course in PATRIC. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2014 - 2014	Short Course in Genomics for Working with Pathogens. EMBL European Bioinformatics Institute, EMBL-EBI, UFMG, Belo Horizonte, Brasil
2014 - 2014	Short Course in GO terms and annotations. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2012 - 2012	Short Course in Introduction to BioNumerics program. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2012 - 2012	Short Course in Introduction to Perl languages. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2012 - 2012	Short Course in Molecular Methods for Microbial Diagnosis, Genomic. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2012 - 2012	Short Course em Rna-seq course. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2012 - 2012	Short Course em International Course in Automated Functional Annot. Centro de Investigaciones Príncipe Felipe, CIPF, Espanha
2011 - 2011	Short Course in Next Generation Sequencing: SOLiD. Universidade Federal do Paraná, UFPR, Curitiba, Brasil
2011 - 2011	Short Course in assembly and annotation of bacterial genomes. Universidade Federal de Minas Gerais, UFMG, Belo Horizonte, Brasil
2011 - 2011	Short Course in II Winter Course of Biochemistry and Molecular Biology. Universidade Federal do Paraná, UFPR, Curitiba, Brasil
2011 - 2011	Short Course in molecular biology transformation. Universidade Federal do Paraná, UFPR, Curitiba, Brasil
2011 - 2011	Short Course in IV Winter Course of Genetics. Universidade Federal do Paraná, UFPR, Curitiba, Brasil
2010 - 2010	Short Course in Stage intensif de Langue française.

Université de Provence Aix Marseille I, Aix-Marceille I, Marseille , França

- **2009 2009** Short Course in Arterial Blood Gas Analysis. Medical Association of Minas Gerais, AMMG, Brazil, AMMG, Brasil
- **2008 2009** French. Alliance Française, BH, Brazil.

### Areas of expertise

- 1. Bioinformatics
- 2. Microbiology
- 3. Genetics
- 4. Parasitology

## Languages

- 1. English Understanding Functional, Speaking Functional, Writing Basic, Reading Fluent
- 2. Spanish Understanding Functional, Speaking Functional, Writing Basic, Reading Fluent
- 3. French Understanding Fluent, Speaking Functional, Writing Functional, Reading Fluent

## Prêmios e títulos

2013 BioVis 2013 - Data Contest Biology Experts Pick, IEEE Computer Society

*S,T &A Production* Bibliographic Production Articles Published in Scientific Journals

1. BARBOSA, EUDES G. V., **ABURJAILE, F. F.**, RAMOS, ROMMEL T.J., CARNEIRO, ADRIANA R., LOIR, Y. L., BAUMBACH, J., MIYOSHI, ANDERSON, SILVA, ARTUR, AZEVEDO, VASCO. Value of a newly sequenced bacterial genome. World Journal of Biological Chemistry., v.5, p.161 - 168, 2014.

2. SOARES, SIOMAR C., SILVA, ARTUR, TROST, EVA, BLOM, JOCHEN, RAMOS, ROMMEL, CARNEIRO, ADRIANA, ALI, AMJAD, SANTOS, ANDERSON R., PINTO, ANNE C., DINIZ, CARLOS, BARBOSA, EUDES G. V., DORELLA, FERNANDA A., **ABURJAILE, FLÁVIA**, ROCHA, FLÁVIA S., NASCIMENTO, KARINA K. F., GUIMARÃES, LUÍS C., ALMEIDA, SINTIA, HASSAN, SYED S., BAKHTIAR, SYEDA M., PEREIRA, ULISSES P., ABREU, VINICIUS A. C., SCHNEIDER, MARIA P. C., MIYOSHI, ANDERSON, TAUCH, ANDREAS, AZEVEDO, VASCO. The Pan-Genome of the Animal Pathogen Corynebacterium pseudotuberculosis Reveals Differences in Genome Plasticity between the Biovar ovis and equi Strains. Plos One., v.8, p.e53818-e53818, 2013.

3. DORELLA, FERNANDA A, GALA-GARCIA, ALFONSO, PINTO, ANNE C, SARROUH, BOUTROS, ANTUNES, CAMILA A, RIBEIRO, DAYANA, **Aburjaile, Flávia**, FIAUX, KARINA K, Guimarães, Luis C, SEYFFERT, NUBIA, EL-AOUAR, RACHID A, SILVA, RENATA, HASSAN, SYED S, CASTO, THIAGO LP, MARQUES, WANDERSON S, RAMOS, ROMMEL, CARNEIRO, ADRIANA, SÁ, PABLO DE, MIYOSHI, ANDERSON, AZEVEDO, VASCO, SILVA, ARTUR. Progression of 'OMICS' methodologies for understanding the pathogenicity of Corynebacterium pseudotuberculosis: the Brazilian experience. Computational and Structural Biotechnology Journal., v.6, p.1 - 7, 2013.

4. RAMOS, ROMMEL THIAGO JUCÁ, CARNEIRO, ADRIANA RIBEIRO, SOARES, SIOMAR DE CASTRO, SANTOS, ANDERSON RODRIGUES DOS, ALMEIDA, SINTIA, GUIMARÃES, LUIS, **FIGUEIRA, FLÁVIA**, BARBOSA, EUDES, TAUCH, ANDREAS, AZEVEDO, VASCO, SILVA, ARTUR. Tips and tricks for the assembly of a Corynebacterium pseudotuberculosis genome using a semiconductor sequencer. Microbial Biotechnology (Online). , v.n/a, p.n/a - n/a, 2012.

5. Soares, Siomar C., Trost, Eva, Ramos, Rommel T.J., Carneiro, Adriana R., Santos, Anderson R., Santos, A.R., Pinto, Anne C., **ABURJAILE, F. F.**, Barbosa, Eudes, Ali, Amjad, DINIZ, CARLOS A.A., Hassan, Syed S., FIAUX, KARINA, Guimarães, Luis C., Bakhtiar, Syeda M., PEREIRA, ULISSES, Almeida, Sintia S., Abreu, Vinícius A.C., ROCHA, FLÁVIA S., DORELLA, FERNANDA A., Miyoshi, Anderson, Silva, Artur, Azevedo, Vasco, Tauch, Andreas. Genome sequence of Corynebacterium pseudotuberculosis biovar equi strain 258 and prediction of antigenic targets to improve biotechnological vaccine production. Journal of Biotechnology., v.166, p.1 - 7, 2012.

6. SILVA, A., LOPES, T., RAMOS, R. T. J, CARNEIRO, A., DORELLA, F., Rocha, F. S., SANTOS, A. R., LIMA, A. R. J., GUIMARAES, L. C., Barbosa, E. G. V., RIBEIRO, D., FIAUX, K., DINIZ, C. A. A., ABREU, V. A. C., ALMEIDA, S. S., HASSAN, S., ALI, A., BAKHTIAR, S., **ABURJAILE, F. F.**, PINTO, A. C., Soares, S. C., PEREIRA, U., Schneider, M. P. C, MIYOSHI, A., EDMAN, J., SPIER, S, AZEVEDO, V.. Complete genome sequence of Corynebacterium pseudotuberculosis Cp267 strain isolated from a Llama. Journal of Bacteriology (Print)., v.194, p.3567 - 3568, 2012.

7. Lopes, T., Silva, A., Thiago, R., Carneiro, A., Dorella, F. A., Rocha, F. S., dos Santos, A. R., Lima, A. R. J., Guimaraes, L. C., Barbosa, E. G. V., BARBOSA, E. G. V., Ribeiro, D., Fiaux, K. K., Diniz, C. A. A., de Abreu, V. A. C., de Almeida, S. S., Hassan, S. S., Ali, A., **ABURJAILE, F. F.**, Bakhtiar, S. M., Pinto, A. C., Soares, S. d. C., Pereira, U. d. P., Schneider, M. P. C., Miyoshi, A., Edman, J., Spier, S., Azevedo, V. Complete Genome Sequence of Corynebacterium pseudotuberculosis Strain Cp267, Isolated from a Llama. Journal of Bacteriology (Print)., v.194, p.3567 - 3568, 2012.

8. Pethick, F. E., Lainson, A. F., YAGA, R., FLOCKHART, A., Smith, D. G. E., DONACHIE, W., Cerdeira, L. T., Silva, A., BOL, E., LOPES, T. S., Barbosa, M. S., PINTO, A. C., dos Santos, A. R., SOARES, S. C., Almeida, S. S., Guimaraes, L. C.,

**ABURJAILE, F. F.**, Abreu, V. A. C., RIBEIRO, D., FIAUX, K. K., Diniz, C. A. A., BARBOSA, E. G. V., Pereira, U. P., Hassan, S. S., Ali, A., Bakhtiar, S. M., DORELLA, F. A., DORELLA, F. A., CARNEIRO, A. R., Ramos, R. T. J., Rocha, F. S., Schneider, M. P. C., MIYOSHI, A., AZEVEDO, V., Fontaine, M. C. Complete Genome Sequence of Corynebacterium pseudotuberculosis Strain 1/06-A, Isolated from a Horse in North America. Journal of Bacteriology (Print). , v.194, p.4476 - 4476, 2012.

9. Pethick, F. E., Lainson, A. F., Yaga, R., Flockhart, A., Smith, D. G. E., Donachie, W., CERDEIRA, L. T., SILVA, A., Bol, E., LOPES, T. S., Barbosa, M. S., PINTO, A. C., dos Santos, A. R., SOARES, S. C., ALMEIDA, S. S., Guimaraes, L. C., **ABURJAILE**, **F. F.**, ABREU, V. A. C., RIBEIRO, D., Fiaux, K. K., Diniz, C. A. A., BARBOSA, E. G. V., Pereira, U. P., Hassan, S. S., ALI, A., Bakhtiar, S. M., DORELLA, F. A., CARNEIRO, A. R., RAMOS, R. T. J., Rocha, F. S., SCHNEIDER, M. P. C., MIYOSHI, A., AZEVEDO, V., AZEVEDO, V., Fontaine, M. C. Complete Genome Sequences of Corynebacterium pseudotuberculosis Strains 3/99-5 and 42/02-A, Isolated from Sheep in Scotland and Australia, Respectively. Journal of Bacteriology (Print). , v.194, p.4736 - 4737, 2012.

10. Santos, A.R., Carneiro, A. R., Ramos, R. T. J., DALL'AGNOL, H., PINTO, A. C., de Castro Soares, S., SANTOS, A. R., Guimaraes, L. C., Almeida, S. S., BARAUNA, R. A., DAS GRACAS, D. A., FRANCO, L. C., Ali, A., Hassan, S. S., NUNES, C. I. P., Barbosa, M. S., **ABURJAILE, F. F.**, Fiaux, K. K., Barbosa, E. G. V., Bakhtiar, S. M., VILELA, D., NOBREGA, F., DOS SANTOS, A. L., CAREPO, M. S. P., Azevedo, V., Schneider, M. P. C., PELLIZARI, V. H., SILVA, A. Genome Sequence of Exiguobacterium antarcticum B7, Isolated from a Biofilm in Ginger Lake, King George Island, Antarctica. Journal of Bacteriology (Print)., v.194, p.6689 - 6690, 2012.

11. SANTOS, A. R., CARNEIRO, A. R., GALA-GARCIA, A., PINTO, A. C., BARH, D., BARBOSA, E. G. V., **ABURJAILE, F. F.**, DORELLA, F. A., ROCHA, F. S., GUIMARAES, L. C., Zurita-Turk, Meritxell, RAMOS, R. T. J., ALMEIDA, S. S., ALMEIDA, S., SOARES, S. C., PEREIRA, U. P., ABREU, V. A. C., SILVA, A., MIYOSHI, A., AZEVEDO, V.. The Corynebacterium pseudotuberculosis in silico predicted pan-exoproteome. BMC Genomics., v.13, p.S5 - S6, 2012.

12. CERDEIRA, L., Schneider, M. P. C, PINTO, A. C., ALMEIDA, S. S., SANTOS, A. R., Barbosa, E. G. V., ALI, A., **ABURJAILE, F. F.**, ABREU, V. A. C., GUIMARAES, L. C., Soares, S. C., DORELLA, F., Rocha, F. S., BOL, E., LOPES, T., Barbosa, M. S., CARNEIRO, A., Juca Ramos, R. T., Coimbra, N. A. d. R, LIMA, A. R. J., Barh, D, Tiwari, S., RAJA, R., ZAMBARE, V., Ghosh, P., Trost, E., Tauch, A, MIYOSHI, A., AZEVEDO, V., SILVA, A.. Complete Genome Sequence of Corynebacterium pseudotuberculosis Strain CIP 52.97, Isolated from a Horse in Kenya. Journal of Bacteriology (Print)., v.193, p.7025 - 7026, 2011.

13. HASSAN, SYED SHAH, GUIMARÃES, LUIS CARLOS, PEREIRA, ULISSES DE PÁDUA, ISLAM, ARSHAD, ALI, AMJAD, BAKHTIAR, SYEDA MARRIAM, RIBEIRO, DAYANA, RODRIGUES DOS SANTOS, ANDERSON, SOARES, SIOMAR DE CASTRO, DORELLA, FERNANDA, PINTO, ANNE CYBELLE, SCHNEIDER, MARIA PAULA CRUZ, BARBOSA, MARIA SILVANIRA, ALMEIDA, SÍNTIA, ABREU, VINÍCIUS, **ABURJAILE, FLÁVIA**, CARNEIRO, ADRIANA RIBEIRO, CERDEIRA, LOUISE TEIXEIRA, FIAUX, KARINA, BARBOSA, EUDES, DINIZ, CARLOS, ROCHA, FLAVIA S., RAMOS, ROMMEL THIAGO JUCÁ, JAIN, NEHA, TIWARI, SANDEEP, BARH, DEBMALYA, MIYOSHI, ANDERSON, MÜLLER, BORNA, SILVA, ARTUR, AZEVEDO, VASCO. Complete genome sequence of Corynebacterium pseudotuberculosis biovar ovis strain P54B96 isolated from antelope in South Africa obtained by rapid next generation sequencing technology. STAND GENOMIC SCI., v.7, p.189 - 199, 2012.

14. SILVA, A., RAMOS, R. T. J., RIBEIRO CARNEIRO, A., CYBELLE PINTO, A., de Castro Soares, S., RODRIGUES SANTOS, A., SILVA ALMEIDA, S., Guimaraes, L. C., **FIGUEIRA ABURJAILE, F.**, VIEIRA BARBOSA, E. G., ALVES DORELLA, F., SOUZA ROCHA, F., SOUZA LOPES, T., KAWASAKI, R., GOMES SA, P., DA ROCHA COIMBRA, N. A., TEIXEIRA CERDEIRA, L., SILVANIRA BARBOSA, M., CRUZ SCHNEIDER, M. P., MIYOSHI, A., SELIM, S. A. K., MOAWAD, M. S., AZEVEDO, V. Complete Genome Sequence of Corynebacterium pseudotuberculosis Cp31, Isolated from an Egyptian Buffalo. Journal of Bacteriology (Print)., v.194, p.6663 - 6664, 2012.

15. Ramos, R. T. J., Silva, A., CARNEIRO, A. R., PINTO, A. C., Soares, S. d. C., Santos, A. R., Almeida, S. S., Guimaraes, L. C., **ABURJAILE, F. F.**, BARBOSA, E. G. V., DORELLA, F. A., DORELLA, F. A., Rocha, F. S., Cerdeira, L. T., Barbosa, M. S., Tauch, A., EDMAN, J., SPIER, S., MIYOSHI, A., Schneider, M. P. C., AZEVEDO, V.. Genome Sequence of the Corynebacterium pseudotuberculosis Cp316 Strain, Isolated from the Abscess of a Californian Horse. Journal of Bacteriology (Print). , v.194, p.6620 - 6621, 2012.

16. Hassan, S. S., SCHNEIDER, M. P. C., RAMOS, R. T. J., CARNEIRO, A. R., RANIERI, A., Guimaraes, L. C., ALI, A., Bakhtiar, S. M., Pereira, U. d. P., dos Santos, A. R., Soares, S. d. C., DORELLA, F., PINTO, A. C., RIBEIRO, D., Barbosa, M. S., ALMEIDA, S., ABREU, V., **ABURJAILE, F.**, FIAUX, K., BARBOSA, E., DINIZ, C., ROCHA, F. S., SAXENA, R., Tiwari, S., ZAMBARE, V., Ghosh, P., PACHECO, L. G. C., DOWSON, C. G., KUMAR, A., BARH, D., MIYOSHI, A., AZEVEDO, V., SILVA, A.. Whole-Genome Sequence of Corynebacterium pseudotuberculosis Strain Cp162, Isolated from Camel. Journal of Bacteriology (Print). , v.194, p.5718 - 5719, 2012.

17. OLIVEIRA, L. C., SARAIVA, T. D. L., SOARES, S. C., RAMOS, R. T. J., SA, P. H. C. G., CARNEIRO, A. R., MIRANDA, F., FREIRE, M., RENAN, W., JUNIOR, A. F. O., SANTOS, A. R., PINTO, A. C., SOUZA, B. M., CASTRO, C. P., Diniz, C. A. A., ROCHA, C. S., MARIANO, D. C. B., DE AGUIAR, E. L., FOLADOR, E. L., Barbosa, E. G. V., **ABURJAILE, F. F.**, GONCALVES, L. A., GUIMARAES, L. C., AZEVEDO, M., AGRESTI, P. C. M., SILVA, R. F., Tiwari, S., ALMEIDA, S. S., Hassan, S. S., PEREIRA, V. B., ABREU, V. A. C., Pereira, U. P., Dorella, F. A., CARVALHO, A. F., PEREIRA, F. L., LEAL, C. A. G., FIGUEIREDO, H. C. P., SILVA, A., MIYOSHI, A., AZEVEDO, V.. Genome Sequence of Lactococcus lactis subsp. lactis NCDO 2118, a GABA-Producing Strain. Genome Announcements. , v.2, p.e00980-14 - e00980-14, 2014.

SANTOS. ANDERSON R, CARNEIRO, ADRIANA, GALA-GARCÍA, 18. EUDES. ALFONSO, PINTO, ANNE, BARH, DEBMALYA, BARBOSA, FLÁVIA, DORELLA, FERNANDA, ROCHA. FLÁVIA, ABURJAILE, GUIMARÃES, LUIS, Zurita-Turk, Meritxell, RAMOS, ROMMEL, ALMEIDA, SINTIA, SOARES, SIOMAR, PEREIRA, ULISSES, ABREU, VINÍCIUS C, SILVA, ARTUR, MIYOSHI, ANDERSON, AZEVEDO, VASCO. The Corynebacterium pseudotuberculosis in silico predicted pan-exoproteome. BMC Genomics. , v.13, p.S6, 2012.

19. ABURJAILE, S. B., **ABURJAILE, F. F.**, BARBOSA, F. H. F.;Barbosa, Flávio H. F.;BARBOSA, F. Variação da concentração de minerais em aquíferos confinados de indústria de exploração de águas minerais naturais no município de Aparecida-SP. Ciência Equatorial., v.1, p.13 - 17, 2011.

20. BARBOSA, F. H. F.;Barbosa, Flávio H. F.;BARBOSA, F, BARBOSA, L., BAMBIRRA, Leandro Henrique Silva, **ABURJAILE, F. F.** Produção de substâncias envolvidas no fenômeno de antagonismo bacteriano. Revista de Biologia e Ciências da Terra., v.11, p.1 - 10, 2011.

21. BARBOSA, F. H. F.;Barbosa, Flávio H. F.;BARBOSA, F, BARBOSA, L., BAMBIRRA, Leandro Henrique Silva, **ABURJAILE, F. F.** Probióticos microrganismos a favor da vida. Revista de Biologia e Ciências da Terra. , v.11, p.11 - 21, 2011.

22. PEREIRA, ULISSES DE PÁDUA, RODRIGUES DOS SANTOS, ANDERSON, HASSAN, SYED SHAH, **ABURJAILE, FLÁVIA FIGUEIRA**, SOARES, SIOMAR DE CASTRO, RAMOS, ROMMEL THIAGO JUCÁ, CARNEIRO, ADRIANA RIBEIRO, GUIMARÃES, LUÍS CARLOS, SILVA DE ALMEIDA, SINTIA, DINIZ, CARLOS AUGUSTO ALMEIDA, BARBOSA, MARIA SILVANIRA, GOMES DE SÁ, PABLO, ALI, AMJAD, BAKHTIAR, SYEDA MARRIAM, DORELLA, FERNANDA ALVES, ZERLOTINI, ADHEMAR, ARAÚJO, FLÁVIO MARCOS GOMES, LEITE, LAURA RABELO, OLIVEIRA, GUILHERME, MIYOSHI, ANDERSON, SILVA, ARTUR, AZEVEDO, VASCO, FIGUEIREDO, HENRIQUE CÉSAR PEREIRA. Complete genome sequence of Streptococcus agalactiae strain SA20-06, a fish pathogen associated to meningoencephalitis outbreaks. Standards in Genomic Sciences., v.8, p.188 - 197, 2013.

### Articles accepted for publication

1. **ABURJAILE, FF**, MADEC, M., PARAYRE, S., AZEVEDO, V., MIYOSHI, A., LE LOIR, Y., FALENTIN, H.. *Propionibacterium freudenreichii* long-term survival in nutrient shortage. Journal of Applied Microbiology (Print). , 2015.

## Book chapters published

1. Roukos, Dimitrios H, SILVA, ARTUR, Carneiro, Adriana R, **Aburjaile, Flávia**, Guimarães, Luis C, Ramos, Rommel TJ, Castro, Thiago LP, Abreu, Vinicius, Silva, Wanderson M, Schneider, Paula, AZEVEDO, VASCO. Comprehensive whole genome and transcriptome analysis for novel diagnostics In: Next-Generation Sequencing & Molecular Diagnostics. 1 ed.London: Future Medicine, p. 64-76, 2013

Home page: http://www.futuremedicine.com/doi/book/10.2217/9781780841861

2. **ABURJAILE, F. F.**, SANTANA, M. P., VIANA, M. V. C., Silva, Wanderson M, FOLADOR, E., SILVA, A., AZEVEDO, V.. Genomics In: A Textbook of Biotechnology.1 ed.Texas : SMGROUP, 2015, p. 20-40. ISBN: 9780996274531

## Book chapters accepted for publication

1. Santana, M., Aburjaile, F.F., Parise, M., Tiwari, S., Pinto, A and Azevedo V. RNAseq: Reveling biological insights in Bacteria, InTech - Next Generation Sequencing, 2015.

### **Articles in Newspapers/Magazines**

1. Soares, S. C., Silva, A, RAMOS, R. T. J, CERDEIRA, L., ALI, A., SANTOS, A. R., PINTO, A. C., Cassiano, AAM, FERNANDES, J. G. A., ABURJAILE, F. F., CARNEIRO, A. R., GUIMARAES, L. C., Barbosa, E. G. V., ALMEIDA, S. S., ABREU, V. A. C., MIYOSHI, A., AZEVEDO, V.. Plasticidade Genômica e Evolução Bacteriana. Microbiologia in Foco, 26 CBM. Foz do Iguaçu, p.31 - 38, 2011.

### Innovation

## **Research Project**

2009 - 2009 Projeto ManAli (Food handlers) Concluded

Integrantes: Flavia Figueira Aburjaile, Fernanda Diniz Prates, Juliana Fernandes, Kenia Cassia Pinto Gazola, Adriana dos Santos

## **Event Production**

## 1. ABURJAILE, F. F.

Member of the Organizing Committee of the II Congress of Microbiology and Parasitology "Oswino Penna Sobrinho", 2009.

### 2. ABURJAILE, F. F.

Member of the Organizing Committee of Academic Week the Health Sciences College in Universidade Fumec, 2008.

### Complementary education in France :

### **Poster presentation:**

• November 04 to 06 (2013)  $\rightarrow$  2<sup>nd</sup> Colloque de Génomique Environnementale (Rennes)

### **Curses Training:**

• October 24 to 26 (2012) → Blast2Go (Automatic Functional Annotation and Data Mining) –Valence – Spain

• April 18 and 19 (2013)  $\rightarrow$  Training: MISTER (Rennes)

• 21 and May 22 (2013) → Training: MIGALE - GALAXY (Jouy-en-Josas)

### French and English curses:

• October  $1^{st}$  to December 21 (2012)  $\rightarrow$  French course (B1) at CIREFE univ. Rennes 2 (4 hours per week)

• February 4 to May 7 (2013)  $\rightarrow$  French classes (B2) at CIREFE univ. Rennes2 (4 hours per week)

• April 4 (2013)  $\rightarrow$  English training

• May 13 to 17 (2013)  $\rightarrow$  Training: Scientific Communication in English and French

• September 30 to December 20 (2013) → French course (C1) at CIREFE univ. Rennes2 (4 hours per week)

## **Subjects and Seminar:**

January 22 (2013) → Mastery of scientific information (module 1) - École VAS
Beaulieu

• 24 and 25 January (2013)  $\rightarrow$  Mastery of scientific information (module 2) - École VAS - Beaulieu

• January 29 (2013)  $\rightarrow$  CM: scientific publication (module 3) - École VAS - Beaulieu

• September 19 (2013)  $\rightarrow$  NGS Seminar, Ilumina (Rennes)

## Total of production

## **Bibliographic Production**

Articles published in scientific journal – 22 Articles accepted in journals - 1 Chapters published - 3 Articles published in annals of events – 2 Presentations in Events (Congress) - 17

## Academic Advisories

Event organization - 2 Participation in events (congress, symposium, other) - 60