

Visual learning in Apis mellifera under virtual reality conditions

Gregory Lafon

▶ To cite this version:

Gregory Lafon. Visual learning in Apis mellifera under virtual reality conditions. Neuroscience. Université Paul Sabatier - Toulouse III, 2022. English. NNT: 2022TOU30016. tel-03714699

HAL Id: tel-03714699 https://theses.hal.science/tel-03714699

Submitted on 5 Jul2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.





En vue de l'obtention du DOCTORAT DE L'UNIVERSITÉ DE TOULOUSE

Délivré par l'Université Toulouse 3 - Paul Sabatier

Présentée et soutenue par Grégory LAFON

Le 25 mars 2022

Apprentissage visuel en réalité virtuelle chez Apis mellifera

Ecole doctorale : BSB - Biologie, Santé, Biotechnologies

Spécialité : **NEUROSCIENCES**

Unité de recherche : CRCA - Centre de Recherches sur la Cognition Animale

> Thèse dirigée par Martin GIURFA et Aurore AVARGUES-WEBER

> > Jury

M. Ludovic Dickel, Rapporteur Mme Elisa Frasnelli, Rapporteure M. Stéphane Viollet, Rapporteur M. Martin GIURFA, Directeur de thèse Mme Aurore Avarguès-Weber, Co-directrice de thèse

1 Table of Contents

2	Acknowledgements	3
3	List of abbreviations	5
4	Aims and goals	6
5	General Introduction	8
6	Prior studies on honey bee visual learning	10
7	Experiments with free-flying bees	11
8	Experiments with free-walking bees	18
9	Visual learning under full immobilization: conditioning of appetitive and aversive reflexes.	23
10	Virtual Reality: an innovative approach to study visual learning in bees	29
11	Studying navigation and attentional processes	30
12	Studying associative learning and memory	35
13	Benefits and caveats of using virtual reality to study visual learning in bees	40
14	What do we know of the underlying mechanism of visual learning?	43
15	The Optic lobes	44
16	The ventrolateral neuropils	46
17	The central complex	48
18	The mushroom bodies	.50
19	Whole circuit mechanisms	54
20	References	59
21 22	Chapter 1: Motion cues from the background influence associative color learning of honey bin a virtual-reality scenario	ees .68
23	Preface	.69
24	Summary	.71
25	Introduction	72
26	Materials and methods	74
27	Results	84
28	Discussion	99
29	References	108
30 31	Chapter 2: Visual learning in a virtual reality environment upregulates immediate early g expression in the mushroom bodies of honey bees	ene 112
32	Preface	113
33	Summary	115

34	Introduction116
35	Results
36	Discussion
37	Methods136
38	References144
39 40	Chapter 3: The neural signature of visual learning under restrictive virtual-reality conditions
41	Preface
42	Summary152
43	Introduction153
44	Results155
45	Discussion165
46	Materials and Methods172
47	References
48 49	Chapter 4: Comparison of associative visual learning in a 3D virtual reality between bumblebees and honeybees
50	Preface
51	Introduction190
52	Materials and Methods182
53	Results
54	Discussion220
55	Conclusion223
56	References
57	General Discussion
58	Virtual reality: progress, limitations and future developments
59	Investigating the brain regions involved in differential visual learning in VR234
60	Bumblebees suitability for VR experiments
61	Conclusion243
62	References
63	

64 Acknowledgements

- This PhD thesis was made possible by the continuous support of many people.
- 66 I would like first to thank Aurore Avarguès-Weber and Martin Giurfa for supervising my PhD
- and for their thorough revision of the various articles produced in this project.
- I also thank sincerely Dr. Ludovic Dickel, Dr. Elisa Frasnelli and Dr. Stephane Viollet foraccepting to be part of my PhD jury and evaluate my work.
- 70 Thanks are also due to Dr. Jean-Christophe Sandoz and Dr. Ricarda Scheiner for accepting to
- be part of my thesis committee and for their helpful advice and guidance during my PhD.
- 72 I'm extremely grateful to Haiyang Geng and Isabelle Massou who are responsible for all the
- cellular and molecular analysis that made chapter 2 and 3, and their associated publications,
- possible. They have done a staggering amount of work for this project and I consider myself
- 75 lucky to have had the opportunity to work with them.
- I am thankful to Alexis Buatois who established the initial VR setup for helping me get started
 with my PhD and for encouraging me to improve the VR.
- I also wish to thank Dr. Mathieu Lihoreau for providing colonies, and material to help us getstarted on the bumblebee experiments.
- I'd like to thank Stephane Kraus and Mathilde Lacombrade for teaching me how to take careof the bumblebees.
- I am grateful to Benjamin Paffhausen and Marco Paoli for many fruitful discussions, not all of
 them about our research. Their kindness and enthusiasm for science made working on this
 project truly enjoyable.

I would also like to thank Maud Combe and Patrick Arrufat for providing the technical support 85 that I needed to develop both the hardware and the software used in this thesis. 86 I am also thankful to Dr. Gabriella de Brito Sanchez for sharing her insights on bee gustation. 87 Thanks also to all the intern that I had the chance to supervise: Rodrigue Fouillet, Juliette 88 Montet, Diane Sam Mine, Emma Giordanengo, Catherine Macri, Clemence Guinnement, Marin 89 Nicolas, Naïs Judan, Eva Blot, and Karolina Pecharova. They have done an extensive amount 90 91 of work for this project and I hope that their time with us was as beneficial to them as it was for 92 me. 93 I am extremely gratefull to Dr Christian Jost for accepting to be my tutor in the DCCE program and helping me throughout my three years as a teaching assistant. 94

I am also gratefull to Dr. Jean-Marc Devaud, Dr. Cédrick Florian, Dr. Guillaume Isabel and Dr
Christian Jost for giving me the opportunity to teach in their courses.

97 Finally, I am extremely grateful to my fellow PhD students for answering all my numerous
98 questions about everything, from university paperwork to plant phylogeny, for helping me pick
99 the color of my graphs and of course for many memorable nights out.

100

- List of abbreviations: 101 102 AL: Antennal Lobe 103 104 AOTu: Anterior Optic Tubercle APIS: Automatic Performance Index System 105 **APL: Anterior Paired Lateral** 106 **CB:** Central Brain 107 **CS:** Conditioned Stimulus 108 109 **CX:** Central Complex DA: Dopamine 110 111 FB: Fan-shaped Body GABA: Gamma-Aminobutyric Acid 112 GLMM: generalized Linear Model 113 114 IEG: Immediate Early Gene LMC: Laminar Monopolar Cell 115 LMM: Linear Mixed Modal 116 LTM: Long Term Memory 117 MB: Mushroom Body 118 NC: No Choice 119 120 **OA:** Octopamine OL: Optic Lobe 121 PB: Protocerebral Bridge 122 PER: Proboscis Extension Response 123 124 RNA: Ribonucleic Acid 125 RT-qPCR: Reverse Transcription Quantitative Polymerase Chain Reaction SER: Sting Extension Response 126 sKC: Small Kenyon Cell 127 **US: Unconditioned Stimulus** 128 UV: Ultra Violet 129 VL: Vertical Lobe 130 131 **VR: Virtual Reality**
- 132

133 Aims and goals

134 The general objective of this doctoral thesis is to explore the underlying mechanism of visual learning in the honeybee Apis mellifera. To do so we decided to use virtual reality (VR), in 135 order to move from the classical studies using free-ly flying bees to a controlled setup in which 136 a tethered animal would learn visual discriminations. Our team had recently developed a new 137 VR setup, which was far from being immersive as it allowed only translational stimuli 138 139 movements (i.e. 2D VR). In order to be able to use VR to its full potential we first worked on upgrading the existing setup to a true 3D virtual environment. This introduced the possibility 140 of enriching the VR with a background that could generated optic flow as the bee moves within 141 142 the virtual world.

This new possibility inspired the first question addressed in this work, namely *how do motion cues from the background influence associative color learning in bees in the 3D VR environment?* To answer this question, we used our new setup to test if and how frontal motion cues generated in the VR and ventral motion cues generated by the movement of the treadmill below the bee affected color discrimination learning. In the first chapter, we present the answer to this question and identify issues that may affect decision-making in VR landscapes.

149 Answering that first question led us to refine both our setup and our conditioning protocols, thus raising our second question: What are the brain regions involved in visual learning? To 150 answer it, we quantified expression of three Immediate Early Genes (IEGs) that serve as 151 markers of neural activity in brain areas and that had been related to bee foraging and 152 orientation: kakusei, Hr38 and Erg1. We analyzed the expression of these IEGs in the calyces 153 of the mushroom bodies, the optic lobes and the rest of the brain after color discrimination 154 learning in VR. More specifically, we asked if the nature of IEG expression, and thus the areas 155 involved in visual learning, change depending on the way in which the animal learns the visual 156

discrimination. We thus compared IEG expression after learning in the 3D environment and
after learning in the 2D environment which set more constraints in terms of stimulus
movements. These two analyses are presented in two separate chapters

The study of the neural mechanisms underlying visual learning requires using invasive approaches to access the brain of the insects, which induces stress and can thus impair behaviors. To potentially mitigate this effect, we performed an additional study using bumblebees *Bombus terrestris*, which could constitute a good alternative to *Apis mellifera* as they are bigger and more robust and resistant to potentially harming procedures. In the last chapter, we explored the performance of bumblebees in a differential learning task in the VR and compared it to that of honeybees.

Overall, our work resulted in a novel and robust 3D VR system that is inexpensive, open source and supports experiments on both bumblebees and honeybees. This system represents therefore a qualitative advance for studies on honey bee visual learning. We also produced the first quantification of IEG expression in the bee brain as a result of associative visual learning and provide data showing the implication of mushroom bodies in this learning form. Taken together our results open the way for a deeper exploration of the mechansims of visual learning through VR experimentation.

174

175 Introduction

176 Honeybees are flying hymenoptera famous for their social organization and their important contribution to the pollination of crops and wild plants. Honeybees are central place foragers 177 that are flower constant (Grant, 1951; Chittka et al., 1999), meaning that they tend to constantly 178 179 return to the same species of flower as long as they are available and profitable, even if other more rewarding flower species are available in the vicinity. In relation with both their 180 eusociality and their status as central place foragers, honeybees have developed a complex 181 system of communication relying on one hand on pheromones and on the other hand on 182 stereotyped movements called "dances", which vary in shape according to the range of 183 184 distances separating the hive and the food source (Frisch et al., 1967). The most studied dance 185 type is the waggle dance, which reports distance and direction of profitable food source to nest mates. It's the discovery of this surprising behavior that made ethologist Karl von Frisch famous 186 (Frisch et al., 1967) and might have played an important role in consolidating the honeybee, 187 among a few other invertebrates, has a major research model in neuroscience and behavior. 188

189 Given honeybees' flower constancy, they have the capacity to learn and memorize the essential 190 traits that characterize the flower species they exploit at a time. Honeybees ability to identify and remember particular flowers species through numerous foraging bouts for periods of time 191 192 that can span several days make them a very enticing species for studying learning and memory (Menzel, 1999). Even more so because their brain is made of about 950 000 neurons for a 193 volume of about 1 mm³, which makes the underlying mechanism of their cognitive abilities 194 more accessible. Hence, for more than fifty years now, honeybees' associative learning abilities 195 196 have been extensively studied (Giurfa, 2007). Studies on honeybee learning have spanned 197 mostly the visual and the olfactory domain but have reached a unique dimension in the latter given the fact that honeybees can be easily conditioned to respond appetitively to a particular 198 odorant while being immobilized (Takeda, 1961; Bitterman et al., 1983; Giurfa and Sandoz, 199

2012). For the conditioning of the Proboscis Extension Response (PER) (Felsenberg et al., 200 201 2011) each bee is restrained in an individual harness such that it can only freely move its antennae and mouth-parts (mandibles and proboscis). When the antennae of a bee are touched 202 203 with sucrose solution, the animal exhibits the PER, i.e. it reflexively extends its proboscis to reach out to the sucrose. Neutral odorants blown to the antennae do not release such a reflex in 204 naive animals. If, however, an odorant is presented immediately before sucrose solution 205 206 (forward pairing), an association is formed which enables the odorant to elicit the PER in a 207 following test. This effect is clearly associative and constitutes a case of classical conditioning (Bitterman et al., 1983), i.e. the odorant can be viewed as the conditioned stimulus (CS) and the 208 209 sucrose solution as the rewarding, unconditioned stimulus (US) (Fig.5). Within this framework, bees learn to associate the odorant with the sucrose reward. Immobilization is crucial in this 210 context as it enabled the use of multiple invasive techniques to study the cellular and molecular 211 212 underpinnings of olfactory learning and memory (Mauelshagen, 1993; Abel et al., 2001; Komischke et al., 2005; Boitard et al., 2015; Carcaud et al., 2016). 213

214 Studies on olfactory learning have been mostly confined to the use of elemental protocols, that rely on the simple unambiguous association of at least two elements like in PER conditioning, 215 216 although more recently, studies on non-elemental olfactory learning, protocol in which the 217 reward or its absence is not associated univocally with the stimulus, have also revealed a capacity to solve non-linear discriminations, i.e. to go beyond simple forms of associative 218 learning (Meyer and Galizia, 2012; Devaud et al., 2015a). Yet, at the same time as it offered 219 the advantage of neural and molecular access, immobility represented a significant burden for 220 221 the possibility of observing the richness and cognitive complexity of free behavior. Experiments 222 with freely flying bees trained to solve discriminations in the visual modality showed precisely that the cognitive capacities of bees under these experimental conditions were highly elaborated 223 224 and parallel to some abilities that were thought a prerogative of vertebrates (Avarguès-Weber

et al., 2011a; Avarguès-Weber and Giurfa, 2013). For instance, free-flying honeybees were 225 226 shown to be able to learn concepts, a relation between different objects that is independent of the physical nature of the objects linked by the relation (Lamberts et al., 1998; Zentall et al., 227 228 2002; Doumas et al., 2008; Zentall et al., 2008; Halford et al., 2010), in the visual modality such has sameness (Giurfa et al., 2001) or above/below (Avarguès-Weber et al., 2011b) and 229 even combinations of those concepts (Avarguès-Weber et al., 2012a). However, the exploration 230 231 of the underlying mechanisms of visual learning, be it simple associations or conceptual learning, has remained elusive due to the impossibility of accessing the nervous system of flying 232 bees solving visual problems. At the same time, conditioning harnessed bees to visual stimuli 233 234 has yielded low rates of success (Avarguès-Weber and Mota, 2016). A goal of this thesis consisted, therefore, in overcoming this historic limitation by establishing a new experimental 235 paradigm allowing the coupled study of visual learning and neural analyses in bees. 236

237 Prior studies on honeybee visual learning

Honeybee visual learning has been studied for more than a century now, and many different 238 techniques and setups have been developed to this end. In order to understand what needs to be 239 done to allow us to progress in our understanding of their visual learning abilities we first need 240 to know what has already been done, what worked, and what didn't. In this part, I will show 241 that looking at the hundred years of scientific development (starting with von Frisch's seminal 242 demonstration of color vision in free-flying bees) clearly outlines the great success of 243 244 experiments on freely moving bees, which contrasts with the milder results obtained so far with immobilized bees. 245

246 Experiments with free-flying bees

In his pioneering work on honeybee vision, Karl von Frisch aimed at demonstrating that honeybees were endowed with color vision, although at that time the status quo was to claim that they were color blind (Hess, 1911). To this end, he trained bees to freely fly to an

experimental set up where they could get sucrose solution in a small Petri dish placed on a blue 250 piece of cardboard; then he tested the bees returning to the experimental site by offering at the 251 same location several targets, one blue, the same used during training, and various shades of 252 grey (Von Frisch, 1914). On all these visual stimuli, an empty Petri dish was presented. The 253 rationale of this experiment was to show that bees would not confound the learned color hue 254 with an achromatic intensity (a shade of grey) that would be common to the blue and grey 255 256 cardboards (Fig.1). He showed indeed that the majority of the bees chose to go to the blue target 257 and not to the achromatic alternative displaying a similar intensity (Von Frisch, 1914). With this experiment, which he repeated for various color hues, he demonstrated unequivocally that 258 259 honeybees are able to perceive colors.



260

Figure 1. Color vision of the honeybees. Experimental results from von Frisch experiment.
 Bees were trained to feed on blue feeders and were tested on a multitude of feeders in random
 arrangement of blue and grey stimuli. The blue feeder received more visits than the achromatic
 ones. Taken from von Frisch 1914.

Since then, many experiments have studied different form of visual learning in free-flight conditions. In this type of experiment, trained honeybee foragers come directly from the hive to an experimental site where the stimuli to be learned and/or discriminated are offered, paired or not with a sucrose reward. The trained bee, marked with a color on its abdomen or thorax in order to identify it, will perform many flights between the hive and the experimental site to collect the food reward and thereby to answer the questions raised by the experimenter. This scenario can be labeled as mainly operant conditioning (Rapaport, 1973) as obtaining or not the reward depends on the bees' actions because it is the decision of the bee to land or not on a given target that defines if the reward is obtained or not. Yet, it also includes Pavlovian associations between the visual stimulus and the reward (or absence of it) following the classic scheme of a CS-US association, and eventually associations between the visual stimulus and the response to be produced.

277 In 1967, Randolf Menzel initiated the study of color memory using free flying experiments (Menzel, 1967). In these experiments, bees were trained to collect sucrose solution on a 278 279 horizontal table in which spectral filters illuminated from below provided the color cues to be 280 learned. Bees were trained with one or more learning trials and varying alternative color (nonrewarded) presented adjacent to the rewarded color. The goal was to quantify the color memory 281 resulting from this training for each wavelength trained. Memory retention was tested in 282 extinction conditions (no reward provided) and presenting the rewarded color against a different 283 color to assess the specificity of the color memory acquired. He demonstrated that bees are able 284 285 to specifically learn the rewarded color and that different spectral colors are learned at different speeds. With violet (413, 428 nm) being the fastest and the most reliably learned with up to 286 85% correct responses during the test, and blue-green (494 nm) being the slowest. 287

Presenting stimuli horizontally constituted a problem for the study of shape and pattern learning as bees could have only a partial view of a pattern perceived upon their approach. It was therefore decided that presenting the stimuli vertically and frontally would preclude this problem as bees would be forced to see the trained stimuli entirely (Wehner, 1967). Since then the study of visual learning in freely flying bees switched to a vertical form of stimulus presentation in the majority of the works that aimed at controlling visual perception properly. Yet, another problem was realized later: what the bee would perceived depended on its distance to the target, i.e. on the visual angle subtended by the stimulus, which determined or not thatthe stimulus was resolvable for the bees' visual system (Srinivasan and Lehrer, 1988).

A way to solve this problem was found by adopting Y-mazes, in which not only stimuli were 297 298 presented vertically on the back walls, but also which allowed controlling the distance from the stimuli to a decision chamber leading to both maze arms. In this way, the visual angle of the 299 visual stimuli could be controlled (Srinivasan and Lehrer, 1988; Hateren et al., 1990; M. Giurfa 300 et al., 1996). In these experiments, forager bees are first trained to collect a sucrose reward at 301 the end of each of the two arms of an experimental Y-maze. Then, during each conditioning 302 trial, individual bees have to choose between a rewarded and a non-rewarded arm, 303 304 distinguishable by adequate cues situated at the end of each arm (Fig.2).



305

Figure 2. Y-maze for honeybees. Prior to this conditioning honeybees learn to come to the entrance of the maze by flying from their hive on their own accord to collect a sucrose reward from each arm of the maze. During each conditioning trial only one arm is rewarded in association with a particular stimulus the bee can see from the decision chamber and has to associate with the reward. Adapted from Avarguès-Weber et al., 2011a. Using these various protocols allowed determining that bees can not only discriminate colors but also a large variety of visual attributes such as shapes and patterns, depth, motion, light intensity, contrast and complex configurations (Menzel, 1967; Zhang et al., 1996; Giurfa and Menzel, 1997; Horridge, 2000; Srinivasan, 2010; Avarguès-Weber et al., 2011a). Throughout the years free flight has been used extensively to analyze visual memory, discrimination and generalization (Zhang et al., 1992, 1999; Giurfa, 2004; Zhang et al., 2005; Dyer et al., 2011; Dyer, 2012; Zhang, 2012).

In the last two decades, however, a 'cognitive revolution' took place and visual learning moved to a different level, namely the study of higher order learning capacities in bees, capacities that until then were considered to be absent in the miniature brains of insects despite the fact that studies had already documented the ability of honeybees to generalize among visual stimuli (Horridge, 2009).

It started with the demonstration of symmetry categorization in bees (Martin Giurfa et al., 323 324 1996). Categorization consists in grouping together stimuli that are recognized as explicitly 325 different but which are classified as similar based on shared attributes. Any unknown exotic bird will be recognized as a "bird" based on the presence of attributes defining this category 326 such as wings, feathers, a beak, etc. In Giurfa et al. study bees were trained to collect a reward 327 328 from vertically presented stimuli, as described earlier (Wehner, 1967), that remained constant only in their degree of symmetry. One group was trained to collect reward on symmetrical 329 stimuli and the other on asymmetrical ones. During the test both groups were able to correctly 330 331 chose the novel symmetrical or asymmetrical stimulus respectively. Bees were thus able perceive the bilateral symmetry and generalize it to novel stimuli. 332

Later, free flying bees were studied for their capacity to learn conceptual relationships, meaning concepts that rely on relationships between stimuli rather than on physical features of the stimuli (Zentall et al., 2002; Avarguès-Weber and Giurfa, 2013). One particular protocol possible to

test this capacity is the 'delayed matching-to-sample' task, in which an animal is presented with 336 a 'sample' and subsequently with two or more secondary stimuli, one of which is identical to 337 the sample. The animal is required to respond to the stimulus just encountered, i.e. to use the 338 339 relational rule 'choose the same as the same previously seen', irrespective of the nature of this sample. The 'delayed non-matching-to-sample' is similar to the matching-to-sample task except 340 that the animal is required to respond to the stimulus that is always different from the sample. 341 In both cases, broadly construed sameness and difference concepts are shown only if the animal 342 exhibits positive transfer to a completely new set of stimuli, which it had not experienced during 343 training (Giurfa et al., 2001). This capacity was shown by Giurfa and collaborators, who trained 344 honeybees, A. mellifera, in a delayed matching-to-sample paradigm to examine whether they 345 could form a concept of sameness and a concept of difference (Giurfa et al., 2001). In the 346 sameness version, each bee entered the maze by flying through a hole in the middle of an 347 348 entrance wall. At the entrance, the bee encountered the sample stimulus. The sample was one of two different stimuli, A or B, alternated in a pseudo-random sequence. The entrance led to a 349 decision chamber, where the bee could choose one of two arms. Each arm carried either 350 stimulus A or stimulus B as secondary stimulus. The bee was rewarded with sucrose solution 351 only if it chose the stimulus that was identical to the sample. If the bees managed to learn the 352 353 original discrimination, they were tested with a new sample and secondary stimuli in 'transfer tests' in extinction conditions (no reward provided): the bees had to choose between stimuli C 354 and D, when the sample was either C or D. In such tests, bees that had been trained to match 355 colors could match achromatic gratings and bees that had been trained to match achromatic 356 gratings could match the colors with a success rate of about 70%, demonstrating thereby the 357 capacity to learn the concept of sameness. Similar experiments demonstrated also the capacity 358 to learn the concept of difference. Further work demonstrated that bees can also handle concepts 359

361 difference and spatial relationships at the same time (Avarguès-Weber et al., 2012a).



362

Figure 3. Rule learning in honeybees. (a) Honeybees trained in a delayed matching-to-sample task to collect sugar solution in a Y-maze where they first get a sample at the entrance and then a choice within the maze and need to choose the stimulus matching the initial sample. (b) Theyare trained on a series of patterns or colors to learn a rule of sameness. Bees trained on the patterns were tested on the colors and vice-versa. In both cases, bees chose the novel stimuli corresponding to the sample. Taken from Avarguès-Weber et al., 2011a.

369 More recently the Y-maze was used to investigate numerical cognition in free-flying honeybees. It was shown that bees are endowed with numeric competence and can count visual 370 items until 4 or 5, different results were obtained in different experiments (Chittka and Geiger, 371 1995; Dacke and Srinivasan, 2008; Gross et al., 2009; Skorupski et al., 2018). Bees could also 372 manage some basic level of addition and subtractions (Howard et al., 2019). In this study 373 honeybees were trained to enter a Y-maze and view a visual sample stimulus presented 374 vertically containing a set of elements of a given color. The color defined the arithmetic 375 376 operation to perform once in the maze. For instance, if three blue items were shown at the entrance of the maze, the blue color indicated addition of one; therefore, the bee entering the 377 378 maze should choose a stimulus displaying four items and not two or five. If the items at the 379 entrance were yellow, the arithmetic operation to perform was subtraction of one. Thus, if three yellow items were shown at the entrance, choice of two within the maze represented the correct option. The color of the elements, and thus the arithmetic problem to be solved, was randomly assigned per bee for each trial. Correct and incorrect options during experiments ranged from one to five elements, and the incorrect option could be higher or lower than the correct option (which also included the sample number as a possible incorrect option). The sample number of three elements was never shown during training and was only used as a novel sample number during testing.

Flying bees are able to perform very difficult tasks when flying through unknown environments 387 relying mainly on the optic flow cues generated by their own motion (Horridge, 1987). Optic 388 389 flow is the speed of movement of an image on the retina, it can be used as a proxy of distance as object that are close appear to be moving faster than object that are further away. In 1989 390 Kirchner and Srinivasan trained bees to receive a food reward at the end of a tunnel where each 391 wall displayed a pattern of vertical black bars and white gratings, to create a texture that would 392 produce optic flow on the retina of the bees, one of the gratings could be moved either in the 393 394 direction of the flight or against it, to reduce or increase the optic flow respectively (Kirchner and Srinivasan, 1989). They showed that bees flied in the middle of the tunnel when the optic 395 flow was equivalent on both sides but when the grating was moved in the same direction as the 396 397 bees' flight, thus reducing optic flow, bees flied closer to the moving grating. Respectively when optic flow was increased bees flied further away from the moving grating. They thus 398 concluded that honeybees use optic flow as a measure of object distance and use this measure 399 to avoid collisions during flights. Since then it was shown that optic flow is also used to control 400 401 speed, height, and to avoid lateral obstacle (Srinivasan et al., 1991; Baird et al., 2006; Baird and 402 Dacke, 2012; Baird et al., 2021). The impressive abilities of bees to fly through complex environment using optic flow to guide them has been reproduce in biomimetic robots that was 403 404 shown to reproduce bees ability to avoid collision in narrow corridors and adjust their speed in wind condition (Roubieu et al., 2014), thereby showing that managing optic flow is sufficientto reproduce the honeybee flying abilities in flight corridors (Fig.3).



407

Figure 3. Automatic speed control and lateral positioning of a miniature hovercraft navigating in a 4 m long straight or tapered corridor (a). The hovercraft is equipped with a bee-inspired autopilot based on the dual optical flow regulators and is endowed with an insectinspired 4-pixel visual system. (b) Chronophotography (~1s time interval) of the hovercraft crossing the corridor. Adapted from Roubieu et al., 2014.

- Free flights experiments have proven to be a very powerful tool in the investigation of honeybees' visual cognition and allowed to uncover the bee incredible learning abilities and cement it as a major model organism in neuroscience. However, because free flight experiments take place in the field they lack the finer control that lab experiments can allow and more importantly, they preclude the use of invasive methods to study the mechanisms of these intriguing performances.
- 419 Experiments with free-walking bees
- 420 The visual learning of honeybees can also be studied in more controlled paradigms in which
- 421 bees walk, generally within reduced setups which force the animals to walk instead of flying.
- 422 For instance, Zhang et al (Zhang et al., 1998) trained bees to walk through a narrow tunnel

423 carrying visual stimuli on the two walls to study their capacity to learn routes based on visual 424 stimuli presented to a single eye, and to then navigate these routes using the other (naive) eye. 425 Bees reaching the end of the tunnel had to turn right or left, one of these choices being correct 426 and leading to reward while the other not. Using the narrow tunnel for a walking bee ensured 427 that what was presented in the lateral walls of the tunnel was the only visible cue to a given eye. 428 They found that stimuli encountered by different eyes could be associated with different routes 429 and that bees could learn to associate a color with a turning direction based on monocular cues.

A similar approach was used by Menzel (Menzel, 2009) who trained bees to turn either left or 430 right in a narrow T-maze depending on the sequence of colors (blue, yellow) experienced at 431 432 four positions in the access arm. In this way he aimed at studying the learning of sequential visual configurations as predictive of reward. The results showed that visual cues differed in 433 their capacity to predict reward when presented alone in a test at one of the four positions of 434 the access tunnel. The position closest to the maze branching had the highest predictive value 435 while that at the entrance of the maze had the lowest value. Thus, the four positions were 436 437 equipped with different salience scores, which reflected probably their contiguity to reward, and which added up independently, although in some tests configuring of sequential patterns 438 was also observed. 439

Walking setups have been also used to study aversive learning as they offered the possibility of 440 delivering punishment (e.g. electric shock) via the tarsi of the walking bees. For instance, 441 Nicholas H. Kirkerud and collaborators established an automated setup to study a passive-442 avoidance task that they called APIS, the Automatic Performance Index System (Kirkerud et 443 al., 2013). It's an enclosed walking channel where the interior is covered with an electric grid, 444 445 and where presentation of odors from either end can be combined with weak electric shocks to form aversive associations. To quantify behavioral responses, the movement of the bee is 446 monitored by an automatic tracking system. Number of escapes from one side to the other, 447

changes in velocity as well as distance and time spent away from the punished odor are 448 449 measured to describe the bee's learning capabilities (Kirkerud et al., 2013). This setup was then adapted for color learning, where one half of the assay is illuminated with one color paired with 450 electric shock and the other half was illuminated with light of a different wavelength and not 451 paired with shocks. The unrestrained bee could run away from the light stimulus and thereby 452 associate one wavelength with punishment, and the other with safety (Fig. 4a) (Kirkerud et al., 453 454 2017). A similar setup (Fig. 4b) was used by Agarwal and collaborators to explore the influence of dopamine (DA) and octopamine (OA) on avoidance learning (Agarwal et al., 2011). In this 455 study free walking honeybees had to learn the association between a mild electric shock and a 456 457 special color cue. After five trials control bees successfully learned the association and stopped going to the color side associated with the shock. OA impaired avoidance learning as OA treated 458 bees spent more time in the shock paired compartment and a lower proportion of insects reached 459 460 complete avoidance. DA on the other hand improved the learning has treated bees spent less time in the shock paired compartment that control bees. Thus the reward and punishment 461 pathways are inter-connected as OA is known to be involved in motivation, reward, and 462 modulation of motor functions in insects (Schwaerzel et al., 2003). Plath et al also used a setup 463 adapted from APIS to explore the role of the central complex and of the mushroom bodies in 464 aversive color learning (Plath et al., 2017). They found that silencing either the central complex 465 or the medial lobes of the mushroom bodies (Fig.15) impaired the ability of the bee to associate 466 the light field with the shock. On the other hand, inactivating one collar region of the mushroom 467 bodies calyx (Fig.15.C) did not affect learning in this assay (Plath et al., 2017). Electric shock 468 associated with color lights in a double-chamber setup was also used, yet not for aversive color 469 discrimination learning, as in the cases mentioned above, but for phototactic suppression based 470 on aversive learning. In this setup termed ICARUS bees learned to suppress their spontaneous 471 attraction toward a blue lit compartment in which they would receive a mild electric shock 472

(Fig.4c) (Marchal et al., 2019). The setup is made of two illuminated chambers, initially both 473 chambers are illuminated in red, not visible to the bee (640 nm), then the chamber not currently 474 occupied by the bee is lit in blue, through positive phototaxy the naive insect will spontaneously 475 enter the lit chamber and receive an electric shock. The delay between the beginning of the 476 illumination and the moment the bee crosses the threshold is recorded and serves as a measure 477 of learning. This protocol was sufficient to induce visual learning as bees were able to 478 successfully repress their spontaneous phototactic response toward the blue light and to create 479 long term memory as honeybees still repressed their attraction to blue light 24 h after the last 480 conditioning trial. Finally, they performed RT-qPCR in individual brains of successfully trained 481 animals focusing on expression levels of the three dopamine-receptor genes Amdop1, Amdop2, 482 and Amdop3. Coherent with Agarwal's results on DA (Agarwal et al., 2011), found an up-483 regulation of the dopaminergic receptor gene Amdop1 in the calvees of the mushroom bodies 484 485 as a result of the conditioning (Marchal et al., 2019).



486

Figure 4. Shuttle boxes for aversive visual learning in bees. In these setups, the bees are located in an elongated chamber where they shuttle back and forth. During a conditioning trial, each half of the chamber is identified by a visual cue (colored sheets of paper or color LEDs) and one of these cues is associated with electric shock delivered by a shock grid whenever the bees enters the compartment. The position of the bees is either assessed by a number of infrared barriers (a) or manually (b and c). (a) APIS setup taken from Kirkerud et al., 2017. (b) Taken from Agarwal et al., 2011. (c) Taken from Marchal et al., 2019.

Both APIS and ICARUS give more control over the timing of the experiments than under free

flight conditions because the whole experiment can take place in the lab and the bees do not

496 return to the hive. It is then possible to perform more invasive experiments like local

497 inactivation of brain structures, using neuropharmacological blockade of target receptors

- 498 (Agarwal et al., 2011), transiently inactivating specific brain structure with anesthetic (Plath et
- al., 2017), or quantifying relative levels of gene expression in key brain structures following
- 500 aversive learning (Marchal et al., 2019).

In order to study appetitive conditioning in more controlled conditions than free flight it is 501 possible to adapt the Y-maze used with free flying honeybees to a semi-restraining version in 502 which bees move freely within a miniature maze but do not return to the hive in between trials 503 504 (Buatois et al., 2018; Bestea et al., 2022). Such a maze had detachable end sections that could be closed and moved to the start of the maze after each choice. In this way, a bee was forced to 505 do consecutive choices by translocating it repeated times to the start of the maze. This setup 506 507 was used, among others, to control for the effect of manipulated appetitive motivation, which would change dramatically if the bee returns to the hive and unload the collected food (Bestea 508 et al., 2022). 509

510 Thanks to those protocols it has been possible to investigate further the underlying mechanisms of visual learning by coupling controlled behavioral experiments with neural analyses of 511 targeted regions of the brain. However, because the insect can move freely, the analyses 512 performed are relatively crude as they proposed 'static' views of neural activation (via gene 513 expression analyses) or because they targeted broad areas via pharmacological blockade. In all 514 515 cases, what is missing is the possibility to couple the study of behavioral performances with an online recording of neural activity, which is of fundamental importance to characterize the 516 neural signature of visual learning and memory. 517

518 Visual learning under full immobilization: conditioning of appetitive and aversive reflexes

519 One of the most widely used protocol to investigate learning and memory in bees is the 520 appetitive reflex elicited by chemosensory contact of sucrose solution with sucrose receptors 521 located on the antennae and the tarsi (Minnich, 1921; Frings, 1944; Frings and Frings, 1949). 522 This response termed PER (Proboscis Extension Reflex) has been used extensively to study 523 olfactory learning and memory (Giurfa, 2007). PER conditioning as also been adapted to other 524 hymenoptera like bumblebees (Laloi et al., 1999; Riveros and Gronenberg, 2009). As described earlier, individually harnessed bees learn the association between a neutral odorant and asucrose reward such that the odorant ends up eliciting PER (Fig.5).



Figure 5. Olfactory conditioning of the proboscis extension reflex. (A) Before conditioning,
stimulation with the neutral odor does not elicit the PER. (B) After forward pairing of odor
stimulation and sucrose solution the honeybee responds to the odor with PER. Taken from
(Scheiner et al., 2013).

532 The PER protocol was originally conceived to study visual learning. In its first version, published in 1957, Kuwabara introduced the protocol to study Pavlovian visual learning. Yet, 533 he realized that learning would be only possible if the antennae of the bee were sectioned. He 534 mentioned that this procedure was necessary to allow the acquisition of color-reward 535 associations and consequent color-dependent PER response because restrained bees with intact 536 antennae apparently developed unspecific PER responses to the water vapor from the small 537 spoon used to deliver sucrose solution as reward (Kuwabara, 1957). Fifteen years after 538 Kuwabara's original report, another study reported results using visual-PER conditioning of 539 540 honeybees with intact antennae. But it required up to 50-60 training trials to achieve learning (Masuhr and Menzel, 1972). Another study reported no significant acquisition of visual-541 induced PER in bees with intact antennae during a 6-trial pre-training phase in which bees had 542 543 to associate a white-light stimulus with sucrose reward (Gerber and Smith, 1998). Only later, the group of Takeo Kubo reproduced Kuwabara's results on visual PER conditioning but 544 sectioning the antennae again (Hori et al., 2006, 2007). The first study compared visual learning 545

performances of antennae-deprived and intact restrained bees, and found that only antennae-546 deprived bees were able to acquire significant visual-induced PER after 20 trials of classical 547 conditioning using green (540 nm) or red light (618 nm) as conditioned stimulus (Hori et al., 548 549 2006) or, in a different work, motion cues which could simulate backward or forward optic flow (Hori et al., 2007). Red light was used "to activate exclusively the L-receptor type", thus as a 550 case of achromatic visual conditioning. In the case of motion cues, bees were able to be 551 552 conditioned equally well to forward and backward movement, and were able to specifically respond to the conditioned motion as bees conditioned to backward motion responded 553 significantly more (44.4%) to backward than to forward motions (14.8%) (Hori et al., 2007). 554

555 More recent works found that intact restrained honeybees can acquire visually-induced PER responses both in absolute and differential visual conditioning paradigms (Dobrin and 556 Fahrbach, 2012; Sakura et al., 2012; Jernigan et al., 2014; Balamurali et al., 2015). Contrary to 557 previous studies (Hori et al., 2007, 2006; T. Mota et al., 2011), some of these authors found that 558 antennae amputation even impaired visual-PER acquisition (Jernigan et al., 2014). The 559 560 potential bias due to responses to water vapor perception when training the bees with intact antennae was not discussed in these papers. Moreover, Dobrin and Fahrbach showed reduced 561 discrimination performances when a wet toothpick was presented to the bees in the CS- trials 562 563 instead of a dry toothpick (Dobrin and Fahrbach, 2012). Thus, when the unambiguous presence of water could not be used as an additional predictive factor of reward, the bees' selective 564 responses to the conditioned color dropped significantly suggesting a crucial influence of this 565 factor. Thus, at the present time, the role and potential interference of the antennae on visual 566 PER conditioning, and the causes for this interference remain unclear. 567

568 With or without antennae, visual learning performances in PER conditioning have never 569 reached the levels usually observed in free-flying bees trained to visual stimuli, often 570 characterized by fast acquisition rates and high percentages of correct choices (70–100%) at the

end of training (M. Giurfa et al., 1996; Dyer and Neumeyer, 2005; Avarguès-Weber et al., 2010) 571 572 or the levels that are characteristic of olfactory PER conditioning, which allows reaching a plateau of 80-90% correct choices in the case of a salient odorant (Bitterman et al., 1983; Giurfa 573 574 and Sandoz, 2012). Also, the number of conditioning trials required to reach a plateau in the learning curve is dramatically different between visual and olfactory conditioning of PER. 575 576 While few trials (usually three to five) are required for successful olfactory conditioning of PER 577 (Bitterman et al., 1983; Guerrieri et al., 2005; Matsumoto et al., 2012), a higher number (6 to 20) of trials is required for visual conditioning of PER and acquisition levels are substantially 578 lower (40%) (Hori et al., 2006, 2007). 579

Therefore, visual PER in its current state does not meet the expectations set by olfactory PER
as it is not only inconvenient to use, with high number of trial and antennae removal, but also
has poor learning performances.

In order to study aversive visual learning in restrained bees, it is possible to use the sting 583 584 extension response (SER), which is defensive behavior of bees elicited by potentially noxious 585 stimuli (Breed et al., 2004). It is possible to elicit SER in laboratory through the application of an electric shock to the thorax (Núñez et al., 1997). This lead to the implementation of an 586 aversive olfactory conditioning protocol (Vergoz et al., 2007). Honeybees were fixed 587 588 individually on a metallic holder so that they build a bridge between two electrodes through which a mild electric shock can be delivered to elicit the SER. A 2s electric shock served as the 589 590 US and was paired with a 5s odor pulse as CS (Fig.6) (Vergoz et al., 2007). This protocol was later adapted for visual learning (Fig.6a) (Theo Mota et al., 2011a) as an alternative to visual 591 PER. Bees were able to discriminate two colors, green and blue, that varied both in their 592 593 chromatic and achromatic properties, reaching 40% of specific response to the reinforced color after 6 trials. They were also able to discriminate colors that varied only in either their chromatic 594 or achromatic properties with the same percentage of success at the last trial. Lastly the authors 595

showed that not only antennae ablation was not necessary for this conditioning but that it 596 decrease bees' performances (Theo Mota et al., 2011a). However, due spatial constraints (the 597 sting had to stay visible to the experimenter) the visual stimulus could only be projected on one 598 eye, even though we know that bees can learn visual exposure through monocular exposure 599 600 (Masuhr and Menzel, 1972) this is limitation for further exploration of the underlying mechanism of visual learning. Moreover, Mota and collaborators' study showed a constant 601 unspecific response of about 20% in addition to the low percentage of conditioned response 602 603 (40%) so visual SER is pretty far from the performances observed with olfactory PER.

Thus visual SER does not meet the expectations set by olfactory PER for reasons similar to theones mentioned for visual PER.

606



Figure 6. Visual conditioning of the sting extension reflex (SER). (a) Experimental setup, a
honeybee was individually harnessed in a holder allowing the delivery of a mild electric shock.
The visual stimulus was presented on a white screen to the right eye of the harnessed bee. Taken
from Theo Mota et al., 2011a. (b) Picture of a bee harnessed in a SER setup. Taken from
(Tedjakumala and Giurfa, 2013) (c) Picture of a sting extension. Taken from (Tedjakumala and
Giurfa, 2013)



622 Virtual Reality: an innovative approach to study visual learning in bees

Virtual reality (VR) is a scenario built on the basis of artificial sensory stimuli, often generated
by computer, that gives the feeling of immersion in an actual world as it allows moving and
interacting within that environment while being in fact stationary in the real world.

For walking insects the development of VR systems started more than 70 years ago with a 626 627 pioneering setup published by Bernhard Hassenstein to characterize for the first time optomotor responses in beetles (Hassenstein, 1950). This setup was not a true VR in the sense that it did 628 629 not create a visual environment for the beetle under study. Yet the visual panorama was coupled to the beetle's movements. The insect was tethered onto a very lightweight 'Y-maze globe' 630 made of thin straws, which turned below the beetle as the beetle 'walked' along a blade of 631 632 straw, thus repeatedly confronting the beetle with Y-maze choices of diverging straws. The 633 tethered beetle could then be exposed to highly controlled, moving visual stimuli, namely a cylinder with vertical black and white stripes, allowing the simultaneous recording of its 634 directional choices on the globe in response to the moving stripes (Hassenstein, 1951). 635

Later a flight simulator was built by Götz to study the optical properties of the compound eye 636 of Drosophila melanogaster. It consisted of a torque meter suspended in the middle of a 637 638 cylinder with textured walls. The fly was suspended by its thorax to the torque meter which was thus able to measure the rotations of the fly as it reacted to the movements of the cylinder walls. 639 640 It was an open loop setup where the fly could react to the stimuli presented to it but could not 641 control their movements (Fig.7) (Götz, 1964). Measuring the optomotor response of the flies to 642 movement of the walls, Götz was able to show that the perception of motion depends only on the temporal, not on the spatial phase relations between periodic intensity variations in 643 644 neighboring ommatidia.



Figure 7. Flight simulator build by Götz. (Left) Electric diagram and schematics of the torque
meter used to measure the optomor respone of the fly. (right) Textured wall used to induce
optomor response in the fly. Taken from Götz, 1964

649 Nowadays thanks to the advancement of computers, the range of virtual realities that could be 650 generated became virtually infinite, ranging from simple shapes and colors to impossible world 651 with non-Euclidian geometries. Thus new approaches and systems were produced to test a wide 652 variety of hypotheses on bee visual behavior.

653 Studying navigation and attentional processes

654 VR for honeybees made its first significant start with a flight simulator build by Luu et al (Luu et al., 2011). A tethered bee was suspended in the middle of four LCD monitors that displayed 655 a moving panorama. They could thus study the behavioral response of bees to being passively 656 exposed to a moving panorama, as a way to examine whether and how optic flow affects body 657 posture during flight. The authors were able to make the tethered bees fly in these experimental 658 659 conditions and noticed that, upon such suspended flight, bees slightly raise their abdomen, a response that is interpreted as a 'streamlining response' (Fig8), presumably to reduce 660 aerodynamic drag. This response was elicited by pure visual exposure (Luu et al., 2011) and 661 was strongest when the image motion was in the direction that would be experienced during 662 forward flight and when it covered the full visual panorama of the bee. It was highly sensitive 663

in the lateral rather than in the frontal and rear fields, and it was also modulated by air-flowcues simulating head-wind (Taylor et al., 2013).



Figure 8. Illustration of the measurement of the streamlining response. The streamlining
response is defined as the orientation of the abdomen relative to the thorax. The sketches
illustrate the definition of the response, and the images give three examples of how this is used
to measure the response. Taken from Luu et al., 2011.

666

More recently another flying VR was established to study the relative importance of motion 671 672 cues and occlusion cues in flight navigation in *Bombus ignitus*. Bumblebees could freely move within an enclosed Plexiglass arena ($62 \times 32 \times 42$ cm), with a LCD monitor at the floor of the 673 arena. The movement of the bee where tracked in to two camera placed orthogonally to each 674 675 other outside the arena. The surface displayed on the monitor could thus be updated according to the movement of the bumblebee thereby creating a virtual floor that could be positioned at 676 677 an arbitrary height compared to the real floor (Fig.9a) (Frasnelli et al., 2018). Bumblebees were first trained to feed from a static target, a blue disc, displayed on the floor of the arena before 678 being submitted to various configurations of virtual floor they had to go through to reach the 679 target. The VR displayed an elevated platform above the floor where the target was positioned, 680 the elevated platform had a hole through which the insect could fly to reach the target below. 681 In order to create the illusion that the elevated platform is above the floor, the authors controlled 682 683 two parameters: motion cues and occlusion cues. Motion cues are the apparent movement speed

of different objects: objects that appear to move fast are close, while slower objects appear to 684 be further away. A typical way to explain that effect is to describe the experience one has when 685 being a passenger on the highway, when looking out the side window, objects near the car 686 appear to move really fast, while objects further away move slower and really distant objects 687 on the horizon appear almost still. Thus, by making the elevated platform move slower than the 688 virtual floor in response to the bee movements, it would appear to be closer to the insect. 689 Occlusion cues are easier to describe; the closest object should hide the object situated behind 690 it. This setup was sufficient to produce a believable illusion for the bumblebees as they tried to 691 avoid the virtual elevated platform, and slowed down and extended their leg when getting close 692 in an attempt to land on it. The genius of this setup, and one of the real strength of VR in general, 693 is that it allowed to create impossible realities where motion cues and occlusion cues were 694 conflicting. It means that the floor would move slower than the platform (motion cues), but 695 696 would mask the platform (occlusion clues). The authors were thus able to evaluate what cues takes precedence for the bumblebee flight navigation by creating this conflicting situation 697 (Fig.9b). The bumblebees flew through the occluding texture and avoided the regions with 698 699 higher motion speed to reach the target (Fig.9b), thereby showing that they prioritize motion cues over occlusion clues for flight navigation (Frasnelli et al., 2018). 700

701



Figure 9. Bumblebee free flight VR arena. (a) Diagram of the flight arena. (b) Behaviour
 results from congruent (top) and incongruent (bottom) VR. Virtual platform is in purple and
 virtual floor in salmon. The green crosses represent the positions at which individuals initially
 descended through the plane of the virtual platform. Adapted from Frasnelli et al., 2018.

Many setups have also been made to study walking bees (Schultheiss et al., 2017). They are 707 called locomotion compensators, or running spheres, use either a light weight ball suspended 708 709 on an airflow or a ball controlled by precise servomotors that compensate the movement of the insect to always keep it at the top of the ball. This thus creates an omnidirectional treadmill on 710 711 which the bee can run indefinitely (Fig.10). Ball movements can be tracked accurately by 712 appropriate optical mouse sensors (Fig10a) (Taylor et al., 2015) or a video camera (Moore et al., 2014). This kind of device has been used for more than half a century to study different 713 aspects of insect behavior, in particular stereotyped responses to environmental stimuli 714 715 (Kramer, 1976). Recent progress in video tracking and computer controlled systems have allowed to present the insect with a visual environment that is directly updated by its 716 717 movements walking stationary on the treadmill (closed-loop). Paulk et al. (Paulk et al., 2014a) used a variant of such a closed-loop VR setup for studying attention-like processes in tethered 718 walking honeybees. Bees walking stationary in the middle of an LED arena were presented 719 with one or two competing green vertical bars separated by 90° and flickering at different 720 frequencies. The goal was to confront the tethered bee with two competing percepts, which 721

would induce sharing attentional resources between them given the fact that bees tend to fixate 722 723 either stimulus. The authors were able to combine the recording of behavioral fixation of these stimuli with extracellular electrophysiological recordings of neural activity in different parts of 724 the bee brain, inspired by earlier work on Drosophila (Van Swinderen, 2012). Using this 725 method, it was shown that attention-like processes had a neural correlate at the level of the optic 726 lobes before the bee displayed a behavioral choice. No such correlate was detected at the level 727 728 of the mushroom bodies probably because of the sparse coding occurring at this level, which 729 renders difficult detecting electrophysiological signals



730

Figure 10. VR setup using a locomotion compensator. A) Global view of the VR system. 1: 731 Semicircular projection screen made of tracing paper. 2: Holding frame to place the tethered 732 bee on the treadmill. 3: Treadmill made of a Styrofoam ball floating on an air cushion. 4: 733 Infrared mouse optic sensors allowing to record the displacement of the ball. 5: Air input. B) 734 The tethering system. 1: Plastic cylinder containing a glass cannula into which a steel needle is 735 736 inserted. 2: Needle attached to the thorax of the bee. 3: Its curved end is fixed to the thorax by means of melted bee wax. C) Example of stimuli presented to the insect, her two colored 737 cuboids. 738

739 The use of closed-loop instead of open-loop controlled visual stimuli seems to be an important

parameter, as it increases the temporal coordination of neural activity in the insect brain (Paulk

- et al., 2014b, 2015). Closed loop conditions also seem to modulate neural activity as early as
- the medulla (Rusch et al., 2021). Indeed, when honeybees had behavioral control over the
- horizontal displacement of the visual scene, a subset of spiking neurons, in the medulla,
746 The VR setups used are constantly evolving, and each study so far has used slightly different 747 parameters and materials (Schultheiss et al., 2017). Most importantly, the techniques used for visual stimulus presentation have changed from LCD to LED screens, as LCD screens do not 748 allow for easy control over parameters such as color, brightness, light polarization or flicker 749 750 frequency (D'Eath, 1998). LED bulbs allow precise control, and arrays of bulbs can be adjusted to match the visual resolution of bee eyes (Reiser and Dickinson, 2008). Another way to display 751 stimuli that is cheaper than LED arrays and more flexible than LED screens is the use of video 752 753 projectors, which offer similar control over color and brightness and can display images on screens of different shapes such as spheres or cylinders around the insect (Buatois et al., 2017, 754 2018). The latest development of VR in Drosophila allowed to build setups as cheap as \$300 755 (Loesche and Reiser, 2021). This setup could probably be adapted to bees by scaling up the 756 treadmill. 757

758 Studying associative learning and memory

The first two studies exploring visual learning and discrimination of honeybees walking 759 stationary on a trackball and facing virtual visual stimuli came out in 2017 (Buatois et al., 2017; 760 761 Rusch et al., 2017). In both cases, a visual projection system displayed different visual stimuli 762 on a semi-cylindrical screen placed in front of the tethered walking bee. After a pre-test assessing naïve preference for the two visual stimuli to be discriminated, bees were trained by 763 pairing one of them with appetitive sucrose solution and the other with aversive quinine 764 765 solution. Training was performed under open-loop conditions, presenting one stimulus at a time. Thus, the tethered bee had no control over the stimulus displacements on the screen. 766 767 Thereafter, bees were tested with the two visual stimuli displayed simultaneously and without reinforcement, to determine whether learning induced a change in the original preference. 768

Buatois et al. showed that when tested in extinction condition (without reward) after 12 trials 769 770 in open-loop around 60% of bees chose the CS+ while the rest either chose the CS- (20%) or didn't make a choice (20%) thus showing that the insects were able to learn the correct 771 772 association under open loop condition in the VR setup. They also showed that when presenting only the CS+ or CS- during the training phase it is not possible to record any discrimination 773 774 between CS+ and CS- as the spontaneous phototactic response of bees leads them to always 775 orientate towards the light (Buatois et al., 2017). Finally they showed that using distilled water 776 or quinine solution as punishment associated with the CS- was more effective to induce learning as bees submitted to either dry toothpick or NaCl solution associated with CS- did not learn the 777 778 discrimination (Buatois et al., 2017). Those results are really important to guide future conditioning protocol in choosing appropriate US, and suggest strongly to condition bees under 779 close-loop condition, in order to offer them a choice between CS+ and CS- at every trial to have 780 781 a chance to measure acquisition performance during the learning phase.

Similarly, Rusch et al showed that about 60% of bees were able to learn the association after 782 783 12 trials, going further they showed that 6 trials were sufficient to get more than 50% of bees to choose the correct stimulus. Bees were able to learn when CS+ and CS- differed both in 784 shape and colors or when both CS were circle of different colors but not when they were either 785 786 of the same color or square differing only in color. The authors conclude that the color and shape are learned in non-additive manner as not all combinations of shape-color variation lead 787 788 to learning and that they should thus be considered carefully when designing an experiment (Rusch et al., 2017). 789

As mentioned earlier, one of the problems potentially underlying the poor learning performances observed in visual PER conditioning is the restriction of active vision, which precluded – for instance – extracting the borders of objects to better detect their presence. In a further study, the question of the role of active vision in the VR setup described above was analyzed. Buatois et al. (Buatois et al., 2018) realized two transfer experiments in which the bees either learned the association within a miniature Y-maze (described earlier), where they were free to move and explore the visual stimuli projected on the back walls, or while being tethered and walking stationary in the VR setup, where the control of the visual stimuli was restricted to a 2D plane (bees could only displace the stimuli laterally to bring them in front of them or to move away from them). In either case, the transfer consisted in testing bees after their initial learning in the opposite setup (i.e. from Y-maze to VR or from VR to Y-maze).

Approximately 60% of the bees learned the visual discrimination in both conditions. Transfer 801 802 from VR to the maze improved significantly the bees' performances: 75% of bees having 803 chosen the rewarded stimulus (CS+) continued doing so and 100% of bees having chosen the punished stimulus (CS-) reverted their choice in favor of the CS+. In contrast, no improvement 804 was seen for these two groups of bees during the reciprocal transfer from the Y-maze to VR. In 805 this case, bees exhibited inconsistent choices in the VR setup. The asymmetric transfer between 806 contexts indicates that the information learned in each environment may be different despite 807 808 the similar learning success. Moreover, it shows that reducing the possibility of active vision 809 and movement freedom in the passage from the maze to the VR impairs the expression of visual 810 learning while increasing them in the reciprocal transfer improves it (Buatois et al., 2018). 811 These results underline the active nature of visual processing in bees and suggests that current VR systems require more work to increase immersion, like for example by adding looming and 812 optic flow to the virtual environment. 813

In 2019 Zwaka et al. published the first results showing electrophysiological recordings from higher order neurons of honeybees submitted to a visual differential conditioning using a VR system that consisted of an air-supported spherical treadmill allowing the stationary walking bee in closed-loop to control a visual environment projected onto a cone-shaped screen from above (Zwaka et al., 2019). They then used this setup to record A3 mushroom body extrinsic

neurons that are known to change their response properties during classical olfactory 819 820 conditioning (Haehnel and Menzel, 2010; Filla and Menzel, 2015) and receive input from Kenyon cells. These neurons provide GBAergic inhibitory feedback onto the mushroom body 821 822 calyces and regulate therefore Kenyon cell activity (Rybak and Menzel, 1993; Grünewald, 1999; Zwaka et al., 2018). They have been found to play a crucial role for non-elementary 823 discriminations in the olfactory domain (Boitard et al., 2015; Devaud et al., 2015a). After 824 conditioning within the VR, a significant increase in response from the recorded units in 825 826 reaction to the rewarded color was found. Yet, this increase was observed in animals for which no behavioral readout of learning was available, thus raising the question of the signification of 827 this variation in neural activity. 828

The interest of honeybees as a model for the study of learning and memory resides in the fact 829 that these animals cannot only solve simple discriminations; they can also solve complex visual 830 tasks relying on categorization, conceptual learning or numerosity (Giurfa et al., 2001; 831 Avarguès-Weber et al., 2011b, 2012a; Howard et al., 2019). A form of higher-order learning is 832 833 the so-called negative patterning discrimination in which a subject has to learn to respond to single stimuli (A, B) but not to their conjunction (AB) (Kehoe and Graham, 1988; Whitlow and 834 Wagner, 1972). The ambiguity of the task resides in the fact that each element (A and B) is as 835 836 often reinforced (when presented alone) as non-reinforced (when presented as a compound). Besides, the problem is difficult given the natural tendency to summation upon compound AB 837 presentation; in other words, if A and B were positively reinforced, the prediction is that AB 838 would be twice as good (Whitlow and Wagner, 1972). Yet, in this discrimination, individuals 839 have to inhibit this summation response and respond only to the single elements. A visual 840 841 version of this protocol was established in the VR environment (Buatois et al., 2020). It was shown that honeybees were able to solve a negative patterning task where A and B were green 842 843 and blue gratings against a dark background, while AB was a green-blue composite grating. When conditioned with rewarded green grating (A+) and rewarded blue grating (B+) versus the non-rewarded composite green and blue grating (AB-) (Fig11.B), 25% of the bees were able to solve both A vs AB and B vs AB tests and about 60% were able to solve at least one of the two. The relative low success is explained by the higher complexity of the task compared to associative learning. Nevertheless, a non-negligible number of bees were able to solve the task thereby proving that honeybees can solve a negative patterning task in VR.



850

- Figure 11. Virtual reality set-up and visual stimuli used for negative patterning. (a) Global
 view of the virtual reality system. (b) Conditioned stimuli: green grating (A), blue grating (B)
- and composite green-blue grating (AB). Taken from Buatois et al., 2020
- VR thus appears to be an appropriate tool for the study of visual learning as it was successfully
- used for both elemental association (Buatois et al., 2017; Rusch et al., 2017) and non-elemental
- learning (Buatois et al., 2020). Moreover, it shows great promises for live electrophysiological
- recording of learning bees (Rusch et al., 2021; Zwaka et al., 2019).

859 Benefits and caveats of using virtual reality to study visual learning in bees

One of the most important benefits of VR setups is the opportunity to combine controlled behavioral analyses with invasive analyses of underlying neural performances (Paulk et al., 2014b; Zwaka et al., 2019; Rusch et al., 2021), which is really difficult in freely-flying or moving insects (Paffhausen et al., 2020, 2021).

864 In VR, parameters of interest can be manipulated with great precision and flexibility, changing simple things like shape and color of stimuli (Rusch et al., 2017), or even creating conflict 865 between properties that are impossible in the natural world (Frasnelli et al., 2018). The 866 complexity of possible stimuli can vary greatly, ranging from very simple open-loop 867 presentations (Buatois et al., 2017; Rusch et al., 2017) to naturalistic, immersive, multimodal 868 scenarios in closed-loop (Fig.12) (Kócsi et al., 2020). Contrary to the real world, VR is under 869 the complete control of the experimenter, which allows precise control over both the timing of 870 stimuli, and the bees themselves, including their entire sensory exposure over the course of the 871 872 experiment (Schultheiss et al., 2017). This is very important for gene expression analysis as it allows to normalize the sensory experience across individuals and thus reduce noise and 873 unspecific response in the results. As tethered bees can be kept for long periods if they are fed 874 875 regularly and controlled for their motivation, this opens up the possibility of studying the neurobiological processes of long-term memory formation. 876



878

Figure 12. Illustration of a naturalistic VR: The Antarium. (Top) Picture of the fully
assembled Antarium. (Bottom) The landscape panorama projected by the Antarium LEDs seen
at 1.5 resolution, about twice the average resolution of ants. Taken from Kócsi et al., 2020.

VR systems still present some limitations. Learning success is reduced under such conditions 882 883 compared to performances of freely-flying bees, i.e. 60% versus the typical 90% to 100% 884 success rates of freely-flying bees trained to discriminate visual stimuli in Y-mazes (Buatois et al., 2017), this might be caused by the tethering that limits free movement and could induce 885 886 some stress in the insect. Moreover, transfer experiments showed an asymmetry in learning 887 success between real world and virtual reality conditions suggesting an important role of active vision in visual learning (Buatois et al., 2018). Thus, VR setup might need to offer the 888 889 possibility for insects to actively scan objects with not only closed loop conditions but also 890 adding a third dimension with looming and optic flow from a virtual background. Additionally, under tethered conditions, bees might be lacking some essential proprioceptive input (such as 891 antennal deflections during flight) for complete multisensory integration. Missing 892

mechanosensory input is known to influence responses of Drosophila to visual stimuli (Mureli 893 et al., 2017), and is believed to be a cause of 'cybersickness' in human applications of VR 894 (Rebenitsch and Owen, 2016). In some bees, tethering may also induce a decrease or a switch 895 896 from the appetitive motivation necessary for visual training to an escape motivation, which will interfere with learning. Also, when keeping bees tethered for longer periods, a proper control 897 of appetitive motivation is necessary as well as regular checks of normal motor behavior to 898 avoid fatigue effects (Schultheiss et al., 2017). Recent development in VR setups for ants also 899 underlined the need to use a treadmill of appropriate weight in order to make sure that the force 900 required by the insect to move the ball is similar to the force required to move its own weight 901 902 on a flat surface (Dahmen et al., 2017). This parameter seems to have been mostly overlooked by previous work on honeybees and could be a crucial first step in developing 3D VR as moving 903 the ball forward would require more force than rotating it around its vertical axis. 904

Finally, one also needs to account for the specific properties of the bee visual system when 905 designing VR setups (D'Eath, 1998; Fleishman and Endler, 2000). Bees have compound eyes 906 with photoreceptor sensitivities peaking in the green, blue and UV regions of the spectrum 907 (Backhaus, 1992). Common technology for creating visual stimuli is, however, designed for 908 human vision, in which, for example, yellow will be a blend of green and red. As bees cannot 909 910 perceive red light, their color perception of such stimulus will be very different (Fleishman et al., 1998). It is possible to produce images taking into account the properties of insects visual 911 system (Vorobyev et al., 1997), and this has been successfully done already in several VR 912 setups (Tedore and Johnsen, 2017; Kócsi et al., 2020). In addition, bee vision has a high 913 temporal resolution, almost 200 Hz (Srinivasan and Lehrer, 1984), which should therefore be 914 915 the minimum frequency of any VR display system. Flicker frequency is an important parameter as Poll et al. showed that honeybees payed more attention to LEDs flickering at 20-25 Hz, 916

while they avoided higher or lower frequencies (50–100 Hz and 2–4 Hz, respectively) (Poll et
al., 2015).

Using a very detailed VR that provides as much information as possible like a three-dimensional 919 920 landscape, with polarized light information and optic flow to produce a virtual world as close as possible to the visual setting in a free-flight experiment, could be a way to overcome the VR 921 922 limitations and produce a "perfect" virtual setting. But doing so would be particularly 923 expensive, in financial terms, but more importantly in terms of the time and technical skills required to build such a setup (Fig.12). On the other hand, we could investigate the importance 924 925 of each parameter like optic flow, depth, polarized light etc. on the bees' learning performances 926 in order to design the minimal VR necessary to produce the coherent behavior required for our purpose. This would be giving us more insight into the insect visual behavior and allow the 927 emergence of a simple VR paradigm that could be easily disseminated. Making VR more 928 accessible is important because it would allow teams with limited resources to also be able to 929 explore underlying mechanisms of visual learning. 930

While it is showing great promises, there's still work to be done in order for VR systems to realize their full potential. The technical bar of entry can be lowered, for example, through the diffusion of open source VR software. And, as suggested by transfer experiments (Buatois et al., 2018), we need a better understanding of the importance of parameters like optic flow on the ability of bees to learn efficiently in VR. It provides above all a valid approach to uncover the neuronal mechanisms of visual learning in bees

937 What do we know of the underlying mechanism of visual learning?

938 Now that we have established what tools were at our disposal and what work was needed to 939 improve them we need to identify what questions will benefit from the application of those 940 tools. Despite the difficulties mentioned earlier to access the brain of learning honeybees, their vision has been studied intensively in the past decades. By drawing from those experiments and
work done on other insects' models, four main regions appear de be involved in visual learning.
From the periphery to the center: the optic lobes, the ventrolateral neuropils, the central complex
and the mushroom bodies (Fig.13) (Ito et al., 2014b).

In this part we'll review the roles of these different structures for visual learning in order to

identify good path of exploration for VR experimentation.



947

The optic lobes are the first level of integration of visual information. It is a relay point for information which arrives from photoreceptors in the compound eyes (Kien and Menzel, 1977).

- 956 There are three types of photoreceptors, S,M, and L (for short-, mid-, and long-range
- wavelength), peaking in the UV (344 nm), blue (436 nm), and green (544 nm) regions of the
- 958 spectrum, respectively, which have been identified in the honeybee retina (Menzel et al., 1986;
- 959 Menzel, 1979; Peitsch et al., 1992). Photoreceptors are connected to the lamina, the outermost

⁹⁴⁸ **Figure 13. The different visual neuronal populations and pathways of the honeybee brain.** 949 The black arrow indicates color stimulation. La = lamina, χo = outer chiasm, me = medulla, χI 950 = inner chiasm, lo = lobula, le = lateral calyx of the mushroom bodies, me = median calyx, α = 951 alpha-lobe, β = beta-lobe, al = antennal lobe, ot = anterior optic tuberculum. MB: mushroom 952 bodies; CC: central complex. Courtesy of M. Giurfa.

⁹⁵³ The Optic lobes:

structure of the optic lobes. It is itself connected to the medulla which is connected to the lobula.
These three structures from the three layers of the optic lobes (Ribi, 1975; Avarguès-Weber et al., 2012b).

963 The lamina is the first visual neuropil in which the axons of the photoreceptors connect to first order processing interneurons, the lamina monopolar cells (LMC) (Menzel, 1974). In 964 honeybees, the lamina was shown to contain mainly neurons exhibiting relatively little response 965 966 variation across a wide range of wavelengths (Menzel, 1974; Ribi, 1975; Kien and Menzel, 1977). This neuropil is made of thousands of optical cartridges, each receiving an axon bundle 967 (containing the nine photoreceptor cell axons) from the overlying ommatidium, as well as the 968 969 axons of four different types of monopolar cells. The spatial arrangement of photoreceptor axons and LMCs within a cartridge remains constant throughout the lamina, thus retaining the 970 retinotopic organization. The outer chiasm forms the connection between the lamina and the 971 second visual neuropil, the medulla, a structure that contains most of the bee visual system 972 neurons (Ribi and Scheel, 1981). 973

974 Fibers coming from the anterior part of the lamina project to the posterior medulla while posterior fibers from the lamina project to the anterior medulla. Thus, the retinotopic 975 organization is retained but reversed in the medulla, which is also organized into a columnar 976 977 pattern. Medulla columns are highly connected by horizontal fibers (serotoninergic or GABAergic) in contrast with the lamina that has few horizontal connections (Ribi, 1975; Bicker 978 979 et al., 1987). In addition, the medulla exhibits a distal proximal laminated architecture consisting of eight identified layers, oriented orthogonally to the long axis of the columns (Ribi 980 and Scheel, 1981). Neurons in the medulla already respond with spectral opponency, i.e., with 981 982 opponent excitation or inhibition depending on photoreceptor-type input (Kien and Menzel, 1977). These color-opponent neurons, which exhibit combination-sensitive excitatory and/or 983

984 inhibitory interactions between two or three photoreceptor classes, represent the principal basis985 of color vision in honeybees.

The third visual neuropil is the lobula, where columnar stratification and retinotopic 986 987 organization are preserved mainly in the outer part (Hertel et al., 1987). The inner chiasm forms the connection between the medulla and the lobula, in which the retinotopic organization is 988 again reversed anteroposteriorly. Chromatic properties of neurons in the medulla are preserved 989 990 and amplified in the lobula, which was also shown to contain distinct color-opponent neurons (Kien and Menzel, 1977; Hertel, 1980; Hertel and Maronde, 1987). Moreover, different types 991 of spatial opponent neurons (i.e., with opponent excitation or inhibition depending on the visual 992 993 field region or on direction in which the stimulus is presented) were also described in the lobula (Hertel et al., 1987; Hertel and Maronde, 1987). 994

995 Inner-layer lobula and medulla neurons, which are more likely to exhibit color-sensitive responses, send projections to anterior brain areas, particularly to the mushroom bodies and the 996 997 anterior ventrolateral protocerebrum (Paulk et al., 2008; Paulk and Gronenberg, 2008; Paulk et 998 al., 2009a; Dyer et al., 2011). Thus, some of the major visual afferences to the mushroom bodies are color-sensitive (Gronenberg, 1986; Mauelshagen, 1993; Ehmer and Gronenberg, 2002). On 999 the other hand, outer lobula and both inner and outer medulla neurons, project to the posterior 1000 1001 protocerebrum (Paulk et al., 2009b, 2009a; Dyer et al., 2011). It seems therefore that achromatic and chromatic pathways are largely segregated in different steps of visual processing in the bee 1002 1003 brain. The optic lobes are thus involved in visual processing with the emergence of color vision 1004 and shape perception thanks to color opponent and spatial opponent neurons.

1005 The ventrolateral neuropils

In bees, the ventrolateral neuropils can be divided in at least five main regions: the anterioroptic tubercle (AOTu), the ventrolateral protocerebrum, the posteriorlateral protocerebrum, the

wedge and the posterior optic tubercle (Ito et al., 2014b). Most ventrolateral neuropils receive 1008 1009 visual input from the medulla and/or lobula and participate in visual processing (Paulk et al., 2009b). As mentioned before, anteroposterior segregation of achromatic and chromatic 1010 1011 processing was found in the input from the medulla and lobula to the ventrolateral protocerebrum of bees (Paulk et al., 2008, 2009b, 2009a; Dyer et al., 2011). Moreover, this 1012 1013 same gradient of achromatic/chromatic segregation in the anteroposterior brain axis seems to 1014 be retained at the level of ventrolateral protocerebrum neurons (Paulk et al., 2009b). The most prominent optic neuropil in the anterior region of the ventrolateral neuropils is the AOTu. The 1015 AOTu of bees is compartmentalized in four distinct units (Fig.14) (Theo Mota et al., 2011b). 1016 1017 The AOTu receives substantial input from the medulla and lobula via the anterior optic tract and send output to lateral accessory lobe via the tubercle accessory lobe tract (Fig.14). 1018 1019 Furthermore, two distinct tracts interconnect the AOTus of both brain hemispheres: the ventral 1020 inter-tubercle tract and the medial inter-tubercle tract (Fig.14). In addition to these four tracts, a specific neuron provides input from the vertical lobe of the mushroom bodies to the AOTu 1021 (Theo Mota et al., 2011b). 1022

Visual information from the dorsal and ventral parts of the bee eye segregate within different
AOTu compartments, both at the level of the visual input via the anterior optic tract and of the
visual output to the contralateral AOTu via intertubercle tracts (Theo Mota et al., 2011b).
Therefore, visual processing in the AOTu of bees includes a notable spatial component
characterized by this dorsoventral segregation.

In vivo calcium imaging revealed that stimulation with distinct monochromatic lights (ultraviolet [UV], blue, and green) matching the sensitivity of the three photoreceptor types of the bee retina induced different signal amplitudes, temporal dynamics, and spatial activity patterns in the AOTu intertubercle network, thus revealing intricate chromatic processing properties. Green light strongly activated both the dorsal and ventral lobes of the AOTu's major unit; blue light activated the dorsal lobe more while UV light activated the ventral lobe more.
Eye stimulation with mixtures of blue and green light induced suppression phenomena in which
responses to the mixture were lower than those to the color components, thus concurring with
color-opponent processing. These data reinforce strongly the idea that there is a spatial
segregation of color processing in the AOTu, which may serve for navigation purposes (Mota
et al., 2013).





- 1049 The central complex
- 1050 The central complex (CX) comprises a group of neuropils in the center of the insect brain. One
- 1051 important role of the CX is generation of motor outputs according to processed internal and
- 1052 external stimuli (Pfeiffer and Homberg, 2014; Plath and Barron, 2015). The CX is essential for

the initiation and termination of walking, turning and climbing behavior in fruit flies (Triphan et al., 2010), cockroaches (Martin et al., 2015) and crickets (Kai and Okada, 2013) and is considered as a site for action selection and goal-directed behavior (Barron and Klein, 2016;
Fiore et al., 2015). A role of the CX in visual learning of spatial features has been shown in various behavioral assays using fruit flies (Neuser et al., 2008; Ofstad et al., 2011; Kuntz et al., 2012, 2017). The CX is also important for polarized light processing and navigation (Pfeiffer and Homberg, 2014; Heinze, 2017).

Using a genetic approach in *Drosophila melanogaster*, it was shown that the fan-shaped body (FB), the largest component of the central complex, houses a memory trace for the pattern parameter 'elevation', and a memory trace for 'contour orientation'(Liu et al., 2006). A following study showed that blocking the ellipsoid body, which is another substructure of the CX connected to the FB, interferes with visual pattern memory (Pan et al., 2009).

In Cataglyphis noda ants a comparison of neuroanatomical changes in the central complex 1065 1066 before and after a learning walks revealed that, under natural light conditions (UV light together 1067 with a naturally changing polarization pattern), the CX undergoes a volume increase. While it is not clear whether or not the neuroanatomical changes found in the CX are triggered by 1068 appropriate sensory exposure or following the formation of spatial memory, those results still 1069 1070 suggest a potential involvement of the CX in visual learning (Grob et al., 2017). Honeybees 1071 with inactivated CX (Fig.15) were unable to avoid a shock paired light despite not displaying 1072 any motor deficit (Plath et al., 2017).

1073 The CX appears to play a role in visual learning in the context of navigation and spatial 1074 orientation as is it involved in processing celestial cues like polarized light, which is crucial for 1075 azimuthal orientation, but also in pattern recognition which is important to recognize 1076 landmarks.

1077 The mushroom bodies

The mushroom bodies contain 170,000 intrinsic neurons called Kenyon cells. At least three
sub-populations can be distinguished within these cells: small-type class I cells, large-type class
I cells and class II cells, more recently a third type called middle-type Kenyon cells was
identified (Kaneko et al., 2016).

1082 Quantifying the expression of Immediate Early Genes (IEGs) kakusei, Kiya et al. showed that 1083 neural activity of a the small-type Kenyon cells is prominently increased in the brains of dancer and forager honeybees. In contrast, the neural activity of the small-and large-type Kenyon cells 1084 is increased in the brains of re-orienting workers, which memorize their hive location during 1085 re-orienting flights. These findings demonstrate that the small-type Kenyon cell-preferential 1086 1087 activity is associated with foraging behavior, suggesting its involvement in information integration during foraging flight, and thus potentially in visual learning (Kiya et al., 2007). 1088 1089 IEGs are gene that are transcribed transiently and rapidly in response to specific stimulations 1090 inducing neural activity without de novo protein synthesis (Bahrami and Drabløs, 2016). Thanks to those properties they offer a good proxy of neuronal activity, in mammals, *c-fos*, 1091 1092 *zif268* and *Arc* are regularly used as such during learning, memory and other forms of cellular plasticity like as long-term potentiation (Minatohara et al., 2016; Gallo et al., 2018; He et al., 1093 2019). In insects, IEGs are used less often as the number of candidate genes serving this goal 1094 1095 is reduced and the reliable detection of their expression is sometimes difficult (Sommerlandt et al., 2019). In their 2007 study, Kiya et al identified such a candidate IEG, kakusei, and 1096 1097 established a method to use it as a marker of neural activity. The kakusei transcript is localized 1098 in the nuclei of neurons and does not encode an open reading frame, suggesting that it functions as a non-coding nuclear RNA (Kiya et al., 2007). 1099

Mushroom bodies are divided into two types of structures: calyces and lobes. The dendrites ofKenyon cells form the calyces and their axons form the pedunculus made up of two lobes

(Mobbs and Young, 1982; Mobbs, 1984; Strausfeld et al., 2003). Calyces are the input region 1102 1103 for various types of sensory information while the lobes act as the output of the system. There are two calves per mushroom body: a median calvx and a lateral calvx. These two structures 1104 1105 have direct neuronal connections with the medulla and the lobula (Paulk et al., 2008). They are made up of three sub-parts: the basal ring, the collar and the lip. The collar receives visual input 1106 , the lip receives the olfactory inputs and the basal ring receives both (Strube-Bloss and Rössler, 1107 1108 2018). The collar region of the calvx is segregated into five layers that receive alternating input from the dorsal or ventral medulla, respectively. A sixth, innermost layer of the collar receives 1109 input from lobula neurons. In the basal ring region of the calyx, medulla neuron terminals are 1110 1111 restricted to a small, distal part. Lobula neurons are more prominent in the basal ring, where they terminate in its outer half (Ehmer and Gronenberg, 2002). 1112

1113 The lobes of mushroom bodies are divided into two parts: vertical lobe and median lobe (Ito et 1114 al., 2014a), which are interconnected. Information from the calyces joins other structures of the 1115 bee brain by passing through these two lobes (Menzel, 1999).

1116 The mushroom bodies have been shown to be involved in both olfactory and visual learning (Komischke et al., 2005; Devaud et al., 2015b; Plath et al., 2017) although their implication in 1117 visual learning is less clear. Using the APIS assay described earlier, Plath et al. studied learning 1118 performance of bees in which different lobes of the mushroom bodies had been transiently 1119 inactivated by microinjection of the reversible anesthetic procaine. Control bees learned to 1120 escape the shock-paired light field and to spend more time in the safe light field after a few 1121 trials. When medial lobe neurons of the mushroom bodies were silenced, bees were no longer 1122 able to associate one light field with shock. By contrast, silencing of one collar region of the 1123 1124 mushroom body calyx (Fig15.C) did not alter behavior in the learning assay in comparison to control treatment (Plath et al., 2017). Those results are coherent with previous olfactory 1125

experiments that showed that inactivation of mushroom body lobes via the injection ofcholinergic antagonist disrupts memory retrieval (Lozano et al., 2001).



Figure 15. Procaine injection sites. (A) Alexa dye injections are shown in magenta (false 1129 color) in the MBC (left), VL (middle) and the CX (right). A DAPI-counterstain and auto-1130 fluorescence of the brain tissue (false colored in cyan) allowed us to identify brain neuropils. 1131 1132 Orientation of all three scans was aligned with rostral (neuraxis) facing upwards. Injections of vehicle (B) and of procaine solution (C) into the MBC. Injections into the VL (D). Injections 1133 1134 of vehicle (E) and of procaine solution (F) into the central body (red dots) and injections located at the border of the lower division of the central body with spread into the noduli (red dots with 1135 black border). MBC, mushroom body calyces; VL, ventral lobes; HL, horizontal lobes; CBU, 1136 upper division of the central body; CBL lower division of the central body; Scale bar = $30 \mu m$. 1137 Taken from Plath et al., 2017. 1138

1139 Similarly, Kamhi et al. studied the effect of procaine inaction of the vertical lobes (VL) of the

- 1140 mushroom bodies on view-based navigation in ants (Myrmecia midas). Experienced foragers
- 1141 were collected, treated, and released in their familiar environment where their behavior was
- 1142 documented. Animals with procaine-inactivated VLs had tortuous paths and were unable to
- 1143 find their nest, whereas control ants were well directed and were the most successful at returning
- 1144 home. Untreated animals walked faster when their gaze was directed toward home, and this

1147 Contrary to Plath et al. results on calyces, in an ant species (*Formica rufa*) lesions of the 1148 mushroom body calyces rendered the ants unable to go toward a previously trained feeder 1149 location (Buehlmann et al., 2020). The discrepancy between those results might be explained 1150 by the fact that Plath et al only silenced one collar region of the calyces which might have been 1151 compensated by the other three collars. More results suggest that the calyces play a part in 1152 visual learning, Li et al found a correlation between success in a visual discrimination task and 1153 microglomeruli density in the collar region of the MB (Li et al., 2017).

Moreover, when bees are trained in the ICARUS setup (described earlier) to inhibit their spontaneous phototaxis by pairing the attracting light with an electric shock (Marchal et al., 2019), learning induced an increase in the dopaminergic receptor gene *Amdop1* in the calyces of the mushroom bodies, consistently with the role of dopaminergic signaling for electric-shock representation in this region of the brain (Unoki et al., 2005; Vergoz et al., 2007; Mizunami et al., 2009; Agarwal et al., 2011; Tedjakumala et al., 2013).

In *Drosophila melanogaster* it was found that inhibition from a single pair of giant GABAergic neurons, the anterior paired lateral (APL) neurons, onto the mushroom bodies (MBs) selectively facilitates behavioral flexibility during visual reversal learning. Indeed, acute disruption of the APL–MB circuit was sufficient to impair visual reversal learning, while flies with dysfunctional APL–MB circuit performed normally in simple forms of visual learning (Ren et al., 2012).

In honeybees inhibition of the MBs was also shown to specifically impair olfactory reversal
learning (Devaud et al., 2007; Boitard et al., 2015). GABAergic inhibitory feedback on the MBs
is provided by A3v and A3d neurons (Bicker et al., 1985; Gronenberg, 1987; Grünewald, 1999).
Both innervate the output region of the MBs (the lobes) but A3v neurons also feedback onto
the input region (the calyces). A3 neurons have been shown to change their response to

rewarded and unrewarded visual stimulus after conditioning (Zwaka et al., 2019). And similarly 1170 1171 to APL neuron in Drosophila, it was shown that inhibition of GABAergic signaling into the MB calyces impairs olfactory reversal learning, but leaves intact the capacity to perform 1172 1173 elemental olfactory conditioning (Boitard et al., 2015; Devaud et al., 2015b). However, inhibition of GABAergic signaling into the lobes instead of the calves had no effect on reversal 1174 learning (Boitard et al., 2015). Even though these experiments were conducted with olfactory 1175 1176 conditioning, it is reasonable to expect similar results for vision as centralization of similar brain functions spares the cost of maintaining similar circuit motifs in different brain areas. 1177 Indeed, in Drosophila it was found that the same subsets of dopaminergic MB neurons drive 1178 1179 formation of both olfactory and visual memories (Vogt et al., 2014). Furthermore, distinct yet partially overlapping subsets of mushroom body intrinsic neurons were shown to be required 1180 for visual and olfactory memories (Vogt et al., 2014). This convergence of different modality 1181 1182 might be an evolutionary conserved design of information processing as such converging inputs of different stimuli into a multisensory area have even been described in humans (Beauchamp 1183 et al., 2008). GABAergic feedback between the lobes (output) and calyces (input) was also 1184 1185 shown to be involved in visual context learning and neural error responses following erroneous behavior (Filla and Menzel, 2015). 1186

1187 Thus the MBs appear to be major integration centers in the honeybee's brain, involved in1188 multiple forms of learning from simple association to more complex reversal learning.

1189 Whole circuit mechanisms

Despite the consequent body of evidence pointing to the MBs as a center for learning and memory, they are not the only structure of the visual system involved in memory formation. A recent study quantifying gene expression kinetic in the brains of honeybees after aversive visual conditioning showed a parallel activation of the optic lobes and the MBs following a similar time course (Avalos et al., 2021). This suggest that sensory neuropils are also involved inassociative learning.

1196 In order to explore the implication of the peripheral processing stages and high-order integration 1197 centers of the insect brain in visual learning, Yilmaz et al. quantified the volumetric changes in 1198 different neuropils directly after color conditioning and, 3 days later, after the establishment of long-term memory (LTM), in Camponotus blandus ants. They found a volume increase of the 1199 1200 OLs, the AOTu, and the fan-shaped body (FB) and protocerebral bridge (PB) which are neuropil of the CX, after color learning and LTM formation. They did not find any specific structural 1201 1202 change in the MBs (Yilmaz et al., 2019), which is coherent with findings in honeybees where 1203 no changes of the number of presynaptic buttons in the collar was found after fine color discrimination (Sommerlandt et al., 2016). However, those results could be explained by the 1204 1205 absence of memory formation in the tested bees as the authors did not test for long term memory. Yilmaz et al. results are also coherent with results mentioned earlier where a role of 1206 the VL but not the collar was suggested for aversive color learning after procaine injections in 1207 1208 the respective areas (Plath et al., 2017).

A volume increase in the OLs might affect processing of color information at the level of color 1209 opponency, the volume of the OLs increased significantly after LTM formation, which may 1210 1211 increase and strengthen the excitatory neuronal connections that are relevant for discrimination behavior (Yang et al., 2004). These results are coherent with the parallel activation of the OLs 1212 with the MB found in honeybees during visual learning (Avalos et al., 2021). Changes in the 1213 volume of the FB, suggest it is a potential region in the CX involved in visual memory formation 1214 after associative color learning. This is consistent with previous findings in Drosophila that 1215 1216 implicate the FB in visual pattern memory formation (Liu et al., 2006; Pan et al., 2009). The volume increase of the AOTu found in ants is coherent with results in bees and locusts, where 1217

the upper unit of the AOTu has been implicated in the processing of chromatic information(Pfeiffer and Homberg, 2007; Mota et al., 2013, 2016).

Thus the MBs appear to be the best candidate for further exploration of the neural basis of visual 1220 learning as they have been shown to play a role in both simple associative conditioning (Plath 1221 et al., 2017) and more complex reversal learning (Devaud et al., 2007; Ren et al., 2012). 1222 1223 However, we can also see that learning actually involves the whole system, from volume modification measured in the optic lobes (Yilmaz et al., 2019) to impaired response to learn 1224 stimulus by silencing the CX (Plath et al., 2017). As such whole brain analysis of the effect of 1225 visual learning on the activation of those different structure, using for example immediate early 1226 1227 gene like *kakusei* (Kiya et al., 2007) as markers, appears like a promising path.

1229 **References**

- Abel, R., Rybak, J., Menzel, R., 2001. Structure and response patterns of olfactory interneurons in the
 honeybee, Apis mellifera. J. Comp. Neurol. 437, 363–383. https://doi.org/10.1002/cne.1289
 Agarwal, M., Giannoni Guzmán, M., Morales-Matos, C., Del Valle Díaz, R.A., Abramson, C.I., Giray, T.,
- 1232Agarwai, M., Glamon Guzman, M., Morales-Matos, C., Der Valle Diaz, K.A., Abranson, C.I., Giray, T.,12332011. Dopamine and Octopamine Influence Avoidance Learning of Honey Bees in a Place1234Preference Assay. PLoS ONE 6. https://doi.org/10.1371/journal.pone.0025371
- Avalos, A., Traniello, I.M., Pérez Claudio, E., Giray, T., 2021. Parallel mechanisms of visual memory
 formation across distinct regions of the honey bee brain. J. Exp. Biol. 224, jeb242292.
 https://doi.org/10.1242/jeb.242292
- Avarguès-Weber, A., de Brito Sanchez, M.G., Giurfa, M., Dyer, A.G., 2010. Aversive Reinforcement
 Improves Visual Discrimination Learning in Free-Flying Honeybees. PLoS ONE 5.
 https://doi.org/10.1371/journal.pone.0015370
- Avarguès-Weber, A., Deisig, N., Giurfa, M., 2011a. Visual Cognition in Social Insects. Annu. Rev.
 Entomol. 56, 423–443. https://doi.org/10.1146/annurev-ento-120709-144855
- Avarguès-Weber, A., Dyer, A.G., Combe, M., Giurfa, M., 2012a. Simultaneous mastering of two
 abstract concepts by the miniature brain of bees. Proc. Natl. Acad. Sci. U. S. A. 109, 7481–
 7486. https://doi.org/10.1073/pnas.1202576109
- Avarguès-Weber, A., Dyer, A.G., Giurfa, M., 2011b. Conceptualization of above and below
 relationships by an insect. Proc. R. Soc. B Biol. Sci. 278, 898–905.
 https://doi.org/10.1098/rspb.2010.1891
- Avarguès-Weber, A., Giurfa, M., 2013. Conceptual learning by miniature brains. Proc. Biol. Sci. 280,
 20131907. https://doi.org/10.1098/rspb.2013.1907
- Avarguès-Weber, A., Mota, T., 2016. Advances and limitations of visual conditioning protocols in
 harnessed bees. J. Physiol.-Paris, SI: Neuroethology 2016 110, 107–118.
 https://doi.org/10.1016/j.jphysparis.2016.12.006
- Avarguès-Weber, A., Mota, T., Giurfa, M., 2012b. New vistas on honey bee vision. Apidologie 43,
 244–268. https://doi.org/10.1007/s13592-012-0124-2
- 1256 Backhaus, W., 1992. Color vision in honeybees. Neurosci. Biobehav. Rev. 16, 1–12.
- 1257 https://doi.org/10.1016/S0149-7634(05)80045-4
- 1258Bahrami, S., Drabløs, F., 2016. Gene regulation in the immediate-early response process. Adv. Biol.1259Regul. 62, 37–49. https://doi.org/10.1016/j.jbior.2016.05.001
- Baird, E., Boeddeker, N., Srinivasan, M.V., 2021. The effect of optic flow cues on honeybee flight
 control in wind. Proc. R. Soc. B Biol. Sci. 288, 20203051.
 https://doi.org/10.1098/rspb.2020.3051
- Baird, E., Dacke, M., 2012. Visual flight control in naturalistic and artificial environments. J. Comp.
 Physiol. A 198, 869–876. https://doi.org/10.1007/s00359-012-0757-7
- Baird, E., Srinivasan, M.V., Zhang, S., Lamont, R., Cowling, A., 2006. Visual Control of Flight Speed and
 Height in the Honeybee, in: Nolfi, S., Baldassarre, G., Calabretta, R., Hallam, J.C.T., Marocco,
 D., Meyer, J.-A., Miglino, O., Parisi, D. (Eds.), From Animals to Animats 9, Lecture Notes in
 Computer Science. Springer, Berlin, Heidelberg, pp. 40–51.
- 1269 https://doi.org/10.1007/11840541_4
- Balamurali, G.S., Somanathan, H., Hempel de Ibarra, N., 2015. Motion cues improve the performance
 of harnessed bees in a colour learning task. J. Comp. Physiol. A 201, 505–511.
 https://doi.org/10.1007/s00359-015-0994-7
- Barron, A.B., Klein, C., 2016. What insects can tell us about the origins of consciousness. Proc. Natl.
 Acad. Sci. 113, 4900–4908. https://doi.org/10.1073/pnas.1520084113
- 1275Beauchamp, M.S., Yasar, N.E., Frye, R.E., Ro, T., 2008. Touch, sound and vision in human superior1276temporal sulcus. NeuroImage 41, 1011–1020.
- 1277 https://doi.org/10.1016/j.neuroimage.2008.03.015

- Bestea, L., Briard, E., Carcaud, J., Sandoz, J.-C., Velarde, R., Giurfa, M., Maria Gabriela, de B.S., 2022.
 The short neuropeptide F (sNPF) promotes the formation of appetitive visual memories in honey bees. Biol. Lett. (in press).
- Bicker, G., Schäfer, S., Kingan, T.G., 1985. Mushroom body feedback interneurones in the honeybee
 show GABA-like immunoreactivity. Brain Res. 360, 394–397. https://doi.org/10.1016/0006 8993(85)91262-4
- Bicker, G., Schäfer, S., Rehder, V., 1987. Chemical Neuroanatomy of the Honeybee Brain, in: Menzel,
 R., Mercer, A. (Eds.), Neurobiology and Behavior of Honeybees. Springer, Berlin, Heidelberg,
 pp. 202–224. https://doi.org/10.1007/978-3-642-71496-2_17
- Bitterman, M.E., Menzel, R., Fietz, A., Schäfer, S., 1983. Classical conditioning of proboscis extension
 in honeybees (Apis mellifera). J. Comp. Psychol. Wash. DC 1983 97, 107–119.
- Boitard, C., Devaud, J.-M., Isabel, G., Giurfa, M., 2015. GABAergic feedback signaling into the calyces
 of the mushroom bodies enables olfactory reversal learning in honey bees. Front. Behav.
 Neurosci. 9, 198. https://doi.org/10.3389/fnbeh.2015.00198
- Breed, M.D., Guzmán-Novoa, E., Hunt, G.J. 3, 2004. Defensive behavior of honey bees: organization,
 genetics, and comparisons with other bees. Annu. Rev. Entomol. 49, 271–298.
 https://doi.org/10.1146/annurev.ento.49.061802.123155
- Buatois, A., Flumian, C., Schultheiss, P., Avarguès-Weber, A., Giurfa, M., 2018. Transfer of Visual
 Learning Between a Virtual and a Real Environment in Honey Bees: The Role of Active Vision.
 Front. Behav. Neurosci. 12, 139. https://doi.org/10.3389/fnbeh.2018.00139
- Buatois, A., Laroche, L., Lafon, G., Avarguès-Weber, A., Giurfa, M., 2020. Higher-order discrimination
 learning by honeybees in a virtual environment. Eur. J. Neurosci. 51, 681–694.
 https://doi.org/10.1111/ejn.14633
- Buatois, A., Pichot, C., Schultheiss, P., Sandoz, J.-C., Lazzari, C.R., Chittka, L., Avarguès-Weber, A.,
 Giurfa, M., 2017. Associative visual learning by tethered bees in a controlled visual
 environment. Sci. Rep. 7. https://doi.org/10.1038/s41598-017-12631-w
- Buehlmann, C., Wozniak, B., Goulard, R., Webb, B., Graham, P., Niven, J.E., 2020. Mushroom Bodies
 Are Required for Learned Visual Navigation, but Not for Innate Visual Behavior, in Ants. Curr.
 Biol. CB 30, 3438-3443.e2. https://doi.org/10.1016/j.cub.2020.07.013
- Carcaud, J., Giurfa, M., Sandoz, J.C., 2016. Parallel Olfactory Processing in the Honey Bee Brain: Odor
 Learning and Generalization under Selective Lesion of a Projection Neuron Tract. Front.
 Integr. Neurosci. 9. https://doi.org/10.3389/fnint.2015.00075
- 1310 Chittka, L., Geiger, K., 1995. Can honey bees count landmarks? Anim. Behav. 49, 159–164.
 1311 https://doi.org/10.1016/0003-3472(95)80163-4
- 1312 Chittka, L., Thomson, J., Waser, N., 1999. Flower Constancy, Insect Psychology, and Plant Evolution.
 1313 Naturwissenschaften 86, 361–377. https://doi.org/10.1007/s001140050636
- 1314 Dacke, M., Srinivasan, M.V., 2008. Evidence for counting in insects. Anim. Cogn. 11, 683–689.
 1315 https://doi.org/10.1007/s10071-008-0159-y
- Dahmen, H., Wahl, V.L., Pfeffer, S.E., Mallot, H.A., Wittlinger, M., 2017. Naturalistic path integration
 of Cataglyphis desert ants on an air-cushioned lightweight spherical treadmill. J. Exp. Biol.
 220, 634–644. https://doi.org/10.1242/jeb.148213
- D'Eath, R.B., 1998. Can video images imitate real stimuli in animal behaviour experiments? Biol. Rev.
 Camb. Philos. Soc. 73, 267–292. https://doi.org/10.1017/S0006323198005179
- Devaud, J.-M., Blunk, A., Podufall, J., Giurfa, M., Grünewald, B., 2007. Using local anaesthetics to
 block neuronal activity and map specific learning tasks to the mushroom bodies of an insect
 brain. Eur. J. Neurosci. 26, 3193–3206. https://doi.org/10.1111/j.1460-9568.2007.05904.x
- Devaud, J.-M., Papouin, T., Carcaud, J., Sandoz, J.-C., Grünewald, B., Giurfa, M., 2015a. Neural
 substrate for higher-order learning in an insect: Mushroom bodies are necessary for
 configural discriminations. Proc. Natl. Acad. Sci. U. S. A. 112, E5854–E5862.
 https://doi.org/10.1073/pnas.1508422112
- 1328Devaud, J.-M., Papouin, T., Carcaud, J., Sandoz, J.-C., Grünewald, B., Giurfa, M., 2015b. Neural1329substrate for higher-order learning in an insect: Mushroom bodies are necessary for

1330 configural discriminations. Proc. Natl. Acad. Sci. U. S. A. 112, E5854–E5862. 1331 https://doi.org/10.1073/pnas.1508422112 Dobrin, S.E., Fahrbach, S.E., 2012. Visual Associative Learning in Restrained Honey Bees with Intact 1332 1333 Antennae. PLOS ONE 7, e37666. https://doi.org/10.1371/journal.pone.0037666 Doumas, L.A.A., Hummel, J.E., Sandhofer, C.M., 2008. A theory of the discovery and predication of 1334 relational concepts. Psychol. Rev. 115, 1–43. https://doi.org/10.1037/0033-295X.115.1.1 1335 Dyer, A.G., 2012. The mysterious cognitive abilities of bees: why models of visual processing need to 1336 1337 consider experience and individual differences in animal performance. J. Exp. Biol. 215, 387-1338 395. https://doi.org/10.1242/jeb.038190 1339 Dyer, A.G., Neumeyer, C., 2005. Simultaneous and successive colour discrimination in the honeybee (Apis mellifera). J. Comp. Physiol. A 191, 547-557. https://doi.org/10.1007/s00359-005-1340 1341 0622-z 1342 Dyer, A.G., Paulk, A.C., Reser, D.H., 2011. Colour processing in complex environments: insights from the visual system of bees. Proc. R. Soc. B Biol. Sci. 278, 952–959. 1343 1344 https://doi.org/10.1098/rspb.2010.2412 1345 Ehmer, B., Gronenberg, W., 2002. Segregation of visual input to the mushroom bodies in the 1346 honeybee (Apis mellifera). J. Comp. Neurol. 451, 362–373. 1347 https://doi.org/10.1002/cne.10355 Felsenberg, J., Gehring, K.B., Antemann, V., Eisenhardt, D., 2011. Behavioural Pharmacology in 1348 Classical Conditioning of the Proboscis Extension Response in Honeybees (Apis mellifera). 1349 JoVE J. Vis. Exp. e2282-e2282. https://doi.org/10.3791/2282 1350 1351 Filla, I., Menzel, R., 2015. Mushroom body extrinsic neurons in the honeybee (Apis mellifera) brain 1352 integrate context and cue values upon attentional stimulus selection. J. Neurophysiol. 114, 1353 2005–2014. https://doi.org/10.1152/jn.00776.2014 1354 Fiore, V.G., Dolan, R.J., Strausfeld, N.J., Hirth, F., 2015. Evolutionarily conserved mechanisms for the 1355 selection and maintenance of behavioural activity. Philos. Trans. R. Soc. B Biol. Sci. 370, 1356 20150053. https://doi.org/10.1098/rstb.2015.0053 1357 Fleishman, L.J., Endler, J.A., 2000. Some comments on visual perception and the use of video 1358 playback in animal behavior studies. Acta Ethologica 3, 15–27. 1359 Fleishman, L.J., McClintock, W.J., D'Eath, R.B., Brainard, D.H., Endler, J.A., 1998. Colour perception 1360 and the use of video playback experiments in animal behaviour. Anim. Behav. 56, 1035-1361 1040. https://doi.org/10.1006/anbe.1998.0894 Frasnelli, E., Hempel de Ibarra, N., Stewart, F.J., 2018. The Dominant Role of Visual Motion Cues in 1362 1363 Bumblebee Flight Control Revealed Through Virtual Reality. Front. Physiol. 9. https://doi.org/10.3389/fphys.2018.01038 1364 Frings, H., 1944. The loci of olfactory end-organs in the honey-bee, Apis mellifera Linn. J. Exp. Zool. 1365 1366 97, 123–134. https://doi.org/10.1002/jez.1400970203 1367 Frings, H., Frings, M., 1949. The Loci of Contact Chemoreceptors in Insects. A Review with New 1368 Evidence. Am. Midl. Nat. 41, 602–658. https://doi.org/10.2307/2421776 1369 Frisch, K. von, 1886-, Frisch, K. von, 1886-, 1967. Dance language and orientation of bees. Harvard 1370 University Press. 1371 Gallo, F.T., Katche, C., Morici, J.F., Medina, J.H., Weisstaub, N.V., 2018. Immediate Early Genes, 1372 Memory and Psychiatric Disorders: Focus on c-Fos, Egr1 and Arc. Front. Behav. Neurosci. 12, 1373 79. https://doi.org/10.3389/fnbeh.2018.00079 1374 Giurfa, M., 2007. Behavioral and neural analysis of associative learning in the honeybee: a taste from 1375 the magic well. J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol. 193, 801–824. 1376 https://doi.org/10.1007/s00359-007-0235-9 Giurfa, M., 2004. Conditioning procedure and color discrimination in the honeybee Apis mellifera. 1377 1378 Sci. Nat. 5, 228–231. https://doi.org/10.1007/s00114-004-0530-z Giurfa, Martin, Eichmann, B., Menzel, R., 1996. Symmetry perception in an insect. Nature 382, 458-1379 1380 461. https://doi.org/10.1038/382458a0

1381 Giurfa, M., Menzel, R., 1997. Insect visual perception: complex abilities of simple nervous systems. 1382 Curr. Opin. Neurobiol. 7, 505–513. https://doi.org/10.1016/S0959-4388(97)80030-X Giurfa, M., Sandoz, J.-C., 2012. Invertebrate learning and memory: Fifty years of olfactory 1383 1384 conditioning of the proboscis extension response in honeybees. Learn. Mem. 19, 54–66. https://doi.org/10.1101/lm.024711.111 1385 1386 Giurfa, M., Vorobyev, M., Kevan, P., Menzel, R., 1996. Detection of coloured stimuli by honeybees: Minimum visual angles and receptor specific contrasts. J. Comp. Physiol. [A] 178, 699–709. 1387 1388 https://doi.org/10.1007/BF00227381 Giurfa, M., Zhang, S., Jenett, A., Menzel, R., Srinivasan, M.V., 2001. The concepts of 'sameness' and 1389 1390 'difference' in an insect. Nature 410, 930–933. https://doi.org/10.1038/35073582 Götz, K.G., 1964. Optomotorische Untersuchung des visuellen systems einiger Augenmutanten der 1391 1392 Fruchtfliege Drosophila. Kybernetik 2, 77–92. https://doi.org/10.1007/BF00288561 1393 Grant, V., 1951. The fertilization of flowers. Sci. Am. 184, 52–57. 1394 Grob, R., Fleischmann, P.N., Grübel, K., Wehner, R., Rössler, W., 2017. The Role of Celestial Compass 1395 Information in Cataglyphis Ants during Learning Walks and for Neuroplasticity in the Central 1396 Complex and Mushroom Bodies. Front. Behav. Neurosci. 11, 226. 1397 https://doi.org/10.3389/fnbeh.2017.00226 1398 Gronenberg, W., 1987. Anatomical and physiological properties of feedback neurons of the mushroom bodies in the bee brain. Exp. Biol. 46, 115–125. 1399 1400 Gronenberg, W., 1986. Physiological and anatomical properties of optical input-fibres to the 1401 mushroom body in the bee brain. J. Insect Physiol. 32, 695–704. 1402 https://doi.org/10.1016/0022-1910(86)90111-3 1403 Gross, H.J., Pahl, M., Si, A., Zhu, H., Tautz, J., Zhang, S., 2009. Number-Based Visual Generalisation in 1404 the Honeybee. PLoS ONE 4, e4263. https://doi.org/10.1371/journal.pone.0004263 1405 Grünewald, B., 1999. Physiological properties and response modulations of mushroom body 1406 feedback neurons during olfactory learning in the honeybee, Apis mellifera. J. Comp. Physiol. 1407 A 185, 565–576. https://doi.org/10.1007/s003590050417 1408 Guerrieri, F., Schubert, M., Sandoz, J.-C., Giurfa, M., 2005. Perceptual and Neural Olfactory Similarity 1409 in Honeybees. PLOS Biol. 3, e60. https://doi.org/10.1371/journal.pbio.0030060 1410 Haehnel, M., Menzel, R., 2010. Sensory representation and learning-related plasticity in mushroom 1411 body extrinsic feedback neurons of the protocerebral tract. Front. Syst. Neurosci. 4, 161. 1412 https://doi.org/10.3389/fnsys.2010.00161 1413 Halford, G.S., Wilson, W.H., Phillips, S., 2010. Relational knowledge: the foundation of higher 1414 cognition. Trends Cogn. Sci. 14, 497–505. https://doi.org/10.1016/j.tics.2010.08.005 1415 Hassenstein, B., 1951. Ommatidienraster und afferente Bewegungsintegration. Z. Für Vgl. Physiol. 33, 1416 301-326. https://doi.org/10.1007/BF00339334 Hassenstein, B., 1950. Wandernde geometrische Interferenzfiguren im Insektenauge. 1417 1418 Naturwissenschaften 37, 45–46. https://doi.org/10.1007/BF00645369 Hateren, J.H. van, Srinivasan, M.V., Wait, P.B., 1990. Pattern recognition in bees: orientation 1419 1420 discrimination. J. Comp. Physiol. A 167, 649-654. 1421 He, Q., Wang, J., Hu, H., 2019. Illuminating the Activated Brain: Emerging Activity-Dependent Tools to Capture and Control Functional Neural Circuits. Neurosci. Bull. 35, 369–377. 1422 1423 https://doi.org/10.1007/s12264-018-0291-x 1424 Heinze, S., 2017. Unraveling the neural basis of insect navigation. Curr. Opin. Insect Sci., 1425 Neuroscience * Pheromones 24, 58-67. https://doi.org/10.1016/j.cois.2017.09.001 1426 Hertel, H., 1980. Chromatic properties of identified interneurons in the optic lobes of the bee. J. 1427 Comp. Physiol. 137, 215-231. https://doi.org/10.1007/BF00657117 1428 Hertel, H., Maronde, U., 1987. The physiology and morphology of centrally projecting visual 1429 interneurons in the honey bee brain. J Exp Biol 133, 301–315. 1430 Hertel, H., Schafer, S., Maronde, U., 1987. The physiology and morphology of centrally projecting 1431 visual interneurones in the honeybee brain. J Exp Biol 133, 283–300.

- 1432 Hess, C., 1911. Experimentelle Untersuchungen zur vergleichenden Physiologie des Gesichtssinnes. 1433 Pflüg. Arch. Für Gesamte Physiol. Menschen Tiere 142, 405–446. 1434 https://doi.org/10.1007/BF01680657
- 1435 Hori, S., Takeuchi, H., Arikawa, K., Kinoshita, M., Ichikawa, N., Sasaki, M., Kubo, T., 2006. Associative visual learning, color discrimination, and chromatic adaptation in the harnessed honeybee 1436 1437 Apis mellifera L. J. Comp. Physiol. A 192, 691–700. https://doi.org/10.1007/s00359-005-1438 0091-4
- 1439 Hori, S., Takeuchi, H., Kubo, T., 2007. Associative learning and discrimination of motion cues in the 1440 harnessed honeybee Apis mellifera L. J. Comp. Physiol. A 193, 825–833. 1441 https://doi.org/10.1007/s00359-007-0234-x
- 1442 Horridge, A., 2009. Generalization in visual recognition by the honeybee (Apis mellifera): A review 1443 and explanation. J. Insect Physiol. 55, 499-511. 1444
 - https://doi.org/10.1016/j.jinsphys.2009.03.006
- 1445 Horridge, A., 2000. Seven experiments on pattern vision of the honeybee, with a model. Vision Res. 1446 40, 2589–2603. https://doi.org/10.1016/S0042-6989(00)00096-1
- 1447 Horridge, G.A., 1987. The evolution of visual processing and the construction of seeing systems. Proc. 1448 R. Soc. Lond. B Biol. Sci. 230, 279–292. https://doi.org/10.1098/rspb.1987.0020
- 1449 Howard, S.R., Avarguès-Weber, A., Garcia, J.E., Greentree, A.D., Dyer, A.G., 2019. Numerical cognition 1450 in honeybees enables addition and subtraction. Sci. Adv. 5, eaav0961. 1451 https://doi.org/10.1126/sciadv.aav0961
- 1452 Ito, K., Shinomiya, K., Ito, M., Armstrong, J.D., Boyan, G., Hartenstein, V., Harzsch, S., Heisenberg, M., 1453 Homberg, U., Jenett, A., Keshishian, H., Restifo, L.L., Rössler, W., Simpson, J.H., Strausfeld, 1454 N.J., Strauss, R., Vosshall, L.B., 2014a. A Systematic Nomenclature for the Insect Brain. 1455 Neuron 81, 755–765. https://doi.org/10.1016/j.neuron.2013.12.017
- 1456 Ito, K., Shinomiya, K., Ito, M., Armstrong, J.D., Boyan, G., Hartenstein, V., Harzsch, S., Heisenberg, M., 1457 Homberg, U., Jenett, A., Keshishian, H., Restifo, L.L., Rössler, W., Simpson, J.H., Strausfeld, 1458 N.J., Strauss, R., Vosshall, L.B., Insect Brain Name Working Group, 2014b. A systematic 1459 nomenclature for the insect brain. Neuron 81, 755–765.
- 1460 https://doi.org/10.1016/j.neuron.2013.12.017
- 1461 Jernigan, C.M., Roubik, D.W., Wcislo, W.T., Riveros, A.J., 2014. Color-dependent learning in restrained 1462 Africanized honey bees. J. Exp. Biol. 217, 337–343. https://doi.org/10.1242/jeb.091355
- 1463 Kai, K., Okada, J., 2013. Characterization of Locomotor-Related Spike Activity in Protocerebrum of 1464 Freely Walking Cricket. Zoolog. Sci. 30, 591–601. https://doi.org/10.2108/zsj.30.591
- 1465 Kamhi, J.F., Barron, A.B., Narendra, A., 2020. Vertical Lobes of the Mushroom Bodies Are Essential for 1466 View-Based Navigation in Australian Myrmecia Ants. Curr. Biol. CB 30, 3432-3437.e3. 1467 https://doi.org/10.1016/j.cub.2020.06.030
- 1468 Kaneko, K., Suenami, S., Kubo, T., 2016. Gene expression profiles and neural activities of Kenyon cell 1469 subtypes in the honeybee brain: identification of novel 'middle-type' Kenyon cells. Zool. Lett. 1470 2, 14. https://doi.org/10.1186/s40851-016-0051-6
- 1471 Kehoe, E.J., Graham, P., 1988. Summation and configuration: Stimulus compounding and negative 1472 patterning in the rabbit. J. Exp. Psychol. Anim. Behav. Process. 14, 320–333. 1473 https://doi.org/10.1037/0097-7403.14.3.320
- 1474 Kien, J., Menzel, R., 1977. Chromatic properties of interneurons in the optic lobes of the bee. J. 1475 Comp. Physiol. 113, 17–34. https://doi.org/10.1007/BF00610451
- 1476 Kirchner, W.H., Srinivasan, M.V., 1989. Freely flying honeybees use image motion to estimate object 1477 distance. Naturwissenschaften 76, 281–282. https://doi.org/10.1007/BF00368643
- Kirkerud, N., Wehmann, H., Galizia, C., Gustav, D., 2013. APIS—a novel approach for conditioning 1478 1479 honey bees. Front. Behav. Neurosci. 7, 29. https://doi.org/10.3389/fnbeh.2013.00029
- 1480 Kirkerud, N.H., Schlegel, U., Giovanni Galizia, C., 2017. Aversive Learning of Colored Lights in Walking 1481 Honeybees. Front. Behav. Neurosci. 11, 94. https://doi.org/10.3389/fnbeh.2017.00094

- 1482 Kiya, T., Kunieda, T., Kubo, T., 2007. Increased Neural Activity of a Mushroom Body Neuron Subtype
 1483 in the Brains of Forager Honeybees. PLOS ONE 2, e371.
 1484 https://doi.org/10.1371/journal.pone.0000371
- Kócsi, Z., Murray, T., Dahmen, H., Narendra, A., Zeil, J., 2020. The Antarium: A Reconstructed Visual
 Reality Device for Ant Navigation Research. Front. Behav. Neurosci. 14, 599374.
 https://doi.org/10.3389/fnbeh.2020.599374
- Komischke, B., Sandoz, J.-C., Malun, D., Giurfa, M., 2005. Partial unilateral lesions of the mushroom
 bodies affect olfactory learning in honeybees Apis mellifera L. Eur. J. Neurosci. 21, 477–485.
 https://doi.org/10.1111/j.1460-9568.2005.03879.x
- Kramer, E., 1976. The orientation of walking honeybees in odour fields with small concentration
 gradients. Physiol. Entomol. 1, 27–37. https://doi.org/10.1111/j.1365-3032.1976.tb00883.x
- Kuntz, S., Poeck, B., Sokolowski, M.B., Strauss, R., 2012. The visual orientation memory of Drosophila
 requires Foraging (PKG) upstream of Ignorant (RSK2) in ring neurons of the central complex.
 Learn. Mem. 19, 337–340. https://doi.org/10.1101/lm.026369.112
- Kuntz, S., Poeck, B., Strauss, R., 2017. Visual Working Memory Requires Permissive and Instructive
 NO/cGMP Signaling at Presynapses in the Drosophila Central Brain. Curr. Biol. 27, 613–623.
 https://doi.org/10.1016/j.cub.2016.12.056
- Kuwabara, M., 1957. Bildung des bedingten Reflexes von Pavlovs Typus bei der Honigbiene, Apis
 mellifica (Mit 1 Textabbildung) 13, 458–464.
- Laloi, D., Sandoz, J. c., Picard-Nizou, A. I., Marchesi, A., Pouvreau, A., Taséi, J. n., Poppy, G., Phamdelègue, M. h., 1999. Olfactory conditioning of the proboscis extension in bumble bees.
 Entomol. Exp. Appl. 90, 123–129. https://doi.org/10.1046/j.1570-7458.1999.00430.x
- Lamberts, K., Lamberts, K., Shanks, D., 1998. Knowledge Concepts and Categories. Psychology Press,
 London. https://doi.org/10.4324/9780203765418
- Li, L., MaBouDi, H., Egertová, M., Elphick, M.R., Chittka, L., Perry, C.J., 2017. A possible structural
 correlate of learning performance on a colour discrimination task in the brain of the
 bumblebee. Proc. R. Soc. B Biol. Sci. 284, 20171323. https://doi.org/10.1098/rspb.2017.1323
- Liu, G., Seiler, H., Wen, A., Zars, T., Ito, K., Wolf, R., Heisenberg, M., Liu, L., 2006. Distinct memory
 traces for two visual features in the Drosophila brain. Nature 439, 551–556.
 https://doi.org/10.1038/nature04381
- Loesche, F., Reiser, M.B., 2021. An Inexpensive, High-Precision, Modular Spherical Treadmill Setup
 Optimized for Drosophila Experiments. Front. Behav. Neurosci. 15, 138.
 https://doi.org/10.3389/fnbeh.2021.689573
- Lozano, V., Armengaud, C., Gauthier, M., 2001. Memory impairment induced by cholinergic
 antagonists injected into the mushroom bodies of the honeybee. J. Comp. Physiol. A 187,
 249–254. https://doi.org/10.1007/s003590100196
- Luu, T., Cheung, A., Ball, D., Srinivasan, M.V., 2011. Honeybee flight: A novel "streamlining" response.
 J. Exp. Biol. 214, 2215–2225. https://doi.org/10.1242/jeb.050310
- Marchal, P., Villar, M.E., Geng, H., Arrufat, P., Combe, M., Viola, H., Massou, I., Giurfa, M., 2019.
 Inhibitory learning of phototaxis by honeybees in a passive-avoidance task. Learn. Mem. 26, 412–423. https://doi.org/10.1101/lm.050120.119
- Martin, J.P., Guo, P., Mu, L., Harley, C.M., Ritzmann, R.E., 2015. Central-Complex Control of
 Movement in the Freely Walking Cockroach. Curr. Biol. 25, 2795–2803.
 https://doi.org/10.1016/j.cub.2015.09.044
- Masuhr, T., Menzel, R., 1972. Learning Experiments on the Use of Side Specific Information in the
 Olfactory and Visual System in the Honey Bee (Apis mellifica), in: Wehner, R. (Ed.),
 Information Processing in the Visual Systems of Anthropods: Symposium Held at the
 Department of Zoology, University of Zurich, March 6–9, 1972. Springer, Berlin, Heidelberg,
- pp. 315–321. https://doi.org/10.1007/978-3-642-65477-0_45
 Matsumoto, Y., Menzel, R., Sandoz, J.-C., Giurfa, M., 2012. Revisiting olfactory classical conditioning
 of the proboscis extension response in honey bees: A step toward standardized procedures.
 J. Neurosci. Methods 211, 159–167. https://doi.org/10.1016/j.jneumeth.2012.08.018

1534 Mauelshagen, J., 1993. Neural correlates of olfactory learning paradigms in an identified neuron in 1535 the honeybee brain. J. Neurophysiol. 69, 609–625. https://doi.org/10.1152/jn.1993.69.2.609 1536 Menzel, R., 2009. Serial Position Learning in Honeybees. PLoS ONE 4, e4694. 1537 https://doi.org/10.1371/journal.pone.0004694 Menzel, R., 1999. Memory dynamics in the honeybee. J. Comp. Physiol. A 185, 323–340. 1538 1539 https://doi.org/10.1007/s003590050392 1540 Menzel, R., 1974. Spectral Sensitivity of Monopolar Cells in the Bee Lamina. J Comp Physiol A 93, 1541 337-346. https://doi.org/10.1007/BF00606801 1542 Menzel, R., 1967. Untersuchungen zum Erlernen von Spektralfarben durch die Honigbiene (Apis 1543 mellifica). Z. Für Vgl. Physiol. 56, 22–62. https://doi.org/10.1007/BF00333562 Menzel, R., Ventura, D.F., Hertel, H., de Souza, J.M., Greggers, U., 1986. Spectral sensitivity of 1544 1545 photoreceptors in insect compound eyes: Comparison of species and methods. J. Comp. 1546 Physiol. A 158, 165–177. https://doi.org/10.1007/BF01338560 Menzel, Randolf, 1979. Spectral Sensitivity and Color Vision in Invertebrates, in: Autrum, H., Bennett, 1547 1548 M.F., Diehn, B., Hamdorf, K., Heisenberg, M., Järvilehto, M., Kunze, P., Menzel, R., Miller, W.H., Snyder, A.W., Stavenga, D.G., Yoshida, M., Autrum, H. (Eds.), Comparative Physiology 1549 1550 and Evolution of Vision in Invertebrates: A: Invertebrate Photoreceptors, Handbook of 1551 Sensory Physiology. Springer, Berlin, Heidelberg, pp. 503–580. https://doi.org/10.1007/978-1552 3-642-66999-6 9 Meyer, A., Galizia, C.G., 2012. Elemental and configural olfactory coding by antennal lobe neurons of 1553 1554 the honeybee (Apis mellifera). J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol. 1555 198, 159-171. https://doi.org/10.1007/s00359-011-0696-8 1556 Minatohara, K., Akiyoshi, M., Okuno, H., 2016. Role of Immediate-Early Genes in Synaptic Plasticity 1557 and Neuronal Ensembles Underlying the Memory Trace. Front. Mol. Neurosci. 8, 78. 1558 https://doi.org/10.3389/fnmol.2015.00078 1559 Minnich, D.E., 1921. An experimental study of the tarsal chemoreceptors of two nymphalid 1560 butterflies. J. Exp. Zool. 33, 172–203. https://doi.org/10.1002/jez.1400330105 1561 Mizunami, M., Unoki, S., Mori, Y., Hirashima, D., Hatano, A., Matsumoto, Y., 2009. Roles of 1562 octopaminergic and dopaminergic neurons in appetitive and aversive memory recall in an 1563 insect. BMC Biol. 7, 46. https://doi.org/10.1186/1741-7007-7-46 1564 Mobbs, P.G., 1984. Neural networks in the mushroom bodies of the honeybee. J. Insect Physiol. 30, 1565 43-58. https://doi.org/10.1016/0022-1910(84)90107-0 Mobbs, P.G., Young, J.Z., 1982. The brain of the honeybee Apis mellifera. I. The connections and 1566 1567 spatial organization of the mushroom bodies. Philos. Trans. R. Soc. Lond. B Biol. Sci. 298, 1568 309-354. https://doi.org/10.1098/rstb.1982.0086 Moore, R.J.D., Taylor, G.J., Paulk, A.C., Pearson, T., van Swinderen, B., Srinivasan, M.V., 2014. FicTrac: 1569 1570 A visual method for tracking spherical motion and generating fictive animal paths. J. Neurosci. Methods 225, 106–119. https://doi.org/10.1016/j.jneumeth.2014.01.010 1571 1572 Mota, T., Giurfa, M., Sandoz, J.-C., 2011. Color modulates olfactory learning in honeybees by an 1573 occasion-setting mechanism. Learn. Mem. 18, 144–155. https://doi.org/10.1101/lm.2073511 1574 Mota, T., Gronenberg, W., Giurfa, M., Sandoz, J.-C., 2013. Chromatic processing in the anterior optic 1575 tubercle of the honey bee brain. J. Neurosci. Off. J. Soc. Neurosci. 33, 4–16. 1576 https://doi.org/10.1523/JNEUROSCI.1412-12.2013 1577 Mota, T., Kreissl, S., Carrasco Durán, A., Lefer, D., Galizia, G., Giurfa, M., 2016. Synaptic Organization 1578 of Microglomerular Clusters in the Lateral and Medial Bulbs of the Honeybee Brain. Front. 1579 Neuroanat. 10, 103. https://doi.org/10.3389/fnana.2016.00103 1580 Mota, Theo, Roussel, E., Sandoz, J.-C., Giurfa, M., 2011a. Visual conditioning of the sting extension 1581 reflex in harnessed honeybees. J. Exp. Biol. 214, 3577–3587. 1582 https://doi.org/10.1242/jeb.062026 1583 Mota, Theo, Yamagata, N., Giurfa, M., Gronenberg, W., Sandoz, J.-C., 2011b. Neural organization and 1584 visual processing in the anterior optic tubercle of the honeybee brain. J. Neurosci. Off. J. Soc. 1585 Neurosci. 31, 11443–11456. https://doi.org/10.1523/JNEUROSCI.0995-11.2011

- 1586 Mureli, S., Thanigaivelan, I., Schaffer, M.L., Fox, J.L., 2017. Cross-modal influence of mechanosensory 1587 input on gaze responses to visual motion in Drosophila. J. Exp. Biol. 220, 2218–2227. 1588 https://doi.org/10.1242/jeb.146282
- 1589 Neuser, K., Triphan, T., Mronz, M., Poeck, B., Strauss, R., 2008. Analysis of a spatial orientation 1590 memory in Drosophila. Nature 453, 1244–1247. https://doi.org/10.1038/nature07003
- 1591 Núñez, J., Almeida, L., Balderrama, N., Giurfa, M., 1997. Alarm Pheromone Induces Stress Analgesia via an Opioid System in the Honeybee. Physiol. Behav. 63, 75-80. 1592 1593 https://doi.org/10.1016/S0031-9384(97)00391-0
- 1594 Ofstad, T.A., Zuker, C.S., Reiser, M.B., 2011. Visual place learning in Drosophila melanogaster. Nature 1595 474, 204–207. https://doi.org/10.1038/nature10131
- 1596 Paffhausen, B.H., Fuchs, I., Duer, A., Hillmer, I., Dimitriou, I.M., Menzel, R., 2020. Neural Correlates of 1597 Social Behavior in Mushroom Body Extrinsic Neurons of the Honeybee Apis mellifera. Front. 1598 Behav. Neurosci. 14, 62. https://doi.org/10.3389/fnbeh.2020.00062
- 1599 Paffhausen, B.H., Petrasch, J., Wild, B., Meurers, T., Schülke, T., Polster, J., Fuchs, I., Drexler, H., 1600 Kuriatnyk, O., Menzel, R., Landgraf, T., 2021. A Flying Platform to Investigate Neuronal 1601 Correlates of Navigation in the Honey Bee (Apis mellifera). Front. Behav. Neurosci. 15, 1602 690571. https://doi.org/10.3389/fnbeh.2021.690571
- 1603 Pan, Y., Zhou, Y., Guo, C., Gong, H., Gong, Z., Liu, L., 2009. Differential roles of the fan-shaped body and the ellipsoid body in Drosophila visual pattern memory. Learn. Mem. 16, 289–295. 1604 1605 https://doi.org/10.1101/lm.1331809
- 1606 Paulk, A.C., Dacks, A.M., Gronenberg, W., 2009a. Color processing in the medulla of the bumblebee 1607 (Apidae: Bombus impatiens). J. Comp. Neurol. 513, 441–456. 1608 https://doi.org/10.1002/cne.21993
- 1609 Paulk, A.C., Dacks, A.M., Phillips-Portillo, J., Fellous, J.-M., Gronenberg, W., 2009b. Visual Processing 1610 in the Central Bee Brain. J. Neurosci. 29, 9987–9999. 1611
 - https://doi.org/10.1523/JNEUROSCI.1325-09.2009
- 1612 Paulk, A.C., Gronenberg, W., 2008. Higher order visual input to the mushroom bodies in the bee, 1613 Bombus impatiens. Arthropod Struct. Dev. 37, 443–458. 1614 https://doi.org/10.1016/j.asd.2008.03.002
- 1615 Paulk, A.C., Kirszenblat, L., Zhou, Y., van Swinderen, B., 2015. Closed-Loop Behavioral Control 1616 Increases Coherence in the Fly Brain. J. Neurosci. 35, 10304–10315. 1617 https://doi.org/10.1523/JNEUROSCI.0691-15.2015
- Paulk, A.C., Phillips-Portillo, J., Dacks, A.M., Fellous, J.-M., Gronenberg, W., 2008. The Processing of 1618 1619 Color, Motion, and Stimulus Timing Are Anatomically Segregated in the Bumblebee Brain. J. 1620 Neurosci. 28, 6319-6332. https://doi.org/10.1523/JNEUROSCI.1196-08.2008
- 1621 Paulk, A.C., Stacey, J.A., Pearson, T.W.J., Taylor, G.J., Moore, R.J.D., Srinivasan, M.V., Swinderen, B. 1622 van, 2014a. Selective attention in the honeybee optic lobes precedes behavioral choices. 1623 Proc. Natl. Acad. Sci. 111, 5006–5011. https://doi.org/10.1073/pnas.1323297111
- 1624 Paulk, A.C., Stacey, J.A., Pearson, T.W.J., Taylor, G.J., Moore, R.J.D., Srinivasan, M.V., Swinderen, B. 1625 van, 2014b. Selective attention in the honeybee optic lobes precedes behavioral choices. 1626 Proc. Natl. Acad. Sci. 111, 5006–5011. https://doi.org/10.1073/pnas.1323297111
- 1627 Peitsch, D., Fietz, A., Hertel, H., de Souza, J., Ventura, D.F., Menzel, R., 1992. The spectral input 1628 systems of hymenopteran insects and their receptor-based colour vision. J. Comp. Physiol. A 1629 170, 23–40. https://doi.org/10.1007/BF00190398
- 1630 Pfeiffer, K., Homberg, U., 2014. Organization and Functional Roles of the Central Complex in the 1631 Insect Brain. Annu. Rev. Entomol. 59, 165–184. https://doi.org/10.1146/annurev-ento-1632 011613-162031
- Pfeiffer, K., Homberg, U., 2007. Coding of Azimuthal Directions via Time-Compensated Combination 1633 1634 of Celestial Compass Cues. Curr. Biol. 17, 960–965.

- Plath, J.A., Barron, A.B., 2015. Current progress in understanding the functions of the insect central
 complex. Curr. Opin. Insect Sci., Neuroscience * Special Section: Insect conservation 12, 11–
 18. https://doi.org/10.1016/j.cois.2015.08.005
- Plath, J.A., Entler, B.V., Kirkerud, N.H., Schlegel, U., Galizia, C.G., Barron, A.B., 2017. Different Roles
 for Honey Bee Mushroom Bodies and Central Complex in Visual Learning of Colored Lights in
 an Aversive Conditioning Assay. Front. Behav. Neurosci. 11.
 https://doi.org/10.3389/fnbeh.2017.00098
- Poll, M.N.V.D., Zajaczkowski, E.L., Taylor, G.J., Srinivasan, M.V., Swinderen, B. van, 2015. Using an
 abstract geometry in virtual reality to explore choice behaviour: visual flicker preferences in
 honeybees. J. Exp. Biol. 218, 3448–3460. https://doi.org/10.1242/jeb.125138
- 1646 Rapaport, A., 1973. [Instrumental deconditioning of the young rat conditioned to an ambient
 1647 temperature of 40 degrees C with the operational technic of B. F. Skinner]. C. R. Seances Soc.
 1648 Biol. Fil. 167, 900–903.
- 1649Rebenitsch, L., Owen, C., 2016. Review on cybersickness in applications and visual displays. Virtual1650Real. 20, 101–125. https://doi.org/10.1007/s10055-016-0285-9
- 1651 Reiser, M.B., Dickinson, M.H., 2008. A modular display system for insect behavioral neuroscience. J.
 1652 Neurosci. Methods 167, 127–139. https://doi.org/10.1016/j.jneumeth.2007.07.019
- 1653 Ren, Q., Li, H., Wu, Y., Ren, J., Guo, A., 2012. A GABAergic inhibitory neural circuit regulates visual
 1654 reversal learning in Drosophila. J. Neurosci. Off. J. Soc. Neurosci. 32, 11524–11538.
 1655 https://doi.org/10.1523/JNEUROSCI.0827-12.2012
- 1656 Ribi, W.A., 1975. The first optic ganglion of the bee. Cell Tissue Res. 165, 103–111.
 1657 https://doi.org/10.1007/BF00222803
- 1658 Ribi, W.A., Scheel, M., 1981. The second and third optic ganglia of the worker bee: Golgi studies of
 1659 the neuronal elements in the medulla and lobula. Cell Tissue Res. 221, 17–43.
 1660 https://doi.org/10.1007/BF00216567
- 1661 Riveros, A.J., Gronenberg, W., 2009. Olfactory learning and memory in the bumblebee Bombus 1662 occidentalis. Naturwissenschaften 96, 851–856. https://doi.org/10.1007/s00114-009-0532-y
- Roubieu, F.L., Serres, J.R., Colonnier, F., Franceschini, N., Viollet, S., Ruffier, F., 2014. A biomimetic
 vision-based hovercraft accounts for bees' complex behaviour in various corridors. Bioinspir.
 Biomim. 9, 036003. https://doi.org/10.1088/1748-3182/9/3/036003
- 1666 Rusch, C., Alonso San Alberto, D., Riffell, J.A., 2021. Visuo-motor feedback modulates neural activities
 1667 in the medulla of the honeybee, Apis mellifera. J. Neurosci. Off. J. Soc. Neurosci.
 1668 https://doi.org/10.1523/JNEUROSCI.1824-20.2021
- 1669 Rusch, C., Roth, E., Vinauger, C., Riffell, J.A., 2017. Honeybees in a virtual reality environment learn
 1670 unique combinations of colour and shape. J. Exp. Biol. 220, 3478–3487.
 1671 https://doi.org/10.1242/jeb.164731
- 1672 Rybak, J., Menzel, R., 1993. Anatomy of the mushroom bodies in the honey bee brain: the neuronal
 1673 connections of the alpha-lobe. J. Comp. Neurol. 334, 444–465.
 1674 https://doi.org/10.1002/cne.903340309
- Sakura, M., Okada, R., Aonuma, H., 2012. Evidence for instantaneous e-vector detection in the
 honeybee using an associative learning paradigm. Proc. R. Soc. B Biol. Sci. 279, 535–542.
 https://doi.org/10.1098/rspb.2011.0929
- Scheiner, R., Abramson, C.I., Brodschneider, R., Crailsheim, K., Farina, W.M., Fuchs, S., Gruenewald,
 B., Hahshold, S., Karrer, M., Koeniger, G., Koeniger, N., Menzel, R., Mujagic, S., Radspieler, G.,
 Schmickl, T., Schneider, C., Siegel, A.J., Szopek, M., Thenius, R., 2013. Standard methods for
 behavioural studies of Apis mellifera. J. Apic. Res. 52.
- Schultheiss, P., Buatois, A., Avarguès-Weber, A., Giurfa, M., 2017. Using virtual reality to study visual
 performances of honeybees. Curr. Opin. Insect Sci. 24, 43–50.
 https://doi.org/10.1016/j.cois.2017.08.003
- Schwaerzel, M., Monastirioti, M., Scholz, H., Friggi-Grelin, F., Birman, S., Heisenberg, M., 2003.
 Dopamine and octopamine differentiate between aversive and appetitive olfactory
 memories in Drosophila. J. Neurosci. Off. J. Soc. Neurosci. 23, 10495–10502.

- Skorupski, P., MaBouDi, H., Galpayage Dona, H.S., Chittka, L., 2018. Counting insects. Philos. Trans. R.
 Soc. B Biol. Sci. 373, 20160513. https://doi.org/10.1098/rstb.2016.0513
- Sommerlandt, F.M.J., Brockmann, A., Rössler, W., Spaethe, J., 2019. Immediate early genes in social insects: a tool to identify brain regions involved in complex behaviors and molecular processes underlying neuroplasticity. Cell. Mol. Life Sci. 76, 637–651.
 https://doi.org/10.1007/s00018-018-2948-z
- Sommerlandt, F.M.J., Spaethe, J., Rössler, W., Dyer, A.G., 2016. Does Fine Color Discrimination
 Learning in Free-Flying Honeybees Change Mushroom-Body Calyx Neuroarchitecture? PLoS
 ONE 11, e0164386. https://doi.org/10.1371/journal.pone.0164386
- Srinivasan, M.V., 2010. Honey Bees as a Model for Vision, Perception, and Cognition. Annu. Rev.
 Entomol. 55, 267–284. https://doi.org/10.1146/annurev.ento.010908.164537
- Srinivasan, M.V., Lehrer, M., 1988. Spatial acuity of honeybee vision and its spectral properties. J.
 Comp. Physiol. A 162, 159–172. https://doi.org/10.1007/BF00606081
- Srinivasan, M.V., Lehrer, M., 1984. Temporal acuity of honeybee vision: behavioural studies using
 moving stimuli. J. Comp. Physiol. A 155, 297–312. https://doi.org/10.1007/BF00610583
- Srinivasan, M.V., Lehrer, M., Kirchner, W.H., Zhang, S.W., 1991. Range perception through apparent
 image speed in freely flying honeybees. Vis. Neurosci. 6, 519–535.
 https://doi.org/10.1017/S095252380000136X
- Strausfeld, N.J., Sinakevitch, I., Vilinsky, I., 2003. The mushroom bodies of Drosophila melanogaster:
 An immunocytological and golgi study of Kenyon cell organization in the calyces and lobes.
 Microsc. Res. Tech. 62, 151–169. https://doi.org/10.1002/jemt.10368
- Strube-Bloss, M.F., Rössler, W., 2018. Multimodal integration and stimulus categorization in putative
 mushroom body output neurons of the honeybee. R. Soc. Open Sci. 5, 171785.
 https://doi.org/10.1098/rsos.171785
- Takeda, K., 1961. Classical conditioned response in the honey bee. J. Insect Physiol. 6, 168–179.
 https://doi.org/10.1016/0022-1910(61)90060-9
- Taylor, G.J., Luu, T., Ball, D., Srinivasan, M.V., 2013. Vision and air flow combine to streamline flying
 honeybees. Sci. Rep. 3. https://doi.org/10.1038/srep02614
- Taylor, G.J., Paulk, A.C., Pearson, T.W.J., Moore, R.J.D., Stacey, J.A., Ball, D., Swinderen, B. van,
 Srinivasan, M.V., 2015. Insects modify their behaviour depending on the feedback sensor
 used when walking on a trackball in virtual reality. J. Exp. Biol. 218, 3118–3127.
 https://doi.org/10.1242/jeb.125617
- Tedjakumala, S.R., Aimable, M., Giurfa, M., 2013. Pharmacological modulation of aversive
 responsiveness in honey bees. Front. Behav. Neurosci. 7, 221.
 https://doi.org/10.3389/fnbeh.2013.00221
- Tedjakumala, S.R., Giurfa, M., 2013. Rules and mechanisms of punishment learning in honey bees:
 the aversive conditioning of the sting extension response. J. Exp. Biol. 216, 2985–2997.
 https://doi.org/10.1242/jeb.086629
- Tedore, C., Johnsen, S., 2017. Using RGB displays to portray color realistic imagery to animal eyes.
 Curr. Zool. 63, 27–34. https://doi.org/10.1093/cz/zow076
- Triphan, T., Poeck, B., Neuser, K., Strauss, R., 2010. Visual Targeting of Motor Actions in Climbing
 Drosophila. Curr. Biol. 20, 663–668. https://doi.org/10.1016/j.cub.2010.02.055
- Unoki, S., Matsumoto, Y., Mizunami, M., 2005. Participation of octopaminergic reward system and
 dopaminergic punishment system in insect olfactory learning revealed by pharmacological
 study. Eur. J. Neurosci. 22, 1409–1416. https://doi.org/10.1111/j.1460-9568.2005.04318.x
- 1733 Van Swinderen, B., 2012. Competing visual flicker reveals attention-like rivalry in the fly brain. Front.
 1734 Integr. Neurosci. 6. https://doi.org/10.3389/fnint.2012.00096
- 1735 Vergoz, V., Roussel, E., Sandoz, J.-C., Giurfa, M., 2007. Aversive Learning in Honeybees Revealed by
 1736 the Olfactory Conditioning of the Sting Extension Reflex. PLOS ONE 2, e288.
 1737 https://doi.org/10.1371/journal.pone.0000288

67

- 1738 Vogt, K., Schnaitmann, C., Dylla, K.V., Knapek, S., Aso, Y., Rubin, G.M., Tanimoto, H., 2014. Shared
 1739 mushroom body circuits underlie visual and olfactory memories in Drosophila. eLife 3,
 1740 e02395. https://doi.org/10.7554/eLife.02395
- 1741 Von Frisch, K., 1914. Der Farbensinn und Formensinn der Biene. Zool Jb Physiol 1–238.
- 1742 Vorobyev, M., Gumbert, A., Kunze, J., Giurfa, M., Menzel, R., 1997. FLOWERS THROUGH INSECT EYES.
 1743 Isr. J. Plant Sci. 45, 93–101. https://doi.org/10.1080/07929978.1997.10676676
- Wehner, R., 1967. Pattern Recognition in Bees. Nature 215, 1244–1248.
 https://doi.org/10.1038/2151244a0
- Whitlow, J.W., Wagner, A.R., 1972. Negative patterning in classical conditioning: Summation of
 response tendencies to isolable and configural components. Psychon. Sci. 27, 299–301.
 https://doi.org/10.3758/BF03328970
- Yang, E.-C., Lin, H.-C., Hung, Y.-S., 2004. Patterns of chromatic information processing in the lobula of
 the honeybee, Apis mellifera L. J. Insect Physiol. 50, 913–925.
 https://doi.org/10.1016/j.jinsphys.2004.06.010
- Yilmaz, A., Grübel, K., Spaethe, J., Rössler, W., 2019. Distributed plasticity in ant visual pathways
 following colour learning. Proc. Biol. Sci. 286, 20182813.
 https://doi.org/10.1098/rspb.2018.2813
- Zentall, T.R., Galizio, M., Critchfield, T.S., 2002. Categorization, Concept Learning, and Behavior
 Analysis: An Introduction. J. Exp. Anal. Behav. 78, 237–248.
 https://doi.org/10.1901/jeab.2002.78-237
- Zentall, T.R., Wasserman, E.A., Lazareva, O.F., Thompson, R.K.R., Rattermann, M.J., 2008. Concept
 learning in animals. Comp. Cogn. Behav. Rev. 3, 13–45.
 https://doi.org/10.3819/ccbr.2008.30002
- 1761 Zhang, S., 2012. Visually Guided Decision Making in Foraging Honeybees. Front. Neurosci. 6, 88.
 1762 https://doi.org/10.3389/fnins.2012.00088
- Zhang, S., Bock, F., Si, A., Tautz, J., Srinivasan, M.V., 2005. Visual working memory in decision making
 by honey bees. Proc. Natl. Acad. Sci. 102, 5250–5255.
- 1765 https://doi.org/10.1073/pnas.0501440102
- Zhang, S.W., Bartsch, K., Srinivasan, M.V., 1996. Maze Learning by Honeybees. Neurobiol. Learn.
 Mem. 66, 267–282. https://doi.org/10.1006/nlme.1996.0069
- 1768 Zhang, S.W., Lehrer, M., Srinivasan, M.V., 1999. Honeybee Memory: Navigation by Associative
 1769 Grouping and Recall of Visual Stimuli. Neurobiol. Learn. Mem. 72, 180–201.
 1770 https://doi.org/10.1006/nlme.1998.3901
- 1771 Zhang, S.W., Lehrer, M., Srinivasan, M.V., 1998. Eye-specific learning of routes and "signposts" by
 1772 walking honeybees. J. Comp. Physiol. A 182, 747–754.
 1773 https://doi.org/10.1007/s003590050219
- 1774 Zhang, S.W., Srinivasan, M.V., Horridge, G.A., 1992. Pattern recognition in honeybees: local and
 1775 global analysis. Proc. R. Soc. Lond. B Biol. Sci. 248, 55–61.
 1776 https://doi.org/10.1098/rspb.1992.0042
- Zwaka, H., Bartels, R., Grünewald, B., Menzel, R., 2018. Neural Organization of A3 Mushroom Body
 Extrinsic Neurons in the Honeybee Brain. Front. Neuroanat. 12, 57.
 https://doi.org/10.3389/fnana.2018.00057
- Zwaka, H., Bartels, R., Lehfeldt, S., Jusyte, M., Hantke, S., Menzel, S., Gora, J., Alberdi, R., Menzel, R.,
 2019. Learning and Its Neural Correlates in a Virtual Environment for Honeybees. Front.
 Behav. Neurosci. 12, 279. https://doi.org/10.3389/fnbeh.2018.00279
- 1783

Chapter 1

1786 Motion cues from the background influence associative color learning of

1787 honey bees in a virtual-reality scenario



scientific reports

OPEN Motion cues from the background influence associative color learning of honey bees in a virtual-reality scenario

Gregory Lafon¹, Scarlett R. Howard^{1,4}, Benjamin H. Paffhausen¹, Aurore Avarguès-Weber^{1,5} & Martin Giurfa^{1,2,3,553}

1790

We developed a fully 3D virtual environment, in which tethered bees walking stationary can explore a virtual arena and investigate and learn 3D objects. The presence of the third dimension thus created a more complex and immersive VR in which we studied the incidence of motions cues, induced either ventrally by displacements of the treadmill or frontally by virtual movements in the VR itself, on visual discrimination in the VR landscape.

We expected that the addition of motion cues would increase learning performances either by creating a more immersive experience or simply by enhancing attention by increasing the amount of movement on screen. However, we found that frontal background motion cues impaired color discrimination. Ventral motion cues did not affect color discrimination but influenced walking parameters. In this chapter, we present the various effects of motion cues on visual learning and motor behavior, andprovide potential explanations for their negative impact on color discrimination.

1803

Check for updates

1804	Motion cues from the background influence associative
1805	color learning of honey bees in a virtual-reality scenario
1806	
1807	
1808	Gregory Lafon ¹ , Scarlett R. Howard ^{1¥} , Benjamin H. Paffhausen ¹ , Aurore
1809	Avarguès-Weber ^{1*} and Martin Giurfa ^{1, 2, 3*}
1810	
1811	
1812	¹ Research Centre on Animal Cognition, Center for Integrative Biology, CNRS (UMR 5169), University of Toulouse,
1813	118 route de Narbonne, F-31062 Toulouse cedex 09, France
1814	² College of Animal Sciences (College of Bee Science), Fujian Agriculture and Forestry University, Fuzhou 350002,
1815	China
1816	³ Institut Universitaire de France (IUF), Paris, France
1817	
1818 1819	[¥] : <i>Present address:</i> School of Life & Environmental Sciences, Melbourne Burwood Campus, Deakin University, Melbourne, VIC, Australia
1820	
1821	
1822	* Senior authorship shared
1823	
1824	Running Title: Motion Cues in Virtual Reality Learning by Bees
1825	Correspondence to: Martin Giurfa
1826	Research Center on Animal Cognition
1827	Center of Integrative Biology
1828	CNRS - University Paul Sabatier - Toulouse III
1829	4R4 Building - 4th Floor
1830	118 Route de Narbonne
1831	31062 Toulouse cedex 9
1832	FRANCE
1833	
1835 Abstract

1836

Honey bees exhibit remarkable visual learning capacities, which can be studied using virtual 1837 reality (VR) landscapes in laboratory conditions. Existing VR environments for bees are 1838 imperfect as they provide either open-loop conditions or 2D displays. Here we achieved a true 1839 3D environment in which walking bees learned to discriminate a rewarded from a punished 1840 virtual stimulus based on color differences. We included ventral or frontal background cues, 1841 1842 which were also subjected to 3D updating based on the bee movements. We thus studied if and 1843 how the presence of such motion cues affected visual discrimination in our VR landscape. Our results showed that the presence of frontal, and to a lesser extent, of ventral background motion 1844 1845 cues impaired the bees' performance. Whenever these cues were suppressed, color discrimination learning became possible. We analyzed the specific contribution of foreground 1846 1847 and background cues and discussed the role of attentional interference and differences in stimulus salience in the VR environment to account for these results. Overall, we show how 1848 1849 background and target cues may interact at the perceptual level and influence associative learning in bees. In addition, we identify issues that may affect decision-making in VR 1850 landscapes, which require specific control by experimenters. 1851

1852

1853 Keywords

1854

1855 Vision – Visual Learning – Color Discrimination – Optic Flow – Motion cues – Background
 1856 – Honey bees

1858 Introduction

1859

Understanding the spatiotemporal processes that guide decision-making in animals and humans is essential in cognitive research and may be facilitated by virtual reality (VR)^{1,2}, which allows generating of immersive spatial environments in well-controlled laboratory settings. In such environments, experiences are simulated based on changes of perceived landscapes or images, which are updated based on the subject's own movements and decisions^{1,2}.

Insects have pioneered the implementation of VR paradigms aimed at studying 1865 perceptual and cognitive capacities. A predecessor of current VR systems is the flight simulator 1866 conceived for the fruit fly Drosophila melanogaster. In this setup, which was first used to study 1867 how optical properties of compound eyes influence optomotor reactions³, a tethered fly flies 1868 stationary in the middle of a cylindrical arena and experiences surrounding visual stimuli that 1869 1870 can be updated by the fly's movements. Newer versions of this apparatus are still used for numerous studies on visual learning and memory in flies⁴⁻⁷. 'Locomotion compensators' were 1871 1872 also developed to study decision-making by walking insects on two-dimensional surfaces. In silk moths and honey bees, for instance, a 'servosphere' - a form of spherical treadmill that 1873 compensates every locomotive movement of a walking insect – was first used to study olfactory 1874 orientation towards controlled odor stimuli such as pheromone components and odor 1875 gradients^{8,9}. Spherical treadmills have been used to study multiple behaviors in different insect 1876 species. In these setups, the walking movements of the insect under study are constantly 1877 monitored and translated into displacements of surrounding visual cues (closed-loop 1878 conditions). The insect can be either free^{10,11} or immobilized¹²⁻¹⁶ by a tether glued onto its body 1879 surface (typically on the thorax). In both cases, the insect walks stationary on a treadmill whose 1880 movements are recorded by captors placed lateral or ventral to the treadmill. 1881

VR setups are particularly useful for the presentation of visual cues and the study of visual performances. Screens consisting of LED bulb arrays are commonly employed to provide simple forms of visual stimulation (¹⁷⁻¹⁹). In addition, stimuli projected onto screens by highrate video-projectors have also been used on walking arthropods (e.g.^{12-14,16,20}). Furthermore, treadmills holding a tethered animal can also be set in natural visual surroundings to study the influence of landscape features on navigation performances¹⁰.

Owing to their status of classic models for the study of visual cognition²¹⁻²³, the visual performances of honeybees (*Apis mellifera*) have recently started to be studied in VR setups. The main drive to develop these studies was the impossibility to access the neural underpinnings of visual performances in free-flying bees, which have been traditionally used to study basic

properties of visual learning and perception²⁴. Immobilized bees have been traditionally 1892 required for population recordings of neural activity in the bee brain^{25,26}, thus precluding the 1893 possibility of recording active visually-driven behaviors. VR setups in which a tethered animal 1894 makes decision based on visual cues represent a suitable solution to overcome these limitations 1895 as they provide access both to behavioral output and to the nervous system of a behaving bee 1896 with restricted mobility^{27,28}. This perspective is supported by recent developments allowing to 1897 record from specific neurons in the brain of walking bees ^{16,27,29-31}. Yet, the development of VR 1898 environments requires considerable work in order to adapt visual displays to the subjective 1899 perception of an insect and determine optimal parameters for immersive sensations from an 1900 insect's perspective. 1901

Prior work allowed the development of virtual-reality (VR) systems in which a tethered 1902 honey bee walks stationary on a spherical treadmill (a Styrofoam ball floating on an air cushion) 1903 while perceiving a virtual environment displayed by a video projector onto a semicircular 1904 screen^{12-16,27}. In most cases, however, the visual stimulation provided consisted of a 2D virtual 1905 environment in which only translational image movement (left-right) was coupled to the bees' 1906 movements, thus providing an imperfect immersive environment. Despite the absence of depth 1907 1908 components, bees learned both elemental (e.g. discrimination between blue vs. green discs or squares)¹²⁻¹⁴ and non-elemental discriminations (e.g. the negative patterning problem in which 1909 responding to a visual compound, but not to its components, has to be suppressed)¹⁵, thus 1910 1911 showing the suitability of VR for the study of visual learning.

Here we introduce an improved version of our prior VR setup in which a custom-made 1912 1913 software allowed us to create a 3D virtual landscape in which bees move and learn to discriminate visual stimuli. This modification introduced depth perception estimated via the 1914 optic flow generated by the bee's own movements as a new variable, whose influence on the 1915 visual discrimination needs to be considered. In this new scenario, motion cues were not only 1916 1917 derived from the targets themselves, but also from the background presented either 'behind' the vertically displayed targets or ventrally, on the walking surface. We therefore studied if and 1918 how the addition of these motion cues to our VR setup affects learning and discrimination in 1919 tethered bees. 1920

1921

1923 Materials and methods

1924 Study species and collection

Honey bee foragers (Apis mellifera) were obtained from the CRCA apiary located in the campus 1925 1926 of the University Paul Sabatier. Foragers were captured at gravity feeders providing 0.88 M 1927 sucrose solution upon landing and before they began feeding. This step is important as it ensures 1928 that only bees with the appropriate appetitive motivation were brought to the laboratory for the visual learning experiments. Captured bees were enclosed in individual glass vials and then 1929 1930 transferred to small cages housing ten bees in average; where they had access to ad libitum water and 300 µl of 1.5 M sucrose solution. They were then kept overnight in an incubator at 1931 1932 28°C and 80% humidity. On the next day, each bee was cooled on ice for 5 minutes to 1933 anesthetize it and attach it to its tether. Bees were handled under red light, which ensured a dark 1934 environment to the insects.

1935 Tethering procedure

Each bee was tethered by means of a 0.06 g steel needle, 0.5 mm in diameter and 40 mm in 1936 length, which was fixed to the thorax by melted beeswax. The needle was placed within a glass 1937 cannula, 1 mm in diameter, which was held within a black plastic cylinder, 1 cm in diameter 1938 1939 and 55 mm in length, which was fixed on a holding frame placed above the treadmill (Fig. 1940 1A,B). This system allowed the bee to adjust its position in the vertical axis once set on the ball, 1941 but did not allow rotational movements. The holding frame consisted of a vertical black, plastic half frame made of two vertical rectangular supports, 105 mm in length, connected to an upper, 1942 1943 horizontal rectangular support, 120 mm in length. The latter held the black cylinder in the middle (Fig. 1B). After being attached to its tether, each bee was placed on a small (49 mm 1944 1945 diameter) Styrofoam ball for familiarization to a provisory set-up and provided with 5 μ l of 1.5 1946 M sucrose solution. Each bee was held for 3 h in this provisory setup, which was kept in the 1947 dark and without visual stimulations.

1948 Virtual reality set-up

The bee was then moved to the VR setup to be trained and tested in a 3D visual environment. To establish this environment, we used a custom software developed using the Unity engine (version 2018.3.11f1), open-source code available at https://github.com/G-Lafon/BeeVR. The software updated the position of the bee within the VR every 0.017 s.

1953 The VR apparatus consisted of a spherical Styrofoam ball, which acted as a treadmill 1954 onto which a stationary bee walked while perceiving an artificial visual landscape displayed in

front of it on a semi-circular screen (Fig. 1A). The ball was 50 mm in diameter and weighted 1955 1.07 g (Fig. 1B). The ideal weight (M) for spheres holding insects walking on locomotion 1956 compensators was suggested to be¹⁰ $M_{sphere} = 2.5*M_{animal}$, which in case of a honeybee 1957 weighting in average 0.09 g yields a sphere weight of 0.23 g. Despite the fact that our sphere 1958 1959 was about 5 times heavier, the bees used in our experiments walked on it without noticeable problems. The ball was positioned within a 3D-printed, hollow, cylindrical support (cylinder: 1960 50 mm high, 59 mm diameter). The cylinder allowed distributing an upwards air flow of 33 1961 L.min⁻¹ produced by an AquaOxy 2000 aquarium pump, and released through a small hole at 1962 the base of the cylindrical support. The Styrofoam ball floated on the resulting air cushion and 1963 the tethered bee walked on it while remaining stationary. If the bee moves forward, the ball 1964 moved backwards and if it intended to turn to the right or the left, the ball moved to the left or 1965 the right, respectively. The ball was white and unmarked (Fig. 1B,C) except in the experiment 1966 where the influence of the ventral optic flow was tested. In this case, we compared the bees' 1967 performance using a white ball (Fig. 1C) and a ball displaying a black and white checkered 1968 pattern made of 7 mm² squares (Fig. 1D). The movements of the ball, and thus the walking 1969 behavior of the bee (i.e. speed, orientation and location in the virtual environment), were 1970 1971 recorded by two infrared optic-mouse sensors (Logitech M500, 1000 dpi) placed at a distance of 7 mm from the sphere and forming an angle of 90° angle relative to each other (i.e. 45° from 1972 the bee body axis; see Fig. 1A). 1973



Figure 1. Experimental setup for 3D virtual-reality (VR) studies in honey bees. A) Global 1976 1977 view of the VR system. 1: Semicircular projection screen made of tracing paper. 2: Holding frame to place the tethered bee on the treadmill. 3: The treadmill was a Styrofoam ball 1978 positioned within a cylindrical support (not visible) and floating on an air cushion. 4: Infrared 1979 1980 mouse optic sensors allowing to record the displacement of the ball and to reconstruct the bee's trajectory. 5: Air arrival. **B**) The tethering system. 1: Plastic cylinder held by the holding frame; 1981 the cylinder contained a glass cannula into which a steel needle was inserted. 2: The needle was 1982 1983 attached to the thorax of the bee. 3: Its curved end was fixed to the thorax by means of melted bee wax. C, D) Two types of Styrofoam balls used for assessing the importance of the ventral 1984 optic flow. C) No ventral optic flow provided. D) Ventral optic flow provided. E) Color 1985 discrimination learning in the VR setup. The bee had to learn to discriminate a rewarded from 1986 a non-rewarded color cuboid. Cuboids were green and blue. In this case color training and 1987 testing was set in the 'Transparent Condition', i.e. no background was provided and the VR 1988 display contained only the two colored cuboids on an empty dark background. F) Same as in 1989 E) but in this case, the vertical background of the VR arena was covered by a vertical grating 1990 made of black and reddish bars. Depending on its movements, the background gave origin to 1991 three different conditions: the 'Vertical Grating - Optic Flow Condition', in which the grating 1992 was set in closed loop conditions with respect to the bee movements; the 'Vertical Grating - No 1993 Optic Flow Condition', in which the grating was moved in synchrony with the bee's gaze so 1994 that no motion cues could be derived from the background; and the 'Rotating Vertical Grating 1995 1996 Condition', in which the grating was displaced in the anti-clockwise direction across the screen at a constant speed, thus generating a constant optic flow that was independent of the bee's 1997 1998 movements.

The ball was positioned in front of a half-cylindrical vertical screen, 268 mm in diameter 2000 and 200 mm height, which was placed at 9 cm from the bee. The screen was made of semi-2001 transparent tracing paper, which allowed presentation of a 180° visual environment to the bee 2002 (Fig. 1A). The visual environment was projected from behind the screen using a video projector 2003 2004 connected to a laptop (Fig. 1A). The video projector was an Acer K135 (Lamp: LED, Maximum Vertical Sync: 120 Hz, Definition: 1280 x 800, Minimum Vertical Sync: 50 Hz, Brightness: 2005 600 lumens, Maximum Horizontal Sync: 100.10³ Hz, Contrast ratio: 10 000:1, Minimum 2006 Horizontal Sync: 30.10^3 Hz). The lag between the motion of the bee and the update of the visual 2007 surrounding was measured by a high-speed camera at 1000 fps (Canon RX10 mkIII). The VR 2008 2009 display started as usual and the hovering motionless ball was quickly moved by hand. A highspeed video containing the ball, the hand and the VR was shot. The number of frames until the 2010 background illumination changed were counted by two researchers independently. This 2011 2012 procedure yielded a lag value of 18.00 ± 2.53 ms (mean \pm S.E.; n =10).

2013

Experiment 1: choosing the red intensity for the achromatic black-red background gratings

Honey bees can perceive a red target in achromatic terms and can discriminate it from black based on its achromatic L-receptor contrast^{32,33}. In order to present a vertical, red and black striped background against which a color discrimination had to be achieved, we first performed an experiment to choose the intensity of red that was most appropriate for our background grating. We thus determined the spontaneous phototactic responses of bees towards a vertical red cuboid, which varied in intensity. A red intensity that was high enough to be perceived should induce phototactic attraction.

The cuboid had a 5×5 cm base and 1 m height so that it occupied the entire vertical 2023 extent of the screen irrespective of the bee's position (Fig. 2A, left). At the beginning of each 2024 trial, it subtended a horizontal visual angle of 6.5° and was positioned either to the left (-50°) 2025 2026 or the right $(+50^{\circ})$ of the tethered bee. Approaching the cuboid resulted in an expansion of its horizontal extent (1.7°/cm). A choice was recorded when the bee approached the cuboid within 2027 an area of 3 cm surrounding its virtual surface and directly faced its center (Fig. 2A, middle and 2028 right). Three different groups of bees were tested, each one with a different red intensity (see 2029 Fig. S1A): Red 10 (RGB: 26, 0, 0; irradiance: 13 µW/cm²; N = 19), Red 50 (RGB: 128, 0, 0; 2030 irradiance: 140 µW/cm²; N = 19) or Red 100 (RGB: 255, 0, 0; irradiance: 1130 µW/cm²; N = 2031

2032 20). Table 1 summarizes the conditions of this Experiment as well as those of the subsequent2033 experiments.

Within each group, each bee was subjected to four consecutive tests in extinction conditions. During a test, the bee faced the red cuboid on one side, and no stimulus on the alternative side. We recorded whether the bee chose the red cuboid or the equivalent empty space on the other side (to account for possible stimulus choice from random locomotion paths). Each test lasted 60 s and the inter-test interval was 10 s.

2039 Experiment 2: the influence of motion cues from a vertical background on color 2040 discrimination

Having chosen a red intensity for the red and black striped vertical background (Red 100; see 2041 2042 above), we trained bees to discriminate between two vertical colored cuboids, one rewarded and the other not (see below). Both cuboids had the same dimensions of the red cuboid 2043 employed in the previous experiment. One was blue (RGB: 0, 0, 255, with a dominant 2044 wavelength of 446 nm) and the other green (RGB: 0, 51, 0, with a dominant wavelength at 528 2045 2046 nm) (Fig. 1E, Fig. 2B left) (see Fig. S1A). Their intensity, measured at the level of the bee eye, was 161,000 μ W/cm² (blue cuboid) and 24 370 μ W/cm² (green cuboid). These values were 2047 shown to elicit the same level of spontaneous attraction^{12,15}. The cuboids were positioned 2048 respectively at -50° and $+50^{\circ}$ from the bee's body axis at the beginning of each trial. As in the 2049 2050 previous experiment, approaching a cuboid within an area of 3 cm surrounding its virtual surface followed by direct fixation of its center was recorded as a choice (Fig. 2B middle and 2051 2052 right).

The background on which the color cuboids were visible was varied to assess the effect 2053 2054 of background motion cues on visual discrimination learning. Four experimental conditions 2055 were defined (see Table 1). In the 'Transparent Condition' (N=24), no background was provided and the VR display contained only the two cuboids on an empty dark background 2056 2057 (Fig. 1E). The residual light from the empty background had a dominant wavelength of 449 nm and an irradiance of 38 µW/cm². In the 'Vertical Grating - Optic Flow Condition' (N=17), the 2058 walls of the virtual arena were covered by a vertical grating made of black (RGB: 0, 0, 0; 2059 irradiance: 45 µW/cm²; dominant wavelength 628 nm) and red bars (RGB: 255, 0, 0; irradiance: 2060 1130 µW/cm²; dominant wavelength 628 nm), each subtending a visual angle of 6° (Fig. 1F). 2061 Moving forward increased this visual angle by 0.18°/cm. In the 'Vertical Grating - No Optic 2062 2063 Flow Condition' (N=17), the same grating made of black and red bars was used but the VR

software moved it in synchrony with the bee's gaze so that no motion cues could be derived from the background. Finally, in the '*Rotating Vertical Grating Condition*' (N=17), the same black and red grating was displaced in the anti-clockwise direction across the wall at a constant speed (12 m/s), thus generating a constant optic flow that was independent of the bee's movements.

2069

2070 Experiment 3: the influence of motion cues from a ventral background on color 2071 discrimination

In order to test the potential impact of ventral motion cues, we trained bees to discriminate the same two vertical colored cuboids used in the previous experiment under two different conditions. While the vertical frontal background remained the same as in the *Transparent Condition* of Experiment 2 (Fig. 1E), the treadmill texture was varied between two groups of bees: in one case it was a plain white surface (Fig. 1C; N=29) while in the other case, it was a black and white checkered pattern made of 7 mm² squares (Fig. 1D; N=38). While the first condition did not provide ventral optic flow, the second condition provided it (see Table 1).

2079

2080 Training and testing procedure for the Experiments 2 and 3

Bees were trained during 10 trials using a differential conditioning procedure (Fig. 2C) in which one of the cuboids (i.e. one of the two colors, green or blue) was rewarded with 1.5 M sucrose solution (the appetitive conditioned stimulus or CS+) while the other cuboid displaying the alternative color (the aversive conditioned stimulus or CS-) was associated with either 60 mM quinine (Experiment 2)³⁴ or 3 M NaCl solution^{35,36} (Experiment 3). The latter was used to increase the penalty for incorrect choices³⁷.

2087 At the beginning of the experiment, bees were presented with a dark screen for 60 s. During training trials, each bee faced the virtual environment with the two cuboids in front of 2088 it. The bee had to learn to choose the CS+ cuboid by walking towards it and centering it on the 2089 2090 screen. Training was balanced in terms of color contingencies (i.e. blue and green equally 2091 rewarded across bees) based on a random assignment by the VR software. If the bee reached the CS+ within an area of 3 cm in the virtual environment (i.e. the chosen cuboid subtended a 2092 2093 horizontal visual angle of 53°) and centered it in its front, the screen was locked on that image 2094 for 8 s (Fig. 2B). This allowed the delivery of sucrose solution in case of a correct choice, or of quinine or NaCl in case of an incorrect choice. Solutions were delivered for 3 s by the experimenter who sat behind the bee and used a toothpick to this end. The toothpick contacted first the antennae and then the mouthparts while the screen was locked on the visual image fixated by the bee.

Each training trial lasted until the bee chose one of both stimuli or until a maximum of 60 s (no choice). Thus, a single choice (or a no choice) was recorded during each training trial. Trials were separated by an inter-trial interval of 60 s during which the dark screen was presented. The bees that were unable to choose a stimulus in at least 5 trials were excluded from the analysis. From 216 bees trained in the Second Experiment, 75 were kept for analysis (~35%). From 272 bees trained in the Third Experiment, 67 bees were kept for analysis (~25%).

2105 After the last training trial, each bee was subjected to a non-reinforced test (Fig. 2C) that 2106 contrary to training trials had a fixed duration of 60 s. During this test, two variables were recorded: the first choice (as defined above) and the time spent fixating the rewarded and the 2107 non-rewarded stimulus. Both variables have been used in prior works performed in our VR 2108 setup to characterize test performances as they may reveal different aspects of behavioral 2109 performances^{12,13,15}. Fixation time (s) was defined as the time spent by each cuboid at the center 2110 of the screen $(\pm 2.5 \text{ mm})$ where it was brought by the bee's motor actions. We used a ray-casting 2111 2112 approach to determine if the object was there and recorded collisions between a ray following the forward vector of the bee and the center of the object. 2113



2115 Figure 2. Choice and discrimination learning tasks in the VR setup. A) Experiment 1: Quantification of the spontaneous phototactic responses of bees towards a red cuboid against 2116 the absence of an equivalent stimulus in the symmetric position (dashed cuboid). Choice of the 2117 2118 red cuboid was recorded if the bee reached a virtual area of a radius of 3 cm centered on the cuboid and fixed it frontally. B) Experiments 2 and 3: Color discrimination learning with a 2119 green and a blue cuboid. One cuboid was rewarded with sucrose solution and the other punished 2120 with either quinine solution (Experiment 2) or saline solution (Experiment 3) delivered by the 2121 experimenter. A choice was recorded when the bee reached an area of a radius of 3 cm centered 2122 on the cuboid and fixed it frontally. The cuboid image was then frozen during 8 s and the 2123 corresponding reinforcement (US) was delivered. C) Experimental schedule of color learning 2124 experiments (Experiments 2 and 3). Bees were trained along 10 conditioning trials that lasted 2125 a maximum of 1 min and that were spaced by 1 min (intertrial interval). After the end of 2126 2127 conditioning, and following an additional interval of 1 min, bees were tested in extinction conditions with the two colored cuboids during 1 min. 2128

Experiment	Condition Training - Test	Background	N
Experiment 1	Red 10	Frontal Black	19
Choice of Red Intensity	No Training – Test: Red 10 vs. Nothing		
choice of Red Intensity	<i>Red</i> 50	Frontal Black	19
	No Training – Test: Red 50 vs. Nothing		
	Red 100	Frontal Black	20
	No Training – Test: Red 100 vs. Nothing		
Experiment 2	Transparent Condition	Frontal Black	24
Frontal Motion Cues	Training & Test: Blue vs. Green		
	Vertical Grating - Optic Flow Condition	Frontal: Black & Red Vertical Stripes	17
	Training & Test: Blue vs. Green	Closed Loop	
	Vertical Grating - No Optic Flow Condition	Frontal: Black & Red Vertical Stripes	17
	Training & Test: Blue vs. Green	Fixed to the Bee's Gaze	
	Rotating Vertical Grating Condition	Frontal: Black & Red Vertical Stripes	17
	Training & Test: Blue vs. Green	Constantly Rotating	
Experiment 3	No Ventral Optic Flow Condition	Frontal: Black	29
Ventral Motion Cues	Training & Test: Blue vs. Green	Ventral: None (White Treadmill)	
	Ventral Optic Flow Condition	Frontal Black	38
	Training & Test: Blue vs. Green	Ventral: (Black and White Treadmill)	

Table 1: Summary of the experimental conditions provided in Experiments 1 to 3. N: sample
size of each condition.

2132 Statistical analysis

Statistical analyses were performed using R software³⁸. In Experiment 1 (red perception), the 2133 2134 first choice of the bees in each test was categorized according to three mutually exclusive categories: Red Stimulus (Red), No Stimulus (NS: choice of the area symmetric to the stimulus 2135 position) and no choice (NC). Individual choices were translated into a binomial format (0 or 2136 1) within each category. For instance, a bee choosing the red cuboid was recorded as (1, 0, 0)2137 for a choice of the red stimulus, choice of the no stimulus and NC, respectively. In Experiments 2138 2 and 3, the first choice in each trial and test was categorized as choice of the CS+, choice of 2139 the CS- or no choice (NC). Thus, a bee choosing the CS+ was recorded as (1, 0, 0) for choice 2140 of the CS+, choice of the CS- and NC, respectively. Data were bootstrapped to plot the 2141 proportion of bees in each category with their corresponding 95 % confidence interval. 2142 Performances were analysed using generalized mixed linear models (GLMM) with a binomial 2143 error structure-logit-link function (glmer function of R package lme4)³⁹. The independent 2144 variables (fixed factors) were the experimental group (Condition), the trial number (Trial; 2145 Experiments 2 and 3), the choice category (*Choice*) and the color of the CS+ when applicable 2146 (Color: Blue or Green). Bee ID was included as a random factor to account for the repeated-2147 measure design. Several models were run by testing interactions between factors and by 2148 dropping each factor subsequently to select the model with the highest explanatory power (i.e. 2149

the lowest AIC value). P-values for each factor or interaction were obtained by comparing models. The Tukey method was used for multiple comparisons within the selected model; z values are reported for these analyses. For all experiments, the modeling results are reported in Tables S1 to S3 in Supplementary Information. During the tests of Experiments 2 and 3, we also recorded the time spent fixating the test alternatives (CS+ vs. CS-). Time values were compared using a Wilcoxon signed rank test.

For the acquisition trials, we recorded motor variables such as the total distance walked 2156 during a trial, the walking speed, and the tortuosity of the trajectories. Tortuosity was calculated 2157 2158 as the ratio between the total distance walked and the distance between the first and the last point of the trajectory connected by an imaginary straight line. When the ratio was 1, or close 2159 to 1, trajectories were straightforward while higher values corresponded to sinuous trajectories. 2160 In addition, we analyzed the latency to make a choice starting from the beginning of a trial to 2161 2162 the moment in which a choice (either for the CS+ or the CS-) was recorded. NC data were excluded from the latency analysis. The analysis of these continuous variables was done using 2163 2164 a linear mixed model (lmer function) in which the individual identity (Bee ID) was a random factor and the experimental condition (Condition) and trial number (Trial) were fixed factors. 2165

For each experimental condition, we represented the bees' cumulative trajectories (CS+ 2166 2167 choosing and CS- choosing bees) in terms of heat maps, which show the cumulative coordinates occupied by the bees either during the ten training trials or during the non-reinforced test to 2168 which they were subjected. Coordinates were binned into 1 cm². Warmer colors depict locations 2169 more frequently occupied (see color bar). The highest frequency is cut down to 10 % of the 2170 maximum on the color bar. This was done to decrease the excessive occupancy frequency of 2171 the starting point at the expense of other locations, given that it was the same for all bees. While 2172 the side of the rewarded stimulus was randomized, it was placed arbitrarily on the left in the 2173 2174 heat maps.



Figure 3. Experiment 1 - Choosing the red intensity for the achromatic black and red 2176 background. Quantification of the spontaneous phototactic responses of bees towards a red 2177 cuboid (see Fig. 2A). Three different intensities were assayed, each with a different group of 2178 2179 bees: Red 10 (RGB: 26, 0, 0; irradiance: 13 µW/cm²; N = 19), Red 50 (RGB: 128, 0, 0; 2180 irradiance: 140 μ W/cm²; N = 19) and Red 100 (RGB: 255, 0, 0; irradiance: 1130 μ W/cm²; N = 20). For each intensity, the figure represents the pooled performance of four consecutive 2181 extinction tests in which the spontaneous attraction towards the red cuboid ('Red') was 2182 quantified. 'NS' (no stimulus) represents the choice of an equivalent empty area in the VR arena 2183 that was opposite to the red cuboid (see Fig. 2A). NC: no choice. Both the Red-50 and the Red-2184 100 intensities were sufficient to render the red stimulus detectable for honeybees. For 2185 2186 subsequent experiments, the Red 100 intensity was chosen.

2187 **Results**

2188 Experiment 1: choosing the red intensity for the achromatic black and red background

In a first experiment, we determined the spontaneous phototactic responses of bees towards a

red cuboid (dominant wavelength 628 nm) varying in intensity. Using different groups of bees,

- 2191 we tested three different intensities to define the one that would be sufficient to induce
- 2192 phototactic attraction: Red 10 (RGB: 26, 0, 0; irradiance: 13 μW/cm²), Red 50 (RGB: 128, 0,
- 2193 0; irradiance: 140 μ W/cm²) and Red 100 (RGB: 255, 0, 0; irradiance: 1130 μ W/cm²). Each bee
- 2194 was tested along four consecutive tests with the same intensity. The model that best fitted the
- 2195 data included an interaction between the red intensity and the bees' choice (*Choice*Intensity*:

2200 In the Red 10 condition (Fig. 3A; N = 19 bees), the bees did not prefer the red cuboid to the alternative symmetrical area displaying no stimulus (Red 10 vs. NS: 11.8% vs. 6.8% of 2201 2202 choices, z = 1.11, p = 0.27). Most of the bees did not choose in this condition (81.6% of the 2203 cases: NC vs. Red 10: z = 7.56, p < 0.0001; NC vs. NS: z = 7.54, p = <0.0001). By contrast, 2204 both in the Red 50 (Fig. 3B; N = 19) and in the Red 100 condition (Fig. 3C; N = 20), bees preferred the red stimulus to the equivalent area displaying no stimulus (Red 50 vs. NS: 48.7% 2205 2206 vs. 2.6%, z = 4.73, p < 0.0001; Red 100 vs. NS: 52.5% vs. 1.3%, z = 4.34, p < 0.0001). The percentage of bees not choosing remained high and similar to that of bees choosing the red 2207 2208 cuboid (Red 50 vs. NC: z = 0.000, p = 1.00; Red 100 vs. NC: z = 0.58; p = 0.53). For both 2209 intensities, the proportion of non-choosing bees was significantly higher than the choice of the 2210 absence of stimulus (Red 50 vs. NS: 48.7% vs. 48.6%, z = 4.73 p < 0.0001; Red 100 vs. NS: 52.5% vs. 47.5%, z = 4.14 p < 0.0001). These results indicate that both the Red-50 and the Red-2211 100 intensities were sufficient to render the red stimulus detectable for honeybees. We therefore 2212 chose the Red-100 intensity for the red-and-black gratings used in the subsequent experiment 2213 as it was the more salient stimulus from the two that were detectable by the bees. We were 2214 confident that Red 100 would not induce higher phototaxis than Red 50 as no differences in 2215 2216 attraction existed between the cuboids displaying these two lights (compare Fig. 3 B and 3 C).

2217

2218 Experiment 2: the influence of motion cues from a vertical frontal background on color

2219 discrimination

Four different frontal background conditions were used to assess the effect of motion cues from 2220 2221 the background during color discrimination learning. In the 'Transparent Condition' (N = 24bees), the blue and green cuboids were displayed against a uniform dark background. In the 2222 'Vertical Grating - Optic Flow Condition' (N = 17 bees), the cuboids were presented against a 2223 red-and-black vertical grating, which was coupled to the bee's movements (closed-loop 2224 conditions). In the 'Vertical Grating - No Optic Flow Condition' (N = 17 bees), the cuboids 2225 were displayed against the same red-and-black grating but motion cues from the background 2226 2227 were suppressed by keeping it constantly fixed to the bee's gaze. Finally, in the 'Rotating Vertical Grating Condition' (N = 17 bees), the cuboids were shown against the same red-andblack grating, which was rotated counterclockwise around the virtual arena at a constant speed, thus generating a constant optic flow even when the bee did not move.

2231 Discrimination learning during training

Figure 4A-D shows the learning curves of the four groups of bees trained to discriminate the 2232 green from the blue cuboid under different background conditions and the cumulative heat maps 2233 displaying the locations of the bees in their trajectories during the ten acquisition trials. 2234 Learning curves were obtained by recording the percentage of bees choosing correctly the CS+ 2235 2236 or the CS- in their first choice, or not choosing any stimulus (NC) during each trial. The best explanatory model of the acquisition performance included a three-way interaction between the 2237 condition, the trial number and the bees' choice ($\chi^2 = 50.11$, df:15, p < 0.001) but no effect of 2238 the nature of the CS+ was found (blue or green: $\chi^2 = 0.000$, df:1, p = 1). For each background 2239 condition, data were thus represented as a CS+ vs. a CS- discrimination irrespective of color 2240 identity. In the heat maps, the rewarded cuboid is represented on the left side although its side 2241 was randomized along the training sequence. 2242

2243 When no grating was present in the background and the colored cuboids were displayed against a dark homogeneous background ('Transparent Condition'; Fig. 4A), bees learned to 2244 respond more to the CS+ than to the CS-. The interaction between trial number and bee choices 2245 was significant ($\chi^2 = 7.99$, df:2, p = 0.02). In the course of the 10 conditioning trials, the 2246 percentages of bees responding to the CS+ and that of bees responding to the CS- evolved 2247 differently (z = 2.51, p = 0.01), thus showing successful discrimination learning. Moreover, the 2248 2249 dynamic of CS+ responding bees was also significantly different from that of the nonresponding (NC) bees (z = 2.17, p = 0.03) while the difference between the dynamic of the CS-2250 responding bees and the NC bees was not different (z = 0.13, p = 0.9). In the corresponding 2251 cumulative heat map, a clear V shape is visible, indicating that the bees did interact equally 2252 with both sides in the VR and walked towards the cuboids. 2253



Figure 4. Acquisition performances in a color discrimination learning task under four different background conditions. Each panel shows 2255 on the left the acquisition curves in terms of the percentage of bees responding to the CS+ (red), to the CS- (black) or not making any choice (NC; 2256 gray) during the ten conditioning trials. The pink, light gray and gray areas around the curves represent the 95% confidence interval of CS+, CS-2257 choices and NC, respectively. On the right of each panel, a heat map shows the cumulative coordinates occupied by the bees trained under each 2258 background condition during the ten training trials. Coordinates were binned into 1 cm². Warmer colors depict locations more frequently occupied 2259 2260 (see color bar). The highest frequency is cut down to 10 % of the maximum on the color bar. While the side of the rewarded stimulus was randomized along conditioning trials, it was placed arbitrarily on the right in the heat maps. A) In the 'Transparent Condition' (N = 24), the blue 2261 and green cuboids were displayed against a dark background. **B**) In the 'Vertical Grating - No Optic Flow Condition' (N = 17), the cuboids were 2262 displayed against the same red-and-black grating but motion cues from the background were suppressed by keeping it constantly fixed to the bee's 2263 gaze. C) In the 'Vertical Grating –Optic Flow Condition' (N = 17), the cuboids were presented against a red-and-black vertical grating, which was 2264 coupled to the bee's movements (closed-loop conditions). **D**) In the 'Rotating Vertical Grating Condition' (N = 17), the cuboids were shown 2265 2266 against the same red-and-black grating, which was rotated counterclockwise around the virtual arena at a constant speed, thus generating a constant 2267 optic flow even when the bee did not move.

2269 When the red-and-black grating was moved in synchrony with the bee's gaze so that no motion cues could be derived from the background ('Vertical Grating - No Optic Flow 2270 Condition'; Fig. 4B), bees did not modify significantly their stimulus choice along trials. There 2271 was a significant interaction between trial number and bee choices ($\chi^2 = 13.6$, df:2, p = 0.001) 2272 but only because of a difference in the dynamic of the NC bees compared to other two categories 2273 2274 (NC vs. CS+: z = 3.44, p < 0.001; NC vs. CS-: z = 2.60, p < 0.001). Although the CS+ and the CS- curves seem to indicate color discrimination, no differences between the dynamics of the 2275 2276 percentages CS+ and CS- choosing bees could be detected (z = 1.20, p = 0.23), probably because of the high overlap in confidence intervals of these curves. The cumulative heat map 2277 2278 representing the locations of the bees during their training trajectories shows that, as in the previous two conditions, bees walked and interacted equally with both sides in the VR. 2279

In the 'Vertical Grating - Optic Flow Condition' (Fig. 4C), the closed loop conditions 2280 included both the cuboids and the background grating, i.e. the bees' movements translated and 2281 expanded not only the cuboids but also the background grating accordingly. The interaction 2282 between trial number and bee choices was not significant in this case ($\chi^2 = 5.16$, df:2, p = 0.08). 2283 Contrarily to the previous condition, bees were unable to learn the difference between the CS+ 2284 2285 and the CS- as no improvement could be detected along the 10 training trials (z = 0.33, p =0.74). Only the dynamics of the non-responding bees was significantly lower than that of bees 2286 selecting either the CS+ (z = 4.63, p < 0.001) or the CS- (z = 4.33, p < 0.001). The cumulative 2287 heat map representing the locations of the bees during their training trajectories shows that, as 2288 in the previous condition, bees walked towards the cuboids. This result indicates that despite 2289 interacting with the cuboids, bees had their color learning impaired by the addition of motion 2290 2291 cues from the background.

Finally, in the 'Rotating Vertical Grating Condition' (Fig. 4D) in which the black-and-2292 red grating was displaced at a constant speed irrespective of the bee movements and gaze, a 2293 similar pattern than for the 'No Optic Flow Condition' was observed. A significant interaction 2294 between trial number and bee choices was found ($\chi^2 = 11.21$, df:2, p = 0.004). Yet, it was again 2295 due to differences in the dynamic of the percentage of NC bees vs. the percentages of CS+ and 2296 CS- bees (NC vs. CS+: z = 3.11, p = 0.002; NC vs. CS- z = 2.71, p = 0.007). The percentage of 2297 bees choosing the CS+ and that of bees choosing the CS- did not evolve differently (z = 0.44; 2298 p = 0.66). In this condition, the cumulative heat map shows that bees also walked and interacted 2299 equally with the two cuboid sides along trials. 2300

We analyzed if and how motion cues from the background affected the displacement of bees during the training trials in our four background conditions (Fig. 5). To this end, we quantified the distance walked, the walking speed and the tortuosity of the trajectories (ratio between the total distance walked and the straight line connecting the first and the last point of the trajectory). We also measured the choice latency in each trial, i.e. the time required to choose a cuboid within a trial.

The distance walked (Fig. 5A) increased slightly, yet significantly, over trials (*Trial*: γ^2 2308 = 6.86, df:1, p = 0.009) but was not significantly affected by the background condition 2309 (*Condition*: $\chi^2 = 5.34$, df:3, p = 0.15). The walking speed (Fig. 5B) also increased during 2310 successive trials (*Trial*: $\chi^2 = 172.9$, df:1, p < 0.001) and revealed a significant interaction with 2311 the background condition (*Trial*Condition*: $\chi^2 = 19.3$, df:3, p < 0.001), which was introduced 2312 by the Optic Flow Condition. In this case, bees decreased their speed at the end of the training, 2313 so that a significant difference was detected against the other background conditions 2314 (*Trial*Condition: 'Optic Flow'* vs. '*Transparent'*: t = 3.64, p < 0.001, *Optic Flow* vs. *No Optic* 2315 Flow: t = 3.79, p < 0.001 and 'Optic Flow' vs. 'Rotating Grating': t = 3.47, p < 0.001). This 2316 decrease was concomitant with an increase in the proportion of bees not choosing (Fig. 4B) so 2317 that it may reveal a reduction in motivation at the end of training in this background condition. 2318 2319 The tortuosity of the trajectories (Fig. 5C) was neither affected by the succession of trials nor by the background condition (*Trial*: $\chi^2 = 0.17$, df:1, p = 0.68; *Condition*: $\chi^2 = 3.62$, df:3, p = 2320 0.31), thus confirming that the structure of motor patterns was similar across the background 2321 conditions. Finally, the analysis of choice latency (Fig. 5D) showed a significant decrease along 2322 trials (*Trial*: $\chi^2 = 21.85$, df:1, p < 0.001; Fig. 5D), suggesting an improvement in the bee's 2323 2324 capacity to navigate in the VR environment. This evolution was independent of the background displayed (*Condition*: $\chi^2 = 1.67$, df:3, p = 0.65) but a tendency towards larger latencies was 2325 observed for the 'Optic Flow Condition'. 2326



Figure 5. Motor and temporal components of bee trajectories during the acquisition trials. 2329 For each background condition, the evolution of A) the distance walked, B) the walking speed, 2330 **(C)** the tortuosity and **D**) the choice latency during training trials is shown. The tortuosity was 2331 the ratio between the total distance walked and the straight line connecting the first and the last 2332 point of the trajectory during a training trial. *Transparent Condition*' (N = 24), 'Vertical Grating 2333 - Optic Flow Condition' (N = 17), 'Vertical Grating - No Optic Flow Condition' (N = 17), 2334 'Rotating Vertical Grating Condition' (N = 17). The dashed lines above and below the curves 2335 represent the 95% confidence interval. 2336

2328

2338 Test Performance

After the end of training, each bee was subjected to a test in which the green and the blue cuboids were presented in extinction conditions (no reinforcement provided). We recorded the percentage of bees choosing correctly the CS+ or the CS- in their first choice, or not choosing (NC) and the time spent fixating the CS+ and the CS- (Fig. 6).

The rewarded color did not affect the first choice during the test (*Color*: $\chi^2 = 0$, df:1, p 2343 = 1), so that performances could be analyzed irrespective of color identity within each 2344 background condition. Only under the 'Transparent Condition' (Fig. 6A), the difference 2345 between the percentages of CS+ and CS- responding bees was significant (CS+ vs CS-, z =2346 2.33, p = 0.02). The difference between the CS+ responding bees and the NC bees was also 2347 significant (CS+ vs. NC: z = 2.83, p = 0.005). On the contrary, no difference was detected 2348 2349 between the CS- responding bees and the NC bees (CS- vs. NC: z = 0.71, p = 0.48). For the other three background conditions, no significant differences were detected between the 2350

percentage of bees choosing the CS+ or the CS- (Fig. 6B: 'Optic Flow Condition'; CS+ vs CS-2351 , z = 1.41, p = 0.16; Fig. 6C: 'No Optic Flow Condition'; CS+ vs CS-, z = 1.0, p = 0.30; Fig. 2352 6D: 'Rotating Grating Condition'; CS+ vs CS-, z = 0.4, p = 0.7). The comparisons with NC 2353 2354 bees in these three conditions were all non-significant except in the 'No Optic Flow Condition' 2355 where CS+ responding bees and NC bees differed significantly (Fig. 6B: 'Optic Flow *Condition*'; CS+ vs. NC: z = 1.05, p = 0.30; CS- vs. NC: z = 0.38, p = 0.70; Fig. 6C: '*No Optic* 2356 *Flow Condition*'; CS+ vs. NC: z = 2.38, p = 0.02; CS- vs. NC: z = 1.55, p = 0.12; Fig. 6D: 2357 *Rotating Grating Condition*'; CS+ vs. NC: z = -1.09, p = 0.28; CS- vs. NC: z = -0.75, p = 0.45). 2358 2359 Overall, the first-choice data show that a significant discrimination between the CS+ and the CS- occurred in the 'Transparent Condition', i.e. in the total absence of background 2360 2361 information.

The analysis of the fixation time confirmed and extended this conclusion (Fig. 6 E-H). 2362 2363 Again, no discrimination learning was observed for the conditions in which motion cues were available from the background; bees spent the same amount of time fixating the CS+ and the 2364 2365 CS- both in the 'Optic Flow Condition' (Fig. 6F; Wilcoxon U rank Test: V = 69, p = 0.63) and in the 'Rotating Grating Condition' (Fig. 6H; V = 83, p = 0.78). On the contrary, and consistent 2366 with the analysis based on the 1st choice, bees in the 'Transparent Condition' learned the 2367 discrimination between the CS+ and the CS- as they spent more time fixating the rewarded 2368 color than the non-rewarded one (Fig. 6E; V = 203, p = 0.049). Interestingly, a significant 2369 discrimination was also observed for the 'No Optic Flow Condition' (Fig. 6G; V = 128, p = 2370 0.012), a condition for which the 1st choice did not reveal significant differences. This result 2371 indicates that the reduction of motion cues inherent to the 'No Optic Flow Condition' also 2372 favored the occurrence of color learning, in agreement with what was observed for the 2373 2374 'Transparent Condition'.

2375 The heat maps displaying the cumulative locations occupied by the bees' trajectories 2376 during the entire test are shown in the bottom of Fig. 6. In these maps, the CS+ is displayed on 2377 the right by convention. In the 'Transparent Condition' (Fig. 6A), besides choosing 2378 significantly more the correct cuboid upon their first choice and spending more time fixating it, bees consistently walked towards the cuboids and inspected them. This tendency was not visible 2379 2380 in the conditions in which motion cues were available from the background (Fig. 6B: 'Optic 2381 Flow Condition' and Fig. 6D: 'Rotating Grating Condition'), thus showing the impairment of performances induced by these cues. In the 'No Optic Flow Condition' (Fig. 6C), bees walked 2382 towards the cuboids and their choice was slightly biased towards the correct color, in 2383

accordance with the longer fixation time elicited by this color. Overall, these results reveal a
negative influence of motion cues from the vertical background on visual-discrimination
learning under VR conditions.





Figure 6. Test performances (1st choice and fixation time) in a color discrimination 2389 learning task under four different background conditions. Panels A-D refer to the 1st choice 2390 and show the percentage of bees responding to the CS+ (red), to the CS- (black) or not making 2391 any choice (NC; gray) during a retention test performed in extinction conditions after a 10-trial 2392 training. Error bars indicate 95% confidence intervals. *: p < 0.05; **: p < 0.01; ns: non-2393 significant. A) In the 'Transparent Condition' (N = 24), the blue and green cuboids were 2394 displayed against a dark background. **B**) In the 'Vertical Grating - Optic Flow Condition' (N = 2395 2396 17), the cuboids were presented against a red-and-black vertical grating, which was coupled to 2397 the bee's movements (closed-loop conditions). C) In the 'Vertical Grating - No Optic Flow

Condition' (N = 17), the cuboids were displayed against the same red-and-black grating but 2398 motion cues from the background were suppressed by keeping it constantly fixed to the bee's 2399 gaze. **D**) In the '*Rotating Vertical Grating Condition*' (N = 17), the cuboids were shown against 2400 the same red-and-black grating, which was rotated counterclockwise around the virtual arena 2401 at a constant speed, thus generating a constant optic flow even when the bee did not move. 2402 2403 Panels E-H refer to the fixation time, i.e. the time spent fixating either the CS+ or the CSduring the test. Bars represent the mean fixation time. Error bars indicate the standard error f 2404 the mean. *: p < 0.05; ns: non-significant. E) 'Transparent Condition' (N = 24). F) 'Vertical 2405 *Grating - Optic Flow Condition*' (N = 17). **G**) '*Vertical Grating - No Optic Flow Condition*' 2406 (N = 17). **H**) '*Rotating Vertical Grating Condition*' (N = 17). The bottom row shows the heat 2407 map corresponding to each condition. Each heat map shows the cumulative coordinates 2408 occupied by the bees under each background condition during the test. Coordinates were binned 2409 into 1 cm². Warmer colors depict locations more frequently occupied (see color bar). The 2410 highest frequency is cut down to 10 % of the maximum on the color bar. The rewarded stimulus 2411 was placed arbitrarily on the right. 2412

2413 Experiment 3: Influence of ventral optic cues on visual discrimination learning

We assessed the importance of the ventral optic flow by training two different groups of bees 2414 to discriminate the green from the blue cuboid in the previous 'Transparent Condition' in which 2415 color learning was possible. The groups differed in the Styrofoam ball onto which the bees 2416 walked. For one group, the ball was homogenously white (Fig. 1C) so that no ventral motion 2417 cues were available to the walking bees ('No Ventral Optic Flow Condition', N = 29 bees). For 2418 2419 the other group, the ball presented a black-and-white checkered pattern made of 7 mm² squares (Fig. 1D; 'Ventral Optic Flow Condition', N = 38 bees) so that ventral optic flow was available 2420 to the walking bees. 2421

2422 Discrimination learning during training

Learning curves were again obtained by recording the percentage of bees correctly choosing 2423 2424 the CS+ or the CS- in their first choice, or not choosing any stimulus (NC) during each trial. Figure 7A,B shows the learning curves obtained under the two ventral optic flow conditions 2425 2426 and the cumulative heat map showing equal interaction with the two cuboid sides along trials. Yet, in this case the model that provided the best fit to the data included a three-way interaction 2427 between choices, trial number and color (*Color*Trial*Choice*: $\chi^2 = 64.30$, df:7, p < 0.001) but 2428 with no significant effect of the type of ball used (*Condition*: $\chi^2 = 0$, df:1, p = 1). This shows 2429 that the availability of ventral optic flow did not influence the bees' performance when the 2430 variable quantified was the stimulus choice and that, on the contrary, a color effect existed. To 2431 analyze this effect, we pooled acquisition performances irrespective of the ventral optic flow 2432 condition, and represented them in terms of a green vs. blue discrimination (Fig. 7C: blue+ vs. 2433 green-; Fig. 7D: blue- vs. green+). 2434



Figure 7. Acquisition performances in a color discrimination learning task under two 2436 different ventral optic-flow conditions. A) Color discrimination learning with motion cues 2437 available ventrally on the treadmill (N= 38). Left: Acquisition curves in terms of the 2438 percentage of bees responding to the CS+ (red), to the CS- (black) or not making any choice 2439 (NC; gray) during the ten conditioning trials. The pink, light gray and gray areas around the 2440 2441 curves represent the 95% confidence interval of CS+, CS- choices and NC, respectively. Right: Heat map showing the cumulative coordinates occupied by the bees trained under this condition 2442 during the ten training trials. Coordinates were binned into 1 cm². Warmer colors depict 2443 2444 locations more frequently occupied (see color bar). The highest frequency is cut down to 10 % of the maximum on the color bar. While the side of the rewarded stimulus was randomized 2445 along conditioning trials, it was placed arbitrarily on the right in the heat maps. B) Color 2446 discrimination learning in the absence of ventral motion cues on the treadmill (N= 29). 2447 Left: Acquisition curves as in A). Right: Heat map as in A). C) Data pooled for the two optic-2448 2449 flow conditions and segregated according to the situation in which the CS+ color was Blue 2450 while the CS- color was Green. Acquisition curves in terms of the percentage of bees responding to the CS+ (blue), to the CS- (green) or not making any choice (NC; gray) during 2451 the ten conditioning trials. The blue, green and gray areas around the curves represent the 95% 2452 2453 confidence interval of blue+, green- choices and NC, respectively. D) Data pooled for the two optic-flow conditions and segregated according to the situation in which the CS+ color 2454 was green while the CS- color was blue. Acquisition curves in terms of the percentage of bees 2455 responding to the CS+ (green), to the CS- (blue) or not making any choice (NC; gray) during 2456 the ten conditioning trials. The green, blue and gray areas around the curves represent the 95% 2457 confidence interval of green+, blue- choices and NC, respectively. 2458

In both color conditions, the percentages of bees choosing the stimuli varied along trials 2459 (*Choice*Trial*: Blue: $\chi^2 = 48.86$, df:2, p < 0.001; Green: $\chi^2 = 15.47$, df:2, p < 0.001). However, 2460 the dynamic of the percentage of bees choosing the CS+ and that of bees choosing the CS-2461 differed significantly only when blue was the rewarded color. In this case, the percentages of 2462 bees responding to blue (CS+) and to green (CS-) along trials differed significantly (Fig. 7C; 2463 *Choice***Trial*: CS+ vs. CS-: z = 4.88, p < 0.001). In addition, both the percentages of bees 2464 responding to the rewarded blue and to the punished green differed significantly from the non-2465 responding bees along trials (CS+ vs. NC: z = 5.93, p < 0.001; CS- vs. NC: z = 2.62, p = 0.009). 2466 2467 Segregating these data between the blue-rewarded bees that experienced the ventral optic flow condition and those that did not experience ventral optic flow yielded the same result. In both 2468 2469 cases, the dynamic of the percentage of bees choosing the blue+ and that of bees choosing green- differed significantly ('Ventral Optic Flow': CS+ vs. CS-: z = 3.62, p < 0.001; 'No 2470 *Ventral Optic Flow*': CS+ vs. CS-: z = 3.31, p < 0.001). When green was the rewarded color, 2471 no significant differences in the percentages of bees responding to green+ and that of bees 2472 2473 responding to blue – was detected along trials, even if the former tended to be higher than the latter (Fig. 7D; Choice*Trial: CS+ vs. CS-: z = 0.60, p = 0.55). Both percentages were 2474 2475 significantly higher than that of bees not responding to any stimulus (CS+ vs. NC: z = 3.58, p < 0.001; CS- vs. NC: z = 3.23, p = 0.001). The same pattern of responses with respect to bees 2476 responding to green+ and to blue- was found when analyzing separately the two optic-flow 2477 conditions ('Ventral Optic Flow': CS+ vs. CS-: z = 0.23, p = 0.82; 'No Ventral Optic Flow': 2478 CS+vs. CS-: z = 1.16, p = 0.25).2479

2480 Motor and temporal components of bee trajectories during training

We analyzed the motor performance of bees in the two conditions described above to determine 2481 2482 if and how ventral motion cues affected the displacement of bees in the VR setup during the training trials (Fig. 8A-D). The distance walked during the acquisition phase (Fig. 8A) was 2483 affected by the presence of ventral optic flow (*Condition*: $\chi^2 = 7.45$, df:1, p = 0.006). With the 2484 checkered ball, the bees walked less. The walking speed during the acquisition phase (Fig. 8B) 2485 was also significantly slower when ventral optic flow was available (*Condition*: $\chi^2 = 6.03$, df:1, 2486 p = 0.01) although it increased significantly over trials for both conditions (*Trial*: $\chi^2 = 85.20$; df: 2487 2488 1, p < 0.0001). The tortuosity of the walking paths (Fig. 8C) decreased over trials (*Trial*: χ^2 =7.95, df: 1, p = 0.005) but was unaffected by the ventral optic flow (*Condition*: χ^2 = 0.56, 2489 df:1, p = 0.45). Finally, the latency before making a choice (Fig. 8D) was stable over trials even 2490 if an apparent decrease was observed in the first trials (*Trial*: χ^2 =1.97; df: 1, p = 0.16), and was 2491

not influenced by the ventral optic flow (*Condition*: $\chi^2 = 0.19$, df:1, p = 0.66). Overall, the significant variation in distance walked and walking speed detected between the two conditions shows that bees were not insensitive to the presence of ventral motion cues. They perceived them and in consequence walked slower and less.



Figure 8. Motor and temporal components of bee trajectories during the acquisition trials. For each ventral optic-flow condition, the evolution of A) the distance walked, B) the walking speed, C) the tortuosity and D) the choice latency during training trials is shown. The tortuosity was the ratio between the total distance walked and the straight line connecting the first and the last point of the trajectory during a training trial. '*Ventral Optic Flow*' (N = 38), '*No Ventral Optic Flow*' (N = 29). The dashed lines above and below the curves represent the 95% confidence interval.

2504 Test Performance

2496

After the end of training, each bee was subjected to a test in which the green and the blue 2505 cuboids were presented in extinction conditions (no reinforcement provided) in the presence or 2506 absence of ventral optic flow. We recorded the percentage of bees correctly choosing the CS+, 2507 2508 the CS- or not choosing (NC). There was no significant effect of the ventral optic flow on test performances when the variable considered was the choice made by the bees (*Condition*: γ^2 = 2509 0, df:1, p = 1). Thus, the results of both groups of bees were pooled (N = 67) and shown as a 2510 single graph (Fig. 9A). In this case, the color of the CS+ did not affect the performance (*Color*: 2511 $\chi^2 = 0$, df:1, p = 1), thus showing that the color effect detected during training was not consistent. 2512 In the test, bees preferred the CS+ over all conditions (CS+ vs. CS-: z = 2.41, p = 0.02; CS+ vs. 2513 NC: z = 5.03, p < 0.0001; CS- vs. NC: z = 3.16, p = 0.002), thus confirming that they had 2514 learned the color discrimination during acquisition. 2515



Fig. 9. Test performances (1st choice and fixation time) in a color discrimination learning 2517 task under two different ventral optic-flow conditions (with ventral optic flow and without 2518 2519 ventral optic flow). A) 1st choice performed during the test. As there were neither significant differences between the two ventral optic-flow conditions nor between the color conditions 2520 2521 (blue or green rewarded), results were pooled and presented as a single bar diagram (N = 67). 2522 The graph shows the percentage of bees responding to the CS+ (red), to the CS- (black) or not 2523 making any choice (NC; gray) during the retention test. Error bars indicate 95% confidence intervals. *: p < 0.05; **: p < 0.01; ***: p < 0.001. B) & C) Fixation time during the test in 2524 2525 the 'No Ventral Optic Flow Condition' and in the 'Ventral Optic Flow Condition', 2526 respectively. In this case, fixation times were separated according to the experimental condition, as different response patterns were observed with and without ventral optic flow. 2527 2528 The graphs show the mean time (\pm S.E.) spent fixating either the CS+ or the CS- during the retention test. B) In the 'No Ventral Optic Flow Condition', bees fixated significantly longer 2529 the CS+ than the CS-. **: p < 0.01. C) In the 'Ventral Optic Flow Condition', bees fixated 2530 equally the CS+ and the CS-. NS: not significant. D) & E) Heat maps showing the cumulative 2531 coordinates occupied by the bees during the test in the 'No Ventral Optic Flow Condition' 2532 and in the 'Ventral Optic Flow Condition', respectively. The CS+ is shown on the right by 2533 convention. Coordinates were binned into 1 cm². Warmer colors depict locations more 2534 frequently occupied (see color bar). The highest frequency is cut down to 10 % of the maximum 2535 on the color bar. D) In the 'No Ventral Optic Flow Condition', bees clearly aimed at the CS+ 2536 2537 besides choosing it more frequently in their first choice. E) In the 'Ventral Optic Flow *Condition*', bees also aimed at the CS+ but in a less clear way. 2538

The analysis of the fixation time showed a significant difference between the conditions 2540 'Ventral Optic Flow' and 'No Ventral Optic Flow'. Bees fixated significantly longer the CS+ 2541 than the CS- in the absence of ventral optic flow (Fig. 9B; V = 320, p = 0.008) while they 2542 fixated equally both stimuli in the presence of ventral optic flow (Fig. 9C; V = 373; p = 0.75). 2543 2544 The first condition is identical to the 'Transparent Condition' previously studied, in which a white ball was used as a treadmill. The results were, therefore, consistent between the two 2545 experiments: bees preferred the CS+ in their first choice and spent more time fixating it. The 2546 second condition shows that ventral motion cues played a distractive role as only when they 2547 2548 were absent, did the fixation time correlate with the bees' choice.

The heat maps displaying the cumulative locations occupied by the bees' trajectories during the entire test are shown in Figs. 9D, E. In the '*No Ventral Optic Flow*' condition (Fig. 9D), bees consistently walked towards the CS+ cuboid and inspected it during the test besides choosing it more frequently upon their first choice. In the '*Ventral Optic Flow*' condition (Fig. 9E), bees still walked towards the CS+ cuboid but in a less clear way.

2554 **Discussion**

We studied the impact of motion cues provided by the background on visual-discrimination 2555 learning by honey bees in virtual-reality conditions. Bees had to learn the difference between 2556 two virtual color cuboids, one of which was rewarded while the other was punished. We focused 2557 2558 both on motion cues derived from a background placed frontally 'behind' the color stimuli, and from a ventral ground, which was perceived in the ventral visual field while the bee walked on 2559 2560 a Styrofoam ball. In the latter case, the perceived optic flow had no direct relation with the cuboids perceived in the frontal field. The color discrimination task was set under closed loop 2561 conditions so that in the case of the grating displayed frontally, both the cuboids and the 2562 background could vary (translation and expansion) according to the bees' movements. Our 2563 results indicate that in VR conditions, frontal but no ventral motion cues from the background 2564 2565 interfered with the learning of colors. Although ventral motion cues did not affect color 2566 learning, they were well perceived as they affected walking distance and speed and impaired fixation time of the rewarded stimulus during the test. 2567

2569 Optic flow and visual performances in insects

Optic flow is the pattern of apparent motion of objects, surfaces, and edges in a visual scene 2570 caused by the relative motion between an observer and a scene ^{40,41}. It can be seen as a vector 2571 field that gives the retinal slip speed of each contrasting object encountered in the environment 2572 2573 when the observer moves and/or when features in the environment move relative to the 2574 observer⁴². Optic flow processing is crucial for navigation as it allows assessing the distance to 2575 objects encountered. Objects closer to an observer move faster in the retinal field than distant 2576 objects, so that approaching a target induces higher optic flow while moving away from it 2577 decreases it. This information is crucial for moving insects as it allows estimating distances in translational segments⁴³⁻⁴⁶ and avoiding collisions with circumventing obstacles and flying 2578 equidistantly from parallel landmarks. For instance, when flying along narrow corridors, insects 2579 use the magnitude of visual motion experienced in each eye to control their position, height and 2580 speed⁴⁷⁻⁴⁹. 2581

Motion cues can be extracted at the edge of objects through parallax and allow 2582 evaluating the distance of targets with respect to their background based on differences in their 2583 relative retinal speed⁵⁰⁻⁵⁴. Edges are therefore contrasting regions in terms of motion-parallax 2584 cues and are privileged by flying insects in their detection and landing strategies⁵¹. Numerous 2585 experiments have documented this fact in honey bees⁵⁰⁻⁵⁴. An interesting example is provided 2586 by experiments in which bees were trained to solve a discrimination between a plain black disk 2587 and a black ring positioned a few centimeters in front of a white background. The targets 2588 provided a good contrast to the background both in terms of intensity as well as in terms of the 2589 2590 motion cues provided at their edges so that bees had no problems in learning this shape discrimination⁵⁵. However, when bees were trained on the same shapes, yet cut from a textured 2591 2592 paper and placed in front of a similarly textured background, the task was impossible for them⁵⁵. 2593 This result shows that motion cues alone, which existed because the textured targets were 2594 placed in front of the textured background, are not always helpful to appreciate shape 2595 differences between targets. Interestingly, this impossible discrimination became possible after 2596 the bees were primed by pre-training them with the easy discrimination involving plain stimuli against the white background. This improvement shows that attentional mechanisms boosted 2597 2598 by the priming procedure are crucial for achieving target/background segmentation. The role of attentional mechanisms will be discussed below. 2599

Ventral motion cues did not influence the color discrimination performance of bees in VR but affected walking parameters and fixation time

Multiple layers of neurons within visual circuits in the bee brain are devoted to the segregated 2603 processing of motion cues, which are essential to estimate distances traveled in translational 2604 pathways⁵⁶. Ventral optic flow is particularly important for insects flying in open spaces. In 2605 consequence, flying above surfaces providing strong optic flow cues is preferred by bumblebees 2606 over flying above featureless backgrounds⁵⁷. Experiments on bumblebees trained to fly along 2607 textured tunnels showed that in tunnels of 60 and 120 cm width, control of the lateral position 2608 2609 was achieved by balancing the magnitude of translational optic flow experienced in the lateral visual field of each eye; yet, in wider tunnels, bumblebees used translational optic flow 2610 2611 perceived in the ventral visual field to control their lateral position and to steer along straight tracks⁵⁷. Ventral optic flow can be used to keep a constant height above the ground using a 2612 2613 feedback control loop in which a set point value of perceived ventral optic flow is maintained constant by varying the lift, a solution that was shown experimentally in flying bees^{58,59} and 2614 that proved to be efficient when implemented in flying robots that needed to keep a constant fly 2615 height⁶⁰. 2616

These and other findings^{59,61-63} clearly show the importance of ventral motion cues for 2617 the translational displacements of flying insects. Although less information is available for 2618 2619 walking insects, experiments performed on desert ants *Cataglyphis fortis* walking in narrow tunnels showed that both the lateral and the ventral optic flow were dispensable for distance 2620 estimation⁶⁴. In these insects, the use of a 'pedometer' was proposed, i.e. a stride integrator that 2621 accounts for stride number and the respective stride length^{65,66}. Although optic flow can be 2622 computed by these ants, as shown by the case of ants transported by nestmates, which rely on 2623 the optic flow perceived during their transport⁶⁷, the primary mechanism to gauge distances is 2624 based on idiothetic cues. 2625

In our experiments, bees walking on a Styrofoam ball were partially affected by the 2626 presence or absence of ventral optic-flow cues (Figs. 7-9). During the training, these cues did 2627 2628 not affect the learning performance measured in terms of color choice (Fig. 7). Yet, we found 2629 an effect of color, suggesting that discrimination learning was better when blue was the rewarded color. However, this effect disappeared during the test (Fig. 9), as the first choice of 2630 the bees revealed that they preferred significantly the CS+, irrespective of its color. Ventral 2631 motion cues affected the other variable recorded during the test, the time spent fixating the 2632 cuboids (Figs. 9 B,C). When these cues were absent, bees fixated more the CS+, consistently 2633

with their first color choice; however, when ventral motion cues were available, they fixated both the CS+ and the CS- to similar extents, even if they preferred the CS+ in their first color choice. Thus, ventral motion cues interfered with the time spent fixating the CS+ during the test.

2638 The absence of effect of ventral motion cues during the training did not mean that bees did not pay attention to them or that they were unable to perceive the difference between the 2639 2640 two walking surfaces. Fig. 8A,B shows that both the distance walked and the walking speed 2641 decreased significantly when ventral motion cues were available, thus showing that bees 2642 perceived them. Their impact on these motor variables indicates, in addition, that such cues are relevant for estimating walking distances. This conclusion goes against the possibility that in a 2643 2644 walking context, bees, like desert ants, rely on a mechanism for estimating distances different from that employed during flight. In fact, it is difficult to conceive how the relevance of optic 2645 2646 flow could be switched off during walking, given its fundamental role for bee navigation.

Alternatively, our findings may indicate that ventral optic-flow cues play a fundamental 2647 role en route to the goal for distance estimation and completion of an intended translational 2648 2649 vector, but not in the immediate vicinity of the goal, when the insect faces the task of close-up object recognition. In the latter situation, translational ventral optic flow may be irrelevant as 2650 2651 the goal has been reached. Last but not least, it is worth considering that our experiments did not create a ventral optic flow in the virtual arena, i.e. below the targets to be discriminated, but 2652 only on the walking treadmill. Including ventral motion cues in the floor of the virtual arena 2653 could affect the choice of the color cuboids in a way similar to that induced by the frontal 2654 motion cues from the background. 2655

2656

Frontal motion cues from the background interfered with the color discrimination performance of bees in VR

Motion cues perceived frontally at the edges of vertically displayed targets allows segregating them from their respective background based on motion parallax cues. This feature extraction improves therefore object identification and landing on targets. Yet, in our experiments, whenever motion cues from the background were available (*'Vertical Grating - Optic Flow Condition'* and *'Rotating Vertical Grating Condition'*; see Fig. 4), color discrimination of objects located in the virtual foreground was impaired. This result is in contradiction with the hypothesis that animals should behave better in more realistic environments and challenges *a*

priori efforts towards enriching our VR environment with additional cues besides those to be 2666 2667 learned and discriminated. Indeed, learning was only possible in the total absence of a frontal background ('Transparent Condition', Fig. 4), suggesting that background cues interfered with 2668 the learning of foreground objects. Interestingly, in the 'Vertical Grating - No Optic Flow 2669 2670 Condition', optic flow from the background was artificially suppressed and yet learning was not apparent even if a tendency towards a segregation of CS+ and CS- curves was observed 2671 (see Fig. 4B). However, when test performances were analyzed in terms of the time spent 2672 fixating the CS+ and the CS-, a significant difference in favor of the former was found, which 2673 2674 is consistent with a learning effect. The fact that performances were not as clear as in the 'Transparent Condition' suggests that the mere presence of the background may have been 2675 2676 distractive for the bees. Thus, both the motion cues emanating from the background, and its illumination conditions, may have interfered with color learning in the VR arena. 2677

2678 A first explanation for this interference could rely on the role of irradiance cues used to establish the background in our VR environment. The background was projected onto the 2679 2680 semicircular screen of our setup by a videoprojector and therefore provided irradiance cues that could have attracted the bees based on positive phototaxis, thus interfering with color 2681 2682 discrimination. As bees are tested in the dark, a situation inherent to the use of a videoprojector, phototaxis may have indeed influenced the behavior of the bees in our VR setup as shown in 2683 experiments performed in open-loop conditions using the same kind of videoprojector-based 2684 display¹². Admittedly, the green and the blue lights used for training the bees had 22 times and 2685 2686 143 times more irradiance than the red used for the background (Red 100: RGB: 255, 0, 0; irradiance: 1130 μ W/cm²). It could be interesting to determine if a similar interference with 2687 color learning would take place when using the other red light that the bees could see (Red 50: 2688 2689 RGB: 128, 0, 0; irradiance: 140 μ W/cm²; see Fig. 3). For this light, the difference of irradiance between colors and background decreases in one order of magnitude with respect to the Red 2690 2691 100 used in our experiments. In theory, using the Red 50 light should not change the main findings reported because the phototactic attraction exerted by this stimulus was identical to 2692 2693 that induced by the Red 100 light (see Figs. 3 B and C).

Another reason for the negative influence of motion cues emanating from the frontal background could be an excessive salience of these cues with respect to those from the foreground objects that had to be discriminated. In natural conditions, background objects provide motion cues that are less salient than those of foreground objects. Although we attempted to reproduce this situation in our VR environment (the expansion of the cuboids during forward motion was 1.7 °/cm while that of a red bar from the background was 0.18
°/cm), the optic flow generated by the background might still have been too high and detracted
bees from efficiently learning the discrimination task.

Finally, in the 'Rotating Vertical Grating Condition', optomotor responses triggered by 2702 2703 the background rotating regularly in front of the bees may have interfered with the color 2704 learning task. To determine if this was the case, we analyzed the cumulative turning exhibited 2705 by the bees in this condition and in the 'Transparent Condition', where no background was 2706 available. Figure S2 shows that the cumulative turning in the direction of the rotating 2707 background (to the left) was significantly higher in the 'Rotating Vertical Grating Condition', which indicates the presence of optomotor responses. These responses may have interfered with 2708 2709 color learning and may be one of the causes of the impaired performance observed.

2710

2711 Chromatic and achromatic vision in the VR setup

2712 Bees were trained to discriminate two different colors against an achromatic background. Blue 2713 and green colors differing in intensity were used to this end (Fig. S1A), which may have resulted 2714 in bees using differences in intensity rather than chromatic differences to solve the task when 2715 motion cues from the background did not interfere (i.e. in the 'Transparent Condition'; see Fig. 4A). Yet, this possibility is ruled out by the performance of the bees itself. In this condition, no 2716 2717 asymmetries in color learning were observed depending on which color was rewarded. Had the bees been guided by achromatic intensity, then significant learning asymmetries should have 2718 2719 emerged: bees trained to the less intense green should show impaired learning, detracted by the 2720 highly intense blue displayed by the alternative non-rewarded stimulus. On the contrary bees 2721 trained to the highly intense blue should have their performances amplified by the attraction induced by the blue light. This was not the case and no color effect was observed in this 2722 experiment. The situation presented in the 'Transparent Condition' was reproduced in the 2723 experiments studying the effect of the ventral optic flow, when the surface of the treadmill was 2724 plain white (see Fig. 7, 'No Optic Flow Condition'). In this case, a color effect consistent with 2725 2726 the use of intensity was visible during the training, as performance was better when blue was 2727 rewarded than when green was rewarded (see Fig. 7C,D). However, this effect disappeared 2728 during the test (see Fig. 9), showing that it was inconsistent and that even the bees rewarded on the green color learned the task. These results indicate that in the absence of distractive motion 2729 cues from the background, the bees were mainly guided by chromatic cues from the blue and 2730 green colors although we cannot definitely rule out an incidence of stimulus intensity in these 2731

experiments. Note that the same colors were used in previous studies performed in our VR setup
and that no color asymmetry was found, which goes against the use of color intensity by the
bees^{12-16,27}.

An additional issue that requires consideration is the possible interference of the red 2735 2736 light (Red 100) used for the background with the color vision system of the bees involved in the blue-green discrimination. Fig. S1 shows that Red 100 could only be perceived via the L 2737 (Green) receptor type, i.e. via an achromatic visual mechanism involving a single receptor type. 2738 Bees can see red (see Fig. 3), not as a color, but as an achromatic stimulus, perceived in terms 2739 of its intensity by the L receptor^{32,33}. Whether Red 100 affected negatively chromatic 2740 discrimination, interfering with the L-receptor type involved in this discrimination is unknown. 2741 2742 VR experiments with freely flying bumblebees trained to land on a virtual horizontal blue target 2743 located on a projected achromatic checkerboard made of random pink (RGB: 255, 127, 127) 2744 and white (RGB: 255, 255, 255) squares showed that the background had no incidence on the bees' performance⁶¹. In this case, the pink light used could potentially stimulate all receptor 2745 types and thus truly affect the color vision system, contrarily to our red light (RGB: 255, 0, 0). 2746 The fact that this was not the case suggests a minor effect, if any, of the red light in our color 2747 discrimination experiments. 2748

2749

2750 Conclusion

Our results point towards deficits in attentional processes underlying color discrimination 2751 whenever motion cues from the background were frontally available in our VR setup. In the 2752 case of ventral motion cues, no interference of color learning was observed, yet, a distractive 2753 2754 effect on the time spent fixating the stimuli was detected during the test. Attention plays a fundamental role in visual discrimination tasks achieved by bees and other insects⁶⁸⁻⁷¹. 2755 2756 Attention is defined as the "ability to focus our perception on one stimulus (or group of related stimuli), while filtering out other simultaneous stimuli that are less relevant at any moment"⁷². 2757 Several studies focusing on color discrimination by bees have underlined the importance of 2758 attention in this context. In particular, differential conditioning protocols -as the one used in 2759 this work – are said to require more attention than absolute conditioning, the simple training of 2760 a single stimulus⁷³, in particular when the stimuli to be discriminated are similar³⁴. The role of 2761 attention in visual object recognition was studied by training bees to choose a colored target 2762 disc among a variable number of differently colored distractor discs⁷⁴. Accuracy and decision 2763

time were measured as a function of distractor number and color. For all color combinations,
decision time increased and accuracy decreased with increasing distractor number, whereas
performance improved when more targets were present.

From this perspective, highly salient irradiance or motion cues from the background may have interfered with attentional processes required to achieve the color cuboid discrimination. Further experiments may explore strategies to reduce their salience and thus enable their perceptual filtering in our VR landscape.
2772 Acknowledgements

We thank two anonymous reviewers for useful comments and suggestions on previous versions of this article. This work was supported by an ERC Advanced Grant ('Cognibrains') to M.G, who also thanks the Institut Universitaire de France (IUF), the CNRS and the University Paul Sabatier/Fondation Catalyses for support. S.R.H. was supported by a post-doctoral fellowship

2777 of the Fyssen Foundation.

2778

2779 Contributions

2780 M.G, A.A.-W and G.L. designed the experiment. G.L built the setup and wrote the VR software. G.L performed all the behavioral experiments. S.R.H. contributed to the behavioral 2781 2782 experiments. Rodrigue Fouillet, Juliette Montet, Diane Sam Mine and Emma Giordanengo also 2783 assisted with the behavioral experiments. Behavioral experiments were supervised by M.G. and A.A.-W. Heat maps and analyses of displacements within the VR were performed by G.L. 2784 and B.P. Statistical analyses were performed by A.A.-W, G.L. and MG. The manuscript was 2785 written by M.G. who also obtained funding. All authors reviewed and approved the final version 2786 of the manuscript. 2787

2788

2789 Ethics Declarations

- 2790 Competing interests
- 2791 The authors declare no competing interests.

2793 **References**

2794 1 Dombeck, D. A. & Reiser, M. B. Real neuroscience in virtual worlds. Curr Opin Neurobiol 22, 3-2795 10 (2012). 2796 Tarr, M. J. & Warren, W. H. Virtual reality in behavioral neuroscience and beyond. Nat 2 2797 Neurosci 5 Suppl, 1089-1092 (2002). 2798 3 Götz, K. G. Optomotorische Untersuchung des visuellen systems einiger Augenmutanten der 2799 Fruchtfliege Drosophila. Kybernetik 2, 77-92 (1964). 2800 4 Wolf, R. & Heisenberg, M. Visual orientation in motion-blind flies is an operant behaviour. 2801 Nature 323, 154-156 (1986). 2802 5 Wolf, R., Voss, A., Hein, S. & Heisenberg, M. Can a fly ride a bycicle? Phil Trans R Soc Lond B 2803 337, 261-269 (1992). 2804 Brembs, B. & Heisenberg, M. The operant and the classical in conditioned orientation of 6 2805 Drosophila melanogaster at the flight simulator. Learn Mem 7, 104-115 (2000). 7 2806 Brembs, B. & Heisenberg, M. Conditioning with compound stimuli in Drosophila 2807 melanogaster in the flight simulator. J Exp Biol 204, 2849-2859 (2001). 2808 8 Kramer, E. in Olfaction and Taste Vol. V (eds D. Denton & J.D. Coghlan) 329-335 (Academic 2809 Press, 1975). 9 2810 Kramer, E. The orientation of walking honeybees in odour fields with small concentration gradients. Physiol Entomol 1, 27-37 (1976). 2811 2812 10 Dahmen, H., Wahl, V. L., Pfeffer, S. E., Mallot, H. A. & Wittlinger, M. Naturalistic path 2813 integration of Cataglyphis desert ants on an air-cushioned lightweight spherical treadmill. J 2814 Exp Biol 220, 634-644 (2017). 2815 11 Goulard, R., Buehlmann, C., Niven, J. E., Graham, P. & Webb, B. A motion compensation 2816 treadmill for untethered wood ants Formica rufa: evidence for transfer of orientation 2817 memories from free-walking training. J Exp Biol 223, jeb228601 (2020). 2818 12 Buatois, A. et al. Associative visual learning by tethered bees in a controlled visual 2819 environment. Sci Rep 7, 12903 (2017). 2820 13 Buatois, A., Flumian, C., Schultheiss, P., Avargues-Weber, A. & Giurfa, M. Transfer of visual 2821 learning between a virtual and a real environment in honey bees: the role of active vision. 2822 Front Behav Neurosci 12, 139 (2018). 2823 14 Rusch, C., Roth, E., Vinauger, C. & Riffell, J. A. Honeybees in a virtual reality environment 2824 learn unique combinations of colour and shape. J Exp Biol 220, 3478-3487 (2017). 2825 15 Buatois, A., Laroche, L., Lafon, G., Avargues-Weber, A. & Giurfa, M. Higher-order 2826 discrimination learning by honeybees in a virtual environment. Eur J Neurosci 51, 681-694 2827 (2020). 2828 16 Zwaka, H. et al. Learning and its neural correlates in a virtual environment for honeybees. 2829 Front Behav Neurosci 12, 279 (2018). 2830 Seelig, J. D. et al. Two-photon calcium imaging from head-fixed Drosophila during optomotor 17 2831 walking behavior. Nat Meth 8, 184-184 (2011). 2832 18 Seelig, J. D. & Jayaraman, V. Neural dynamics for landmark orientation and angular path 2833 integration. Nature 521, 186-191 (2015). 2834 19 Paulk, A. C. et al. Selective attention in the honeybee optic lobes precedes behavioral 2835 choices. Proc Natl Acad Sci U S A 111, 5006-5011 (2014). 2836 20 Peckmezian, T. & Taylor, P. W. A virtual reality paradigm for the study of visually mediated 2837 behaviour and cognition in spiders. Anim Behav 107, 87-95 (2015). 2838 Avargues-Weber, A., Mota, T. & Giurfa, M. New vistas on honey bee vision. Apidologie 43, 21 244-268 (2012). 2839 2840 22 Avargues-Weber, A., Deisig, N. & Giurfa, M. Visual cognition in social insects. Annu Rev 2841 Entomol 56, 423-443 (2011). 23 2842 Giurfa, M. & Menzel, R. Insect visual perception: complex abilities of simple nervous systems. 2843 Curr Opin Neurobiol 7, 505-513 (1997).

2844 24 Avarguès-Weber, A. & Mota, T. Advances and limitations of visual conditioning protocols in 2845 harnessed bees. J Physiol Paris 110, 107-118 (2016). 2846 25 Mota, T., Yamagata, N., Giurfa, M., Gronenberg, W. & Sandoz, J. C. Neural organization and 2847 visual processing in the anterior optic tubercle of the honeybee brain. J Neurosci 31, 11443-2848 11456 (2011). 2849 26 Mota, T., Gronenberg, W., Giurfa, M. & Sandoz, J. C. Chromatic processing in the anterior 2850 optic tubercle of the honeybee brain. J Neurosci 33, 4-16 (2013). 27 2851 Rusch, C., Alonso San Alberto, D. & Riffell, J. A. Visuo-motor feedback modulates neural 2852 activities in the medulla of the honeybee, Apis mellifera. J Neurosci 41, 3192-3203 (2021). Schultheiss, P., Buatois, A., Avargues-Weber, A. & Giurfa, M. Using virtual reality to study 2853 28 2854 visual performances of honeybees. Curr Opin Insect Sci 24, 43-50 (2017). 2855 29 Jin, N., Paffhausen, B. H., Duer, A. & Menzel, R. Mushroom body extrinsic neurons in walking 2856 bumblebees correlate with behavioral states but not with spatial parameters during 2857 exploratory behavior. Front Behav Neurosci 14, 590999 (2020). 2858 30 Paffhausen, B. H. et al. Neural correlates of social behavior in mushroom body extrinsic 2859 neurons of the honeybee Apis mellifera. Front Behav Neurosci 14, 62 (2020). 2860 31 Duer, A., Paffhausen, B. H. & Menzel, R. High order neural correlates of social behavior in the 2861 honeybee brain. J Neurosci Methods 254, 1-9 (2015). Reisenman, C. E. & Giurfa, M. Chromatic and achromatic stimulus discrimination of long 2862 32 2863 wavelength (red) visual stimuli by the honeybee Apis mellifera. Arthropod Plant Interact 2, 2864 137-146 (2008). 2865 33 Chittka, L. & Waser, N. M. Why red flowers are not invisible to bees. Isr J Plant Sci 45, 169-2866 183 (1997). 2867 34 Avarguès-Weber, A., de Brito Sanchez, M. G., Giurfa, M. & Dyer, A. G. Aversive reinforcement improves visual discrimination learning in free-flying honeybees. PLoS One 5, e15370 (2010). 2868 2869 35 de Brito Sanchez, M. G., Serre, M., Avargues-Weber, A., Dyer, A. G. & Giurfa, M. Learning 2870 context modulates aversive taste strength in honey bees. J Exp Biol 218, 949-959 (2015). 2871 36 Aguiar, J., Roselino, A. C., Sazima, M. & Giurfa, M. Can honey bees discriminate between 2872 floral-fragrance isomers? J Exp Biol 221 (2018). 2873 37 Bestea, L. et al. Peripheral taste detection in honey bees: what do taste receptors respond 2874 to? Eur J Neurosci (2021). 2875 38 R Development Core Team. R: A Language and Environment for Statistical Computing. (The R 2876 Foundation for Statistical Computing, 2016). 2877 39 Bates, D., Mächler, M., Bolker, B. & Walker, S. Fitting linear mixed-effects models using Ime4. 2878 J Stat Softw 67, 1-48 (2015). 2879 40 Gibson, J. J. Visually controlled locomotion and visual orientation in animals. Br J Psychol 49, 2880 182-194 (1958). 2881 41 Lee, D. N. The optic flow field: the foundation of vision. Philos Trans R Soc Lond B Biol Sci 290, 2882 169-179 (1980). 2883 42 Koenderink, J. J. Optic flow. Vision Res 26, 161-179 (1986). 2884 43 Esch, H. E. & Burns, J. E. Honeybees use optic flow to measure the distance of a food source. 2885 Naturwissenschaften 82, 38-40 (1995). 44 2886 Esch, H. E. & Burns, J. E. Distance estimation by foraging honeybees. J Exp Biol 199, 155-162 2887 (1996). 2888 45 Esch, H. E., Zhang, S., Srinivasan, M. V. & Tautz, J. Honeybee dances communicate distances 2889 measured by optic flow. *Nature* **411**, 581-583 (2001). 2890 46 Tautz, J. et al. Honeybee odometry: performance in varying natural terrain. PLoS Biol 2, E211 2891 (2004).2892 47 Srinivasan, M. V., Zhang, S., Altwein, M. & Tautz, J. Honeybee navigation: nature and 2893 calibration of the "odometer". Science 287, 851-853 (2000). 2894 48 Lecoeur, J., Dacke, M., Floreano, D. & Baird, E. The role of optic flow pooling in insect flight 2895 control in cluttered environments. *Sci Rep* **9**, 7707 (2019).

2896	49	Si, A., Srinivasan, M. V. & Zhang, S. Honeybee navigation: properties of the visually driven
2897	50	odometer'. <i>J Exp Biol</i> 206 , 1265-1273 (2003).
2898 2899	50	Lehrer, M., Srinivasan, M. V., Zhang, S. W. & Horridge, G. A. Motion cues provide the bee's visual world with a third dimension. <i>Nature</i> 332 , 356-357 (1988).
2900	51	Lehrer, M. & Srinivasan, M. V. Object detection by honeybees: Why do they land on edges? J
2901		Comp Physiol A 173 , 23-32 (1993).
2902 2903	52	Srinivasan, M. V., Lehrer, M., Kirchner, W. H. & Zhang, S. W. Range perception through apparent image speed in freely flying honeybees. <i>Vis Neurosci</i> 6 , 519-535 (1991).
2904	53	Lehrer, M., Srinivasan, M. V. & Zhang, S. W. Visual edge-detection in the honeybee and Its
2905		chromatic properties. Proc R Soc Lond B Biol Sci 238 , 321-330 (1990).
2906	54	Srinivasan, M. V., Lehrer, M. & Horridge, G. A. Visual figure ground discrimination in the
2907	-	honeybee - the role of motion parallax at boundaries. Proc R Soc Lond B Biol Sci 238 , 331-350
2908		(1990)
2909	55	7hang, S. W. & Srinivasan, M. V. Prior experience enhances nattern discrimination in insect
2910	55	vision Nature 368 330-332 (1994)
2911	56	Paulk A C Phillins-Portillo I Dacks A M Fellous I M & Gronenberg W The processing
2912	50	of color motion and stimulus timing are anatomically segregated in the humblebee brain
2912		Neurosci 28 , 6319-6332 (2008)
2913	57	Linander N. Baird F. & Dacke M. How humblebees use lateral and ventral ontic flow cues
2914	57	for position control in environments of different proximity <i>I Comp Physiol</i> A 203 343-351
2016		(2017)
2017	58	Portalli G. Ruffier F. & Franceschini N. Honeybees change their height to restore their ontic
2917	50	flow / Comp Physiol A 196 , 307-313 (2010)
2910	59	Portelli G. Serres J. R. & Ruffier F. Altitude control in honeybees: joint vision-based learning
2920	55	and guidance. Sci Rep 7, 9231 (2017).
2921	60	Franceschini, N., Ruffier, F. & Serres, J. A Bio-Inspired Flying Robot Sheds Light on Insect
2922		Piloting Abilities. <i>Curr Biol</i> 17 , 329-335 (2007).
2923	61	Frasnelli, E., Hempel de Ibarra, N. & Stewart, F. J. The dominant role of visual motion cues in
2924		bumblebee flight control revealed through virtual reality. <i>Front Physiol</i> 9 , 1038 (2018).
2925	62	Portelli, G., Ruffier, F., Roubieu, F. L. & Franceschini, N. Honeybees' speed depends on dorsal
2926		as well as lateral, ventral and frontal optic flows. PLoS One 6, e19486 (2011).
2927	63	Giurfa, M., Zaccardi, G. & Vorobyev, M. V. How do bees detect coloured targets using
2928		different regions of their compound eyes. J Comp Physiol A 185, 591-600 (1999).
2929	64	Ronacher, B., Gallizzi, K., Wohlgemuth, S. & Wehner, R. Lateral optic flow does not influence
2930		distance estimation in the desert ant <i>Cataglyphis fortis</i> . J Exp Biol 203 , 1113-1121 (2000).
2931	65	Wittlinger, M., Wehner, R. & Wolf, H. The ant odometer: Stepping on stilts and stumps.
2932		Science 312 , 1965-1967 (2006).
2933	66	Wittlinger, M., Wehner, R. & Wolf, H. The desert ant odometer: a stride integrator that
2934		accounts for stride length and walking speed. J Exp Biol 210 , 198-207 (2007).
2935	67	Pfeffer, S. E. & Wittlinger, M. Optic flow odometry operates independently of stride
2936		integration in carried ants. Science 353 , 1155-1157 (2016).
2937	68	Avargues-Weber, A. & Giurfa, M. Cognitive components of color vision in honey bees: how
2938		conditioning variables modulate color learning and discrimination. J Comp Physiol A 200. 449-
2939		461 (2014).
2940	69	Heisenberg, M., Wolf, R. & Brembs, B. Flexibility in a single behavioral variable of <i>Drosophila</i> .
2941		learn Mem 8, 1-10 (2001).
2942	70	Sareen P. Wolf R & Heisenberg M. Attracting the attention of a fly. <i>Proc. Natl. Acad. Sci</i>
2943		<i>U.S.A.</i> 108 , 7230-7235 (2011).
2944	71	van Swinderen, B. Attention in Drosonhila, Int Rev Neurophiol 99 , 51-85 (2011)
2945	72	Miller, S. M., Ngo, T. T. & van Swinderen, B. Attentional switching in humans and flies: rivalry
2946		in large and miniature brains. Front Hum Neurosci 5. 188 (2012).

- 2947 73 Giurfa, M. Conditioning procedure and color discrimination in the honeybee *Apis mellifera*.
 2948 *Naturwissenschaften* **91**, 228-231 (2004).
- 294974Spaethe, J., Tautz, J. & Chittka, L. Do honeybees detect colour targets using serial or parallel2950visual search? J Exp Biol 209, 987-993 (2006).

Chapter 2

2953 Visual learning in a virtual reality environment upregulates immediate early

2954 gene expression in the mushroom bodies of honey bees



communications biology

ARTICLE

https://doi.org/10.1038/s42003-022-03075-8 OPEN

Check for updates

Visual learning in a virtual reality environment upregulates immediate early gene expression in the mushroom bodies of honey bees

Haiyang Geng^{1,2,5}, Gregory Lafon ^{1,5}, Aurore Avarguès-Weber¹, Alexis Buatois^{1,4,6}, Isabelle Massou^{1,6} & Martin Giurfa ^{1,2,3,6⊠}

2957

2958 In the previous chapter, we established a protocol that allowed conditioning tethered bees walking stationary to discriminate 3D colored stimuli. With this tool, we can train bees to solve 2959 a visual task while providing to every insect the same visual experience during conditioning. 2960 We decided to use our new setup to investigate the brain regions and visual pathways 2961 underlying visual learning. To do so, we quantified the expression of immediate early genes 2962 (IEGs) in different brain regions after conditioning. IEGs are considered as useful markers of 2963 2964 neural activity so that we compared their levels of expression between learners and non-2965 learners. Since every bee received the same visual experience in the VR, the main source of variability should come from whether or not an individual learned the task. By focusing on 2966 kakusei, Hr38 and Egr1, three IEGs that have been related to bee foraging and orientation, we 2967 2968 found that, compared to non-learners, learners exhibited Egr1 upregulation in the calyces of the mushroom bodies. This indicates an involvement of mushroom body calices in associative color 2969 learning. and the usefulness of *Egr1* as a marker of neural activity induced by this phenomenon. 2970

2972	
2973	Visual learning in a virtual reality environment upregulates
2974	immediate early gene expression in the mushroom bodies of honey
2975	bees
2976	
2977	Haiyang Geng ^{*1,2} , Gregory Lafon ^{1*} , Aurore Avarguès-Weber ¹ , Alexis
2978	Buatois ^{1,¥,§} , Isabelle Massou ^{1,§} , Martin Giurfa ^{1,2,3§}
2979	
2980 2981	¹ Research Centre on Animal Cognition, Center for Integrative Biology, CNRS, University of Toulouse, 118 route de Narbonne, F-31062 Toulouse cedex 09, France.
2982 2983	² College of Animal Sciences (College of Bee Science), Fujian Agriculture and Forestry University, Fuzhou 350002, China.
2984	³ Institut Universitaire de France, Paris, France (IUF).
2985	
2986	
2987	
2988	* These authors contributed equally
2989	[§] These authors jointly supervised this work
2990	$^{\mathtt{Y}}$ Present address: Institute of Neuroscience and Physiology, Department of Neurochemistry and
2991	Psychiatry, University of Gothenburg, Su Sahlgrenska, 41345 Göteborg, Sweden.
2992	
2993	Corresponding author: Dr. Martin Giurfa
2994	Research Centre on Animal Cognition, CNRS – UPS, 31062 Toulouse cedex 9, France
2995	martin.giurfa@univ-tlse3.fr
2996	

2998 ABSTRACT

2999 Free-flying bees learn efficiently to solve numerous visual tasks. Yet, the neural underpinnings 3000 of this capacity remain unexplored. We used a 3D virtual reality (VR) environment to study visual learning and determine if it leads to changes in immediate early gene (IEG) expression 3001 in specific areas of the bee brain. We focused on *kakusei*, *Hr38* and *Egr1*, three IEGs that have 3002 3003 been related to bee foraging and orientation, and compared their relative expression in the calyces of the mushroom bodies, the optic lobes and the rest of the brain after color 3004 discrimination learning. Bees learned to discriminate virtual stimuli displaying different colors 3005 3006 and retained the information learned. Successful learners exhibited Egr1 upregulation only in the calvces of the mushroom bodies, thus uncovering a privileged involvement of these brain 3007 regions in associative color learning and the usefulness of *Egr1* as a marker of neural activity 3008 3009 induced by this phenomenon.

3010

3011

3012 Keywords: Vision – Visual Learning – Virtual Reality – Honey Bee Brain – Immediate Early
 3013 Genes – *Kakusei – Hr38 – Egr1 –* Mushroom Bodies

3015 INTRODUCTION

Invertebrate models of learning and memory have proved to be extremely influential to 3016 determine where and when such experience-dependent plasticity occurs in the nervous system¹⁻ 3017 ⁶. One of these models is the domestic honey bee *Apis mellifera*, which has been intensively 3018 investigated for its visual and olfactory learning capacities^{5,7,8}. Yet, the knowledge gained on 3019 the mechanisms of these abilities is disparate. While an extensive body of research has 3020 accumulated on the neural bases of olfactory learning and memory in bees⁹, practically nothing 3021 is known about the neural and molecular underpinnings of their visual learning and memory^{10,11}. 3022 This asymmetry is due to the fact that olfactory learning protocols use harnessed bees that learn 3023 3024 to extend their proboscis to an odorant that has been forward-paired with sucrose water, while 3025 visual learning protocols use free-flying bees trained to choose a visual target where they collect sucrose reward^{5,10}. Whilst the harnessing situation of olfactory-learning protocols facilitates the 3026 use of invasive techniques to record neural activity, the use of bees that commute freely between 3027 3028 the hive and the experimental site precludes an equivalent access to visual neural circuits.

Virtual-reality (VR) environments constitute a valuable tool to overcome this limitation. 3029 In such environments, tethered bees walking stationary on a treadmill are exposed to a 3030 controlled visual environment that allows studying decision making based on visual cues¹²⁻¹⁷. 3031 3032 Under these conditions, bees learn and memorize simple and higher-order visual discrimination 3033 problems, which enables coupling the study of this visual learning with mechanistic analyses of brain activity^{16,17}. VR setups may differ according to the degree of variation introduced by 3034 3035 the bee movement into the visual environment. In closed-loop conditions, this variation is 3036 contingent with the movements of a tethered bee, thus creating a more immersive environment. In prior works, we introduced a 2D VR environment in which a tethered bee could displace 3037 laterally (from left to right and vice versa) a color stimulus on a frontal screen according to its 3038 association with sucrose reward of absence of reward^{12,14,18}. Here we moved towards a more 3039

realistic 3D VR environment which allowed, in addition, for stimulus expansions and retractions depending on forward or backward movements, respectively. In this arena, bees may therefore learn to discriminate colors but can also explore in a less restricted way the virtual world proposed to them.

3044 One way to detect brain regions and pathways activated in this scenario is the quantification of immediate early genes (IEGs) in neural tissues¹⁹. IEGs are transcribed 3045 transiently and rapidly in response to specific stimulations inducing neural activity without de 3046 *novo* protein synthesis²⁰. In mammals, IEGs such as *c-fos*, *zif268* and *Arc* are regularly used as 3047 markers of neural activity during learning, memory and other forms of cellular plasticity such 3048 as long-term potentiation²¹⁻²³. In insects, the use of IEGs as neural markers is less expanded as 3049 3050 the number of candidate genes serving this goal is still reduced and the reliable detection of their expression is sometimes difficult²⁴. Three of the IEGs reported for the honey bee are 3051 3052 interesting as they have been related to a foraging context in which learning plays a fundamental role. The first one, termed kakusei (which means 'awakening' in Japanese) is a nuclear non-3053 3054 coding RNA transiently and strongly induced in the brain of European workers by seizures that can be induced by awakening them from anesthesia²⁵. It is also activated after the experience 3055 of dancing in the hive following a foraging flight and in pollen foragers so that it seems related 3056 to the neural excitation resulting from foraging activities²⁶. This IEG is activated within a 3057 3058 subtype of Kenvon cells, the constitutive neurons of the mushroom bodies, which are a higherorder center in the insect brain²⁷. A second IEG is the hormone receptor 38 gene (Hr38), which 3059 is a transcription factor conserved among insects and other species including humans²⁸, and 3060 which has been indirectly related to learning and memory in honey bees and other insects^{29,30}. 3061 *Hr38* is also upregulated by foraging experiences in honey bees²⁹ and bumblebees³⁰ and by 3062 orientation activities upon hive displacement³¹. The third gene is the early growth response 3063 gene-1 (*Egr1*), whose expression is induced in the brain of honey bees and bumble bees upon 3064

foraging^{29,30} and orientation flights³², and which seems to be controlled by circadian timing of foraging³³. None of these IEGs has been studied so far in the context of associative learning and memory formation in the honey bee.

We thus focused on these IEGs to characterize neural activation induced by visual 3068 learning in the brain of bees under 3D VR conditions. Bees had to learn to discriminate a 3069 rewarded color from a punished color³⁴⁻³⁷ and should retain this information in a short-term 3070 retention test. Our goal was to determine if successful learning and retention activate 3071 specifically certain regions in the brain, in particular the mushroom bodies, whose importance 3072 for olfactory learning and memory has been repeatedly stressed^{5,38}, yet with a dramatic lack of 3073 3074 equivalent evidence in the visual domain. Our results show that successful learners exhibited *Egr1* upregulation only in the calyces of the mushroom bodies, thus uncovering a privileged 3075 involvement of these brain regions in associative color learning. 3076

3077 **RESULTS**

3078 Color learning under 3D VR conditions

Honey bee foragers were captured at an artificial feeder to which they were previously trained 3079 and brought to the laboratory where a tether was glued on their thorax. (Fig. 1A,B). They could 3080 be then attached to a holder that allowed adjusting their position on a treadmill, a polystyrene 3081 3082 ball floating on a constant airflow produced by an air pump (see Methods for details). The VR 3083 setup consisted of this treadmill placed in front of a semi-cylindrical semi-transparent screen made of tracing paper (Fig. 1A). The movements of the walking bee on the treadmill were 3084 recorded by two infrared optic-mouse sensors placed on the ball support perpendicular to each 3085 3086 other.



3087

Figure 1. Experimental setup and 3D environment. A) Global view of the VR system. 1: 3088 3089 Semicircular projection screen made of tracing paper. 2: Holding frame to place the tethered 3090 bee on the treadmill. 3: The treadmill was a Styrofoam ball positioned within a cylindrical support (not visible) floating on an air cushion. 4: Infrared mouse optic sensors allowing to 3091 3092 record the displacement of the ball and to reconstruct the bee's trajectory. 5: Air arrival. The video projector displating images on the screen from behind can be seen on top of the image. 3093 **B)** The tethering system. 1: Plastic cylinder held by the holding frame; the cylinder contained 3094 a glass cannula into which a steel needle was inserted. 2: The needle was attached to the thorax 3095 of the bee. 3: Its curved end was fixed to the thorax by means of melted bee wax. C) Color 3096 3097 discrimination learning in the VR setup. The bee had to learn to discriminate two vertical 3098 cuboids based on their different color and their association with reward and punishment. Cuboids were green and blue on a dark background. Color intensities were adjusted to avoid 3099 phototactic biases independent of learning. 3100

3101

Bees were trained to discriminate a green from a blue vertical cuboid against a black 3102 background during ten conditioning trials (Fig. 1C; see Supplementary Fig. 1 for color 3103 characteristics). Training consisted in pairing one of the cuboids (CS+) with a rewarding 1 M 3104 sucrose solution and the other (CS-) with an aversive 3M NaCl solution^{39,40} (Fig. 2). Bees 3105 performed equally irrespective of the color trained (z = -0.97, p = 0.33). They were subdivided 3106 3107 according to their test performance to distinguish those which showed successful discrimination (i.e. choice of the CS+; "learners") from those which did not ("non-learners"). This distinction 3108 allowed subsequent brain gene analyses according to learning success. Bees that were unable 3109 to choose a stimulus in at least 5 trials were excluded from the analysis. Acquisition was 3110 significant for *learners* (n=17) during conditioning trials (Fig. 3A; CS*Trial effect: χ^2 =33.68, 3111

df:2, p<0.0001), confirming the occurrence of learning. Indeed, the percentages of bees 3112 responding to the CS+ and to the CS- differed significantly along trials (CS+ vs. CS-: CS*Trial; 3113 3114 z=-5.46, p<0.0001). Significant differences were also found when comparing the percentages of non-responding bees against the CS+ responding bees and against the CS- responding bees 3115 3116 (NC vs. CS+: CS*Trial; z=8.14, p<0.0001; NC vs. CS-: CS*Trial; z=4.59, p<0.0001). Non*learners* (n=18) did also show a significant interaction (Fig. 3B; CS*Trial effect: χ^2 =7.66, df:2, 3117 p=0.02), but this was introduced by the percentage of non-responding bees. These bees differed 3118 3119 significantly along trials both from the bees responding to the CS+ (NC vs. CS+: CS*Trial; z=6.10, p<0.0001) and from the bees responding to the CS- (NC vs. CS-: CS*Trial; z=6.07, 3120 p<0.0001). On the contrary, the percentages of bees responding to the CS+ and to the CS- did 3121 not vary along trials (CS+ vs. CS-: CS*Trial; z=-0.07, p=1), consistently with the absence of 3122 3123 learning.





Figure 2. Choice criterion and conditioning protocol for color discrimination learning. A) 3125 **Choice criterion.** Left: A bee facing the two virtual cuboids. Center: A bee approaching a target 3126 cuboid; the cuboid has not yet been centered by the bee (gray area). Right: A bee having 3127 centered the target cuboid (gray area). A choice was recorded when the bee reached an area of 3128 a radius of 3 cm centered on the cuboid and fixed it frontally. The cuboid image was then frozen 3129 3130 during 8 s and the corresponding reinforcement (US) was delivered. B) Conditioning protocol. 3131 Bees were trained along 10 conditioning trials that lasted a maximum of 1 min and that were spaced by 1 min (intertrial interval). After the end of conditioning, and following an additional 3132 interval of 1 min, bees were tested in extinction conditions with the two colored cuboids during 3133 1 min. 3134

3135

We next asked if differences between learners and non-learners could be due to differences in motor components. To answer this question, we analyzed for each conditioning trial the total distance walked, the walking speed, and the tortuosity of the trajectories.

Tortuosity was calculated as the ratio between the total distance walked and the distance 3139 between the first and the last point of the trajectory connected by an imaginary straight line. 3140 When the ratio was 1, or close to 1, trajectories were straightforward while higher values 3141 corresponded to sinuous trajectories. The distance travelled (Fig. 4A) did neither vary along 3142 trials (*Trial:* χ^2 =0.24, df:1, p=0.62) nor between *learners* and *non-learners* (*Condition:* χ^2 =1.10, 3143 df:1, p=0.30; Condition*Trial: χ^2 =0.71, df:1, p=0.40). Tortuosity (Fig. 4B) varied along trials 3144 (*Trial:* χ^2 =14.53, df:1, p<0.001) but not between *learners* and *non-learners* (*Condition*: 3145 χ^2 =0.08, df:1, p=0.80; Condition*Trial: χ^2 =0.42, df:1, p=0.52). Finally, the walking speed (Fig. 3146 4C) increased significantly along trials (*Trial:* χ^2 =30.49, df:1, p<0.0001) but did not vary 3147 between *learners* and *non-learners* (*Condition:* χ^2 =1.43, df:1, p=0.23); in this case, however, 3148 the interaction between Trial and Condition was significant (χ^2 =4.68, df:1, p<0.05). This 3149 suggests that *learners* were slower than *non-learners*, which is reminiscent of a speed-accuracy 3150 trade off reported in numerous experiments in bees⁴¹⁻⁴³. 3151



Figure 3. Discrimination learning in the VR setup. A) Acquisition performance of *learners* 3153 3154 (i.e. percentage of bees that chose the CS+ in the non-reinforced test; n=17). The red, black and grey curves show the percentages of bees choosing the CS+, the CS- or not making a choice 3155 (NC), respectively. Bees learned the discrimination between CS+ and CS-. B) Acquisition 3156 performance of non-learners (i.e. percentage of bees that chose the CS- or did not make a 3157 3158 choice in the non-reinforced test; n=18). These bees did not learn to discriminate the CS+ from the CS-. C) Test performances of learners. Percentage of bees choosing in their first choice 3159 the CS+ (FC CS+), the CS- (FC CS-) or not making a choice (NC). Per definition, learners 3160 chose the CS+ in this test. Different letters on top of bars indicate significant differences 3161 (GLMM; p<0.05). D). Test performances of non-learners. Percentage of bees choosing in 3162 3163 their first choice the CS+ (FC CS+), the CS- (FC CS-) or not making a choice (NC). Per definition, non-learners did not choose the CS+. Different letters on top of bars indicate 3164 3165 significant differences (GLMM; p<0.05). In all panels, error bars indicate the 95% confidence 3166 interval.

Finally, in the non-reinforced test, per definition *learners* (n=17; Fig. 3C) chose correctly the CS+ (100% of the bees) while *non-learners* (n=18; Fig. 3D) did either chose the CS- (72.22%) or did not perform any choice (27.78%). We thus focused on differences between *learners* and *non-learners* in the subsequent IEG analyses to uncover possible changes in neural activity induced by learning.



3173

3174

Figure 4. Motor components of learners (n=17) and non-learners (n = 18) in the VR setup 3175 during conditioning. A) Distance travelled (cm) during each conditioning trial. B) Tortuosity 3176 3177 of the trajectories (see text for explanation) during each conditioning trial. C) Walking speed (cm/s) during each conditioning trial. The dashed lines above and below the curves represent 3178 the 95% confidence interval. Comparisons between curves refer to the significance of the 3179 interaction between the factors Trial (1 to 10) and Condition (learners vs. non-learners). All 3180 comparisons referring to Condition alone were non-significant. LMM; *: p<0.05; NS: non-3181 3182 significant.

Type of gene	Target	Primer sequence 5' ≽3'	Amplicon length (bp)	E (%)	R ²
Target genes	Kakusei	CTACAACGTCCTCTTCGATT (forward) CCTACCTTGGTATTGCAGTT (reverse)	149	96.4	0.991
	Hr38	TGAGATCACCTGGTTGAAAG (forward) CGTAGCAGGATCAATTTCCA (reverse)	118	106	0.995
	Egrl	GAGAAACCGTTCTGCTGTGA (forward) GCTCTGAGGGTGATTTCTCG (reverse)	138	109	0.991
Reference genes	Efl	AAGAGCATCAAGAGCGGAGA (forward) CACTC TTAATGACGCCCACA (reverse)	148	106	0.993
	Actin	TGCCAACACTGTCCTTTCTG (forward) AGAATTGACCCACCAATCCA (reverse)	156	110	0.995

Table 1. Primer sequences used to quantify RNA expression of genes of interest and reference genes by RT-qPCR. Amplicon length (bp), efficiency (E, %) and the coefficient of correlation obtained for the standard curve (\mathbb{R}^2) are also shown. *Hr38*: Hormone receptor 38 gene; *Egr1*: Early growth response gene-1; *Ef1a*: Elongation factor 1 α gene.

3188

3189 IEG analyses in the honey bee brain following color learning under 3D VR conditions

We aimed at determining if visual learning in VR induces post learning transcriptional changes, 3190 which might participate in amplifying neural activity reflecting associative color learning. To 3191 3192 this end, we performed RT-qPCR in individual brains of *learners* and *non-learners*, which were 3193 collected 1h after the retention test and placed in liquid nitrogen until brain dissection. We analyzed relative expression levels of kakusei, Hr38 and Egr1 (see Table 1) in three main brain 3194 regions⁴⁴ (Fig. 5A): the optical lobes (OL), the upper part of the mushroom bodies (i.e. the 3195 mushroom-body calyces or MB Ca) and the remaining central brain (CB), which included 3196 mainly the central complex, the subesophageal zone and the peduncula and lobes (α and β 3197 lobes) of the mushroom bodies. Two reference genes were used for the normalization, $Efl \alpha$ 3198 (E=106%) and Actin (E=110%), which proved to be the best choice for the normalization (see 3199 3200 Table 1). The Cq values of these reference genes for the different conditions of this experiment

are shown in Supplementary Fig. 2. Stability was granted for both genes and experimental groups (*learners* and *non-learners*) for the MB and the CB. In the case of the OL, *Ef1* α varied significantly between groups. Thus, normalization used the product of the two reference genes for MB and CB while only actin could be used to normalize OL data. No cross-comparisons between brain regions or genes were performed.

Figure 5 B-D shows the relative normalized expression of *kakusei* for the three brain 3206 3207 regions considered in the case of learners and non-learners. No significant variations of relative expression were found between these two groups for the three regions considered (two-sample 3208 3209 t test; Fig. 5B, OL: t₂₉=0.83, p=0.42; Fig. 5C, MB: t₂₉=1.09, p=0.29; Fig. 5D, CB: t₂₉=1.04, p=0.31). Thus, *kakusei* was unable to reveal learning-induced variations in neural activity under 3210 our experimental conditions. The normalized expression of Hr38 (Fig. 5 E-G) was also 3211 insufficient to uncover learning related differences between learners and non-learners (Fig. 5E, 3212 OL: t₂₉=0.37, p=0.72; Fig. 5F, MB: t₂₉=0.99, p=0.33; Fig. 5G, CB: t₂₉=0.44, p=0.67). However, 3213 3214 a significant upregulation of Egr1 expression was found in the mushroom bodies of learners when compared to *non-learners* (Fig. 5I, t₂₉=2.40, p=0.02). Differences in *Egr1* expression 3215 between *learners* and *non-learners* were neither found in the OL (Fig. 5H, t₂₉=1.48, p=0.15) 3216 3217 nor in the CB (Fig. 5J, t₂₉=0.17, p=0.86), thus showing that learning-dependent variation in IEG 3218 expression was circumscribed to the calyces of the mushroom bodies and that Egrl was more sensitive than both Hr38 and kakusei to detect changes in neural activity induced by associative 3219 3220 learning.







Figure 5. *Egr1*, but neither *kakusei* nor *Hr38*, shows significant variation of relative expression in the mushroom bodies following visual associative learning in a 3D VR environment. A) Honey bee brain with sections used for quantifying IEG expression.

Yellow labels indicate the brain regions used for the analysis: MB: mushroom body; CB: central 3226 brain; OL: optic lobes. The dashed lines indicate the sections performed. Ca: calvx of the 3227 mushroom body; li: lip; co: collar; \Box and \Box : \Box and \Box lobes of the mushroom body; CC: 3228 3229 central complex; AL: antennal lobe; SEZ: subesophagic zone; OL: optic lobe; Me: medulla; lo: lobula. (B-D) Relative normalized expression of kakusei, of Hr38 (E-G) and of Egr1 (H-J) 3230 3231 in three main regions of the bee brain, the optic lobes (B, E, H), the calyces of the 3232 mushroom bodies (C, F, I) and the central brain (D, G, J). The expression of each IEG was normalized to the expression of two genes of reference (Actin and Ef1 \Box) in the case of the MB 3233 and the CB, and of Actin alone in the case of the OL (see Supplementary Figure 2). The range 3234 of ordinates was varied between target genes to facilitate appreciation of data scatter. IEG 3235 3236 expression was analyzed in individual brains of bees belonging to two categories: learners (conditioned bees that responded correctly and chose the CS+ in their first choice during the 3237 non-reinforced test; n=17) and non-learners (conditioned bees that did not choose the CS+ in 3238 their first choice during the non-reinforced test; n=14). The range of ordinates was varied 3239 3240 between target genes to facilitate appreciation of data scatter. Box plots show the mean value in red. Error bars define the 10th and 90th percentiles. Red boxes indicate cases in which 3241 significant variations were detected. Different letters on top of box plots indicate significate 3242 differences (two-sample t test; p < 0.05). 3243

3244

3245 **DISCUSSION**

Our work shows that visual discrimination learning under virtual-reality conditions leads to an 3246 3247 enhancement of IEG expression in the case of Egr1 in the calyces of the mushroom bodies in successful honey bee learners. Learning success did not correlate with differences in distance 3248 travelled or tortuosity of trajectories, i.e. with differences in exploratory drive (Fig. 4), but was 3249 3250 correlated with differences in walking speed as *learners* tended to be slower than *non-learners*. Although strictly speaking the two categories did not differ with respect to this parameter, the 3251 significant interaction between Trial and Condition suggests a speed-accuracy trade off in 3252 which individuals taking more time to decide can improve the accuracy of their decisions⁴¹⁻⁴³. 3253 Differences in Egr1 expression were thus related to learning success and not to differences in 3254 3255 exploratory components. For the other two IEGs analyzed, kakusei and Hr38, no learningdependent changes could be detected in the different brain regions considered, even if prior 3256 reports indicated similar levels of expression for the three IEGs in the brain of bees engaged in 3257

foraging^{29,30,33,45} and orienting around the hive²⁹⁻³¹. Our work demonstrates therefore that this similarity does not necessarily reflect a relationship with associative learning and memory as only *Egr1* acted as *bona fide* marker of learning success in the bee brain under our experimental conditions and revealed the implication of the calyces of the mushroom bodies in associative visual learning and memory in honey bees.

3263

3264 Differential expression of IEGs in the honey bee brain as related to visual learning

3265 *Kakusei* did not vary in the brain regions considered, under the experimental conditions defined 3266 in our work. This IEG does not have orthologous genes in other taxa and its role in honey bees 3267 is unclear. It is induced by seizures following anesthesia^{25,27,45,46} and thermal stimulation⁴⁶, but 3268 also by foraging and reorientation activity following hive displacement^{25,31,45}. These 3269 experiences increase *kakusei* expression in the mushroom bodies²⁵ but also in the optic 3270 lobes^{25,27,45} and the dorsal lobe²⁷. Our results suggest that its enhanced expression in foragers 3271 or in orienting bees is not necessarily related to learning occurring in these contexts.

Differential expression of *kakusei* with respect to an inducing treatment (typically, an induced seizure) starts around 15 min post treatment^{25,31,46} but continues during longer periods which may go beyond 60 min⁴⁶. Thus, the waiting time of 60 min between test and brain freezing in our experiments was appropriate to detect changes in *kakusei* as a result of associative visual learning. However, as other temporal analyses of *kakusei* expression reported a decay in expression beyond 30 min²⁵, the possibility that our sampling period was too long to capture changes in *kakusei* expression cannot be excluded.

This concern does not apply to Hr38 and Egr1, for which temporal expression analyses showed a systematical increase at the time chosen for our experiments³⁰. As in the case of *kakusei*, no learning-related changes were detected in Hr38 expression across the brain regions considered. This hormone receptor gene has been indirectly related to learning and memory in honey bees and other insects^{29,30} and is also upregulated by foraging experiences in honey bees²⁹ and bumblebees³⁰ and by orientation activities upon hive displacement³¹. Despite its involvement in these activities, it did not reveal learning-dependent changes in neural activity in the experimental context defined by our setup and training protocol.

Only Egr1 reported a significant variation in the mushroom body calvees of *learners* in 3287 relation to non-learners (Fig. 5). As for the two other IEGs, the expression of this early growth 3288 response gene is enhanced in the brain of honey bees and bumblebees upon foraging^{29,30} and 3289 orientation flights³². Yet, in this case, Egrl was sensitive enough to report differences in neural 3290 3291 activity related to learning success in our experimental conditions. Learners and non-learners were identical in their experience and handling all along the experiment and they only differed 3292 in learning success. Thus, differences in Egrl expression demonstrate that associative color 3293 learning is accompanied by increased neural activity in the calyces of the mushroom bodies. 3294

3295

3296 The role of mushroom bodies for visual learning and memory

3297 Although the crucial role of mushroom bodies for the acquisition, storage and retrieval of olfactory memories has been extensively documented in bees^{7,38,47} and other insect species^{2,3,48}, 3298 3299 less is known about their implication in visual learning and memory. In the honey bee, the fact that visual learning was mainly studied using free-flying bees trained to choose visual targets 3300 precluded its study at the cellular level¹³. The neural circuits for color processing are known in 3301 the bee brain⁴⁹⁻⁵² but evidence about plasticity-dependent changes in these circuits remains 3302 scarce. Such changes could occur at multiple stages, as is the case in olfactory circuits mediating 3303 olfactory learning⁹. Upstream the mushroom bodies, inner-layer lobula and inner medulla 3304 neurons project to both the mushroom bodies and the lateral protocerebrum ^{49,50,53} and exhibit 3305 color sensitivity, color opponency and temporally complex patterns including adaptation and 3306 entrainment ^{49,53,54}. These patterns are important for color coding and discrimination and could 3307 be subjected to experience-dependent changes in activity⁵⁵. 3308

The implication of mushroom bodies in visual learning and memory in the bee is 3309 expected given the parallels between visual and olfactory inputs at the level of the calyces. 3310 Whilst afferent projection neurons convey olfactory information to a subdivision of the calyces, 3311 the lip⁵⁶, afferent neurons from the lobula and the medulla, which are part of the optic lobes, 3312 convey visual information to other calvx subdivisions, the collar and the basal ring 50,57 . In spite 3313 3314 of this similarity, studies addressing the role of mushroom bodies in honey bee visual learning and memory remain rare. The recent development of protocols for the study of *aversive* visual 3315 3316 learning (association between a color light and an electric shock delivered to walking bees enclosed in a box compartment)^{44,58} has showed the possible implication of mushroom bodies 3317 3318 in this form of learning. In a pharmacological study, in which one half of a chamber was illuminated with one color and paired with shock while the other half was illuminated with a 3319 different color not paired with shock, bees learned to escape the shock-paired light and spent 3320 more time in the safe light after a few trials⁵⁹. When ventral lobe neurons of the mushroom 3321 bodies were silenced by procaine injection, bees were no longer able to associate one light with 3322 3323 shock. By contrast, silencing one collar region of the mushroom body calyx did not alter behavior in comparison with that of controls⁵⁹. The latter result does not exclude a role for the 3324 calyces in visual learning, as blocking one of four collar regions may not have a significant 3325 impact on learning. In a different study, bees were trained to inhibit their spontaneous 3326 phototaxis by pairing the attracting light with an electric shock⁴⁴. In this case, learning induced 3327 an increase in the dopaminergic receptor gene Amdop 1 in the calyces of the mushroom bodies, 3328 consistently with the role of dopaminergic signaling for electric-shock representation in the bee 3329 brain^{60,61}. 3330

In the fruit fly, the study of the role of mushroom bodies for visual learning and memory has yielded contradictory results. Flies suspended within a flight simulator learn to fly towards unpunished visual landmarks to avoid heat punishment delivered to their thorax; mushroom

body deficits do not affect learning so that these structures were considered dispensable for 3334 visual learning and memory⁶². Similarly, learning to discriminate colors in a cylindrical 3335 container made of a blue-lit and a yellow-lit compartment, one of which was associated with 3336 aversive shaking, was not affected in mushroom body mutants⁶³. Visual place learning by flies 3337 walking within a cylindrical arena displaying landmarks can also take place in the absence of 3338 functional mushroom bodies but requires the central complex⁶⁴. Yet, the dispensability of 3339 mushroom bodies for visual learning and memory in fruit flies has been questioned by 3340 experiments in which appetitive and aversive color learning and discrimination were studied in 3341 an arena in which blue and green colors were presented from below. Walking flies learned both 3342 3343 the appetitive (based on pairing one color with sugar) and the aversive discrimination (based on pairing one color with electric shock) but failed if mushroom body function was blocked 3344 using neurogenetic tools⁶⁵. Thus, the role of mushroom bodies for visual learning and memory 3345 3346 in fruit flies may be both task- and learning specific. In addition, the dominance of olfactory inputs to the mushroom bodies may overshadow their role for visual learning in Drosophila. 3347

3348

3349 IEG expression within the mushroom bodies in relation to visual learning

Kenyon cells are the constitutive neurons of the mushroom bodies. Their somata are located 3350 3351 both within the mushroom-body calyces and adjacent to them. Thus, our brain sectioning (see Fig. 4A) collected them massively. Detecting IEG activation in the mushroom bodies upon 3352 visual learning may be particularly difficult as learning-dependent changes in neural activity 3353 may be subtle due to the characteristic sparse neural activity observed at the level of the calyces. 3354 This reduced activity, which has been revealed in studies on olfactory coding⁶⁶⁻⁶⁸ and odor-3355 related learning⁶⁹, can also be a hallmark of visual processing and visual learning. Sparse neural 3356 3357 coding of odorants is in part due to GABAergic inhibition by feedback extrinsic mushroombody neurons acting on Kenyon cells^{70,71}, the constitutive neurons of the mushroom bodies. 3358

These GABAergic neurons, present both in bees and flies^{70,72,73}, suppress Kenyon cell activity 3359 to maintain sparse, neural coding, and may render difficult detecting variations of IEG 3360 expression in the calyces. Yet, we were able to find differences that were dependent on the 3361 experience of the animals analyzed. Such differences might vary according to the difficulty of 3362 the learning problem considered. For instance, higher GABAergic input is required in the 3363 calyces to solve non-linear discriminations, in which subjects have to inhibit response 3364 summation to the simultaneous presentation of stimuli A and B, which are rewarded when 3365 presented alone but non-rewarded when presented together. Bees that learn to solve this 3366 discrimination in the olfactory domain require inhibitory GABAergic feedback in the calvces 3367 to this end⁴⁷. Such a requirement could translate into a different form of IEG expression in this 3368 brain region as a consequence of a more complex discrimination learning. 3369

Recent work on gene expression in the Kenyon-cells of honey bees revealed the 3370 3371 existence of various cell subtypes/populations with unique gene expression profiles and cell body morphology⁷⁴. Among these populations, small Kenyon cells (sKC)⁷⁵, formerly called 3372 inner Kenyon cells⁷⁶, are found in the central, inner core of the MB calyces and express 3373 3374 preferentially three genes, EcR, E74 and Hr38, the latter being higher in the brain of foragers than in nurses⁷⁴. Unfortunately, no information on *Egr1* was reported in this analysis. Yet, 3375 another study that did not distinguished between Kenvon-cell subtypes reported that the 3376 expression of Egrlis enriched in Kenvon cells compared to the rest of the brain³² and that this 3377 enrichment might be related to learning and memory given its association with the orientation 3378 flights of bees³² and with foraging activities^{29,30,77}. However, the sensory cues and behavioral 3379 3380 programs participating in both foraging and orientation are multiple so that it is difficult to sustain such a claim in the absence of a controlled learning experiment. For instance, Egrl is 3381 also upregulated in the brain of honey bees upon seizure induction⁷⁸, with no relation to foraging 3382

or orientation. Only specific experiments like the one performed in this work can reveal whetherincreases in this and other IEGs reflect neural activity induced by associative learning.

Consistently with the notion that sKCs may be particularly relevant for learning and 3385 memory formation, phosphorylated (activated) cAMP-response element binding protein 3386 (pCREB) is enriched in these sKCs in the honey bee⁷⁹. CREB is a nuclear protein that modulates 3387 the transcription of genes required for the cellular events underlying long-term memory (LTM) 3388 formation in both invertebrates and vertebrates⁸⁰⁻⁸³ and its activation leads also to the expression 3389 3390 of IEGs. It is thus possible that the increased expression of *Egr1* induced by visual learning and memory formation is localized within sKCs, and that this increase results from CREB 3391 3392 activation. In our experiments, the reinforced tests were done shortly after the last conditioning trial and only one hour elapsed since the end of the test and the collection of brains for IEG 3393 analysis (a time necessary for the expression of the IEGs selected). This period does not 3394 3395 correspond with the temporal requirements for olfactory LTM formation in the standard view of memory dynamics in the honey bee, where a protein-synthesis dependent LTM is expected 3396 after 24-h post conditioning⁸⁴. However, recent work on olfactory memory formation has shown 3397 3398 that protein-synthesis dependent memories arise much earlier and with less conditioning trials than previously thought⁸⁵. Whether our visual conditioning leads to protein-synthesis dependent 3399 LTM, mediated by CREB activation, remains to be determined. 3400

Taken together, our results show both the implication of mushroom bodies in appetitive visual learning in honey bees and the usefulness of Egrl as a marker of neural activity induced by these phenomena under our experimental conditions. The learning success in our VR setup was 50%, which contrasts with the higher learning rates observable for similar color discriminations in the case of free-flying bees. This decrease may be due to several reasons such as the impossibility to return to the hive between rewarded experiences, the tethering conditions and the resulting reduction in active vision. As the tethering impedes, in part, free movements, it may affect the possibility of actively scanning the images perceived, impairing thereby the possibility of extracting target information and learning. In spite of these restrictions, our setup allowed to segregate between *learners* and *non-learners* and achieve relevant analyses to answer questions on the neural and molecular underpinnings of associative visual learning. It constitutes therefore a valuable tool for further studies on the mechanisms of visual cognition in bees.

3414 The protocol used to train the bees in our work consisted in an elemental discrimination between a rewarded and non-rewarded color. Yet, bees are well known for remarkable visual 3415 performances, which include the non-elemental learning of concepts and relational rules⁸⁶⁻⁸⁸. It 3416 is therefore possible that different forms of learning, which recruit different brain regions⁴⁷, 3417 may reveal experience-dependent neural activation through different IEGs and with different 3418 temporal dynamics. Moreover, IEG upregulation may not always be the hallmark of successful 3419 3420 learning as in some cases inhibition of neural activity may be crucial for plastic changes in behavior. Thus, addressing if IEG expression varies qualitatively and quantitatively according 3421 3422 to learning type and complexity is of fundamental importance. Furthermore, including different 3423 intervals post conditioning is important to characterize possible activity changes related to the formation of different memory phases in different regions of the bee brain. Last, but not least, 3424 our results highlight the value of virtual-reality conditions for further explorations of the neural 3425 and molecular underpinnings of visual learning and memory in bees. 3426

3427

3428 METHODS

Honey bee foragers (*Apis mellifera*) were obtained from colonies located in our apiary at the University Paul Sabatier. Only foragers caught upon landing on a gravity feeder filled with a 0.9 M sucrose solution were used in our experiments to ensure high appetitive motivation. Captured bees were brought to the laboratory where they were placed on ice for five minutes to anesthetize them and facilitate the fixation of a tether glued to their thorax by means of melted wax (Fig. 1A). After being attached to the tether, each bee was placed on a small (49 mm diameter) Styrofoam ball for familiarization with the treadmill situation. Bees were provided with 5 μ l of 1.5 M sucrose solution and kept for 3 h in this provisory setup in the dark. They were then moved to the VR arena and used for the experiments.

Once in the VR setup, the bee was attached to a holder that allowed adjusting its position on the treadmill (Fig. 1B), a polystyrene ball (diameter: 5 cm, weight: 1.07 g) held by a 3Dprinted support and floating on a constant airflow produced by an air pump (airflow: 555ml/s; Aqua Oxy CWS 2000, Oase, Wasquehal, France).

3442

3443 VR setup

The VR setup consisted of the treadmill and of a half-cylindrical vertical screen made of semi-3444 3445 transparent tracing paper, which allowed presentation of a 180° visual environment to the bee 3446 (diameter: 268 mm, height: 200 mm, distance to the bee: 9 cm Fig. 1AB) and which was placed in front of the treadmill. The visual environment was projected from behind the screen using a 3447 video projector connected to a laptop (Fig. 1A). The video projector was an Acer K135 (Lamp: 3448 LED, Definition: 1280 x 800, Brightness: 600 lumens, Contrast ratio: 10 000:1, Minimum 3449 Vertical Sync: 50 Hz, Maximum Vertical Sync: 120 Hz, Minimum Horizontal Sync: 30.10³ Hz, 3450 Maximum Horizontal Sync: 100.10^3 Hz)¹⁴. The movements of the walking bee on the treadmill 3451 were recorded by two infrared optic-mouse sensors (Logitech M500, 1000 dpi, Logitech, 3452 Lausanne, Switzerland) placed on the ball support perpendicular to each other. 3453

Experiments were conducted under 3D closed-loop conditions, i.e. rotations of the ball displaced the visual stimuli not only laterally but also towards the bee. To generate these conditions, we developed a custom software by means of the Unity engine (version 2018.3.11f1). The open-source code is available at https://github.com/G-Lafon/BeeVR. The software updated the position of the bee within the VR every 0.017 s. A displacement of 1 cm on the ball corresponds to an equivalent displacement in the VR landscape. Moving 1 cm on the ball towards an object increased the visual angle of the object by ca. 1.7°. Based on the ball movements, our software calculated the position of the walking bee and its heading, and determined which object was centered on the screen.

3463

3464 Visual stimuli

Bees had to discriminate two vertical cuboids (Fig. 1C) based on their different colors and association with reward and punishment. The colors of the cuboids (see Supplementary Fig. 1) were blue (RGB: 0, 0, 255, with a dominant wavelength of 450 nm and an irradiance of 161000 μ W) and green (RGB: 0, 100, 0, with a dominant wavelength of 530 nm and an irradiance of 24370 μ W/cm2). They were displayed on a black background (RGB: 0, 0, 0). These colors were chosen based on previous work showing their successful learning in the VR setup¹⁴.

Each cuboid had a 5×5 cm base and 1 m height so that it occupied the entire vertical extent of the screen irrespective of the bee's position. The cuboids were positioned at -50° and $+50^{\circ}$ from the bee's body axis at the beginning of each trial. Approaching a cuboid within an area of 3 cm surrounding its virtual surface followed by direct fixation of its center was recorded as a choice (Fig. 2A).

3476

3477 Conditioning and testing at the treadmill

3478 Bees were trained using a differential conditioning, which promotes better learning 3479 performances owing to the presence of penalized incorrect color choice that result in an 3480 enhancement of visual attention³⁶.

Bees were trained during 10 consecutive trials using a differential conditioning procedure (Fig. 2B) in which one of the cuboids (i.e. one of the two colors, green or blue) was rewarded with 1.5 M sucrose solution (the appetitive conditioned stimulus or CS+) while the other cuboid displaying the alternative color (the aversive conditioned stimulus or CS-) was associated with 3 M NaCl solution. The latter was used to increase the penalty of incorrect choices^{40,89,90}. To avoid directional biases, the rewarded and the punished color cuboids were swapped between the left and the right side of the virtual arena in a pseudo random manner along trials. Moreover, a reconstruction of the trajectories of the bees analyzed did not show side biases.

3490 A dark screen was shown initially to the bees. During training trials, each bee faced the two cuboids. The bee had to choose the CS+ cuboid by walking towards it and centering it on 3491 the screen. Colors were equally and randomly assigned to the CS+ and the CS- category during 3492 3493 training. If the bee reached the CS+ within an area of 3 cm in the virtual environment (i.e. if the cuboid chosen by the bee subtended 53° in its horizontal extent) and centered it, the screen was 3494 locked during 8 s to ensure fixation. This allowed the delivery of sucrose solution in case of a 3495 3496 correct choice, or of NaCl in case of an incorrect choice. Solutions were delivered for 3 s by the experimenter who sat behind the bee and used a toothpick to this end. The toothpick touched 3497 first the antennae and then the mouthparts during the 8 s in which the screen was locked on the 3498 cuboid fixated by the bee. Each training trial lasted until the bee chose one of both stimuli or 3499 for a maximum of 60 s (no choice). Trials were separated by an inter-trial interval of 60 s during 3500 3501 which the dark screen was presented. Bees that were unable to choose a stimulus (i.e. that did not fulfill the criterion of a choice defined above) in at least 5 trials were excluded from the 3502 analysis. From 216 bees trained, 75 were kept for analysis (~35%). 3503

After the last training trial, each bee was subjected to a non-reinforced test that lasted 60 s (Fig. 2B). Test performance allowed distinguishing *learners* (i.e. bees that chose the CS+ as their first choice in the test) from *non-learners* (i.e. bees that either chose the CS- in their first test choice or that did not make any choice during the test). IEG expression was compared between these two groups, which had the same sensory experience in the VR setup and whichdiffered only in their learning success.

3510

3511 Brain dissection

One hour after the test, bees were decapitated, and the head was instantly frozen in a nitrogen 3512 solution. The period between post-test and brain collection was chosen to allow induction of 3513 the three IEGS studied (typically, 15 or more min in the case of kakusei^{25,46} and 30-60 min in 3514 the case of $Hr38^{31}$ and $Egr1^{30}$). The frozen bee head was dissected on dry ice under a 3515 microscope. First, the antennae were removed and a window was cut in the upper part of the 3516 3517 head capsule, removing the cuticle between the compound eyes and the ocelli. Second, the glands and tracheae around the brain were removed. Third, the retinas of the compound eyes 3518 3519 were also removed.

The frozen brain was cut in three main parts for IEG analyses (Fig. 4A): the optic lobes 3520 (OL), the upper part of the mushroom bodies (the mushroom-body calyces, MB Ca) and the 3521 remaining central brain (CB), which included mainly the central complex (CC), the 3522 3523 subesophageal zone (SEZ) and the peduncula of the mushroom-bodies (α and β lobes). Samples were stored at -80 °C before RNA extraction. During the dissection process, one of these three 3524 3525 regions was lost in 4 non-learners brains As only bees for which all regions were available were kept in the analyses, the sample sizes of the *non-learners* differ between the behavioral (n=18) 3526 and the molecular analyses (n=14). 3527

3528

3529 RNA extraction and reverse transcription

The RNAs from the three sections mentioned above (OL, MB Ca and CB) were extracted and
purified using the RNeasy Micro Kit (Qiagen). The final RNA concentration obtained was
measured by spectrophotometry (NanoDropTM One, Thermo Scientific). A volume of 10 μl

containing 100 ng of the RNA obtained was used for reverse transcription following the procedure recommended in the Maxima H Minus First Strand cDNA Synthesis kit (Thermoscientific, 0.25 μ l of random hexamer primer, 1 μ l of 10 mM dNTP mix, 3.75 μ l of nuclease free H₂O, 4 μ l 5X RT Buffer and 1 μ l Maxima H Minus Enzyme Mix).

3537

3538 **Quantitative Polymerase Chain Reaction (RT-qPCR)**

All the primers used for target and reference genes generated amplification products of approximately 150 pb. The efficiencies of all reactions with the different primers used were between 95 and 110 % (Table 1). Their specificity was verified by analyzing melting curves of the qRT-PCR products (see Supplementary Fig. 2). Two reference genes (*Ef1* α and *Actin*) were used for normalization.

Expression was quantified using a SYBR Green real-time PCR method. Real-time PCR 3544 3545 were carried out in 384-Well PCR Plates (Bio-Rad) cover with Microseal 'B' PCR Plate Sealing Film (Bio-Rad). The PCR reactions were performed using the SsoAdvancedTM Universal 3546 SYBR® Green Supermix (Bio-Rad) in a final volume of 10 µl containing 5 µl of 2X 3547 SsoAdvancedTM Universal SYBR® Green Supermix, 2 µl of cDNA template (1:3 dilution 3548 from the reverse transcription reaction), 0.5 µl of 10 µmol of each primer and 2 µl of ultrapure 3549 3550 water. The reaction conditions were as follows: 95 °C for 30 s followed by 40 cycles of 95 °C for 10 s, 55 °C for 30 s and a final step at 95 °C for 10 s followed by a melt curve from 55 °C 3551 to 95 °C with 0.5 °C per second. The reaction was performed in a CFX384 Touch Real-Time 3552 PCR Detection System (Bio-Rad) and analyzed with the software Bio-Rad CFX Manager. 3553

Each sample was run in triplicates. If the triplicates showed too much variability (SD > 0.3), the furthest triplicate was discarded. If the two remaining triplicates still showed too much variability (SD > 0.3) the sample was discarded. The samples were subjected to a relative quantification and normalization. First for each sample and for each reference gene per brain

region, the relative quantity (Qr) was computed using the difference between the mean Ct value of each sample and the highest mean Ct value (Δ Ct), using the following formula: Qr= (1+E)^{Δ Ct} (with E= efficiency of the reaction). Then a normalization factor for each sample was obtained computing the geometric mean of the relative quantities obtained for the reference genes in the corresponding samples (Δ \DeltaCt).

3563

3564 Data analysis and statistics

3565 Behavioral data

The first choice of the bees was recorded during the conditioning trials and the non-reinforced test. In this way, we established for each trial and test the percentages of bees choosing first each of the stimuli displayed or not choosing a stimulus (\pm 95% confidence interval).

Test percentages were analyzed within groups by means of a generalized linear mixed model (GLMM) for binomial family in which the individual identity (Bee) was considered as a random factor (individual effect) while the choice category (CS+, CS-, NC) was fitted as a fixed effect; z values with corresponding degrees of freedom are reported throughout for this kind of analysis.

For each acquisition trial, we recorded motor variables such as the total distance walked, 3574 the walking speed, and the tortuosity of the trajectories⁹¹. Tortuosity was calculated as the ratio 3575 between the total distance walked and the distance between the first and the last point of the 3576 trajectory connected by an imaginary straight line. When the ratio was 1, or close to 1, 3577 trajectories were straightforward while higher values corresponded to sinuous trajectories⁹¹. 3578 3579 The analysis of these continuous variables was done using a linear mixed model (lmer function) 3580 in which the individual identity (Bee ID) was a random factor and the experimental condition (Condition) and trial number (Trial) were fixed factors⁹¹. Statistical analyses were performed 3581 using with R 3.5.1⁹². The package lme4 was used for GLMMs and LMMs. 3582
Statistical differences in gene expression were assessed for reference genes to check for stability and for target genes within a given brain region using One-Factor ANOVA for independent groups in the case of multiple comparisons or two-sample t test in the case of dual comparisons. Pots hoc comparisons between groups were performed by means of a Tukey test following ANOVA. No cross-comparisons between brain regions or genes were performed due to withinarea normalization procedures. Statistical analyses were done either with R 3.5.1 software⁹² or with Statistica 13 Software (TIBCO® Data Science).

3591

3592

3593 **REPORTING SUMMARY**

Further information on research design is available in the Nature Research Reporting Summarylinked to this article.

3596

3597 DATA AND CODE AVAILABILITY

The datasets generated during this study are available at figshare.com with the following accession ID: <u>https://figshare.com/s/1e868800d08a17dc300e</u>

3600

3601 ACKNOWLEDGMENTS

We thank three anonymous reviewers for constructive criticisms and Shiori Iino and Takeo Kubo for providing useful information on the timing of IEG expression. We also thank Benjamin H. Paffhausen, Marco Paoli and Dorian Champelovier for valuable discussions. This work was supported by an ERC Advanced Grant ('Cognibrains') to M.G, who also thanks the Institut Universitaire de France (IUF), the CNRS and the University Paul Sabatier for support.

144

3608 CONTRIBUTIONS

3609 GL performed the behavioral experiments. HG dissected and sectioned the brains of the bees

trained in the VR setup and performed all the molecular analyses. Behavioral experiments were

3611 supervised by AB, AAW and MG. Molecular experiments were supervised by IM and MG

3612 Statistical analyses on behavioral data were performed by GL and MG. Statistical analyses on

3613 gene-expression data were performed by HG and MG. The manuscript was written by MG who

3614 also obtained the funding. All authors reviewed and approved the final version of the

- 3615 manuscript.
- 3616

3617 ETHICS DECLARATIONS

- 3618 **Competing interests**
- 3619 The authors declare no competing interests.
- 3620
- 3621

3622 **REFERENCES**

3623 1 Giurfa, M. Cognition with few neurons: higher-order learning in insects. Trends in 3624 neurosciences 36, 285-294 (2013). 3625 2 Heisenberg, M. Mushroom body memoir: from maps to models. Nat Rev Neurosci 4, 266-275 3626 (2003).3627 3 Cognigni, P., Felsenberg, J. & Waddell, S. Do the right thing: neural network mechanisms of memory formation, expression and update in Drosophila. Current opinion in neurobiology 49, 3628 3629 51-58 (2018). 3630 4 Benjamin, P. R., Kemenes, G. & Kemenes, I. Non-synaptic neuronal mechanisms of learning and memory in gastropod molluscs. Frontiers in bioscience : a journal and virtual library 13, 3631 4051-4057 (2008). 3632 3633 Giurfa, M. Behavioral and neural analysis of associative learning in the honeybee: a taste 5 3634 from the magic well. J Comp Physiol A 193, 801-824 (2007). Kandel, E. R. The molecular biology of memory storage: a dialogue between genes and 3635 6 synapses. Science (New York, N.Y.) 294, 1030-1038 (2001). 3636 7 Menzel, R. Memory dynamics in the honeybee. J Comp Physiol A 185, 323-340 (1999). 3637 Menzel, R. The honeybee as a model for understanding the basis of cognition. Nature Rev 3638 8 3639 Neurosci 13, 758-768 (2012).

3640 9 Giurfa, M. & Sandoz, J. C. Invertebrate learning and memory: fifty years of olfactory 3641 conditioning of the proboscis extension response in honeybees. Learning & memory (Cold 3642 Spring Harbor, N.Y.) 19, 54-66 (2012). 3643 10 Avargues-Weber, A., Deisig, N. & Giurfa, M. Visual cognition in social insects. Annual review 3644 of entomology 56, 423-443 (2011). Avargues-Weber, A., Mota, T. & Giurfa, M. New vistas on honey bee vision. Apidologie 43, 3645 11 3646 244-268 (2012). 3647 12 Buatois, A. et al. Associative visual learning by tethered bees in a controlled visual 3648 environment. Sci Reports 7, 127903 (2017). 3649 13 Schultheiss, P., Buatois, A., Avarguès-Weber, A. & Giurfa, M. Using virtual reality to study 3650 visual performances of honeybees. Curr Opin Insect Sci 24, 43-50 (2017). 3651 14 Buatois, A., Flumian, C., Schultheiss, P., Avargues-Weber, A. & Giurfa, M. Transfer of visual 3652 learning between a virtual and a real environment in honey bees: the role of active vision. 3653 Front Behav Neurosci 12, 139 (2018). 3654 15 Rusch, C., Roth, E., Vinauger, C. & Riffell, J. A. Honeybees in a virtual reality environment 3655 learn unique combinations of colour and shape. J Exp Biol 220, 3478-3487 (2017). 3656 16 Zwaka, H. et al. Learning and its neural correlates in a virtual environment for honeybees. Front Behav Neurosci 12, 279 (2018). 3657 Rusch, C., Alonso San Alberto, D. & Riffell, J. A. Visuo-motor feedback modulates neural 3658 17 3659 activities in the medulla of the honeybee, Apis mellifera. J Neurosci 41, 3192-3203 (2021). 3660 18 Buatois, A., Laroche, L., Lafon, G., Avargues-Weber, A. & Giurfa, M. Higher-order 3661 discrimination learning by honeybees in a virtual environment. The European journal of 3662 neuroscience 51, 681-694 (2020). 3663 19 Clayton, D. F. The genomic action potential. *Neurobiol Learn Mem* 74, 185-216 (2000). 20 3664 Bahrami, S. & Drablos, F. Gene regulation in the immediate-early response process. Adv Biol 3665 Regul 62, 37-49 (2016). 3666 21 Minatohara, K., Akiyoshi, M. & Okuno, H. Role of immediate-early genes in synaptic plasticity 3667 and neuronal ensembles underlying the memory trace. Frontiers in molecular neuroscience 8, 3668 78 (2015). 22 Gallo, F. T., Katche, C., Morici, J. F., Medina, J. H. & Weisstaub, N. V. Immediate early genes, 3669 3670 memory and psychiatric disorders: focus on *c-Fos, Egr1* and *Arc. Front Behav Neurosci* 12, 79 3671 (2018). 23 He, Q., Wang, J. & Hu, H. Illuminating the activated brain: emerging activity-dependent tools 3672 3673 to capture and control functional neural circuits. Neuroscience bulletin 35, 369-377 (2019). 3674 24 Sommerlandt, F. M. J., Brockmann, A., Roessler, W. & Spaethe, J. Immediate early genes in 3675 social insects: a tool to identify brain regions involved in complex behaviors and molecular 3676 processes underlying neuroplasticity. Cell Mol Life Sci 76, 637-651 (2019). 3677 25 Kiya, T., Kunieda, T. & Kubo, T. Increased neural activity of a mushroom body neuron subtype 3678 in the brains of forager honeybees. PLoS One 2, e371 (2007). 3679 26 Kiya, T. & Kubo, T. Dance type and flight parameters are associated with different mushroom 3680 body neural activities in worker honeybee brains. PLoS One 6, e19301 (2011). 3681 27 Kiya, T., Kunieda, T. & Kubo, T. Inducible- and constitutive-type transcript variants of kakusei , 3682 a novel non-coding immediate early gene, in the honeybee brain. Insect Mol Biol 17, 531-536 3683 (2008). 3684 28 Fujita, N. et al. Visualization of neural activity in insect brains using a conserved immediate 3685 early gene, Hr38. Curr Biol 23, 2063-2070 (2013). 3686 29 Singh, A. S., Shah, A. & Brockmann, A. Honey bee foraging induces upregulation of early 3687 growth response protein 1, hormone receptor 38 and candidate downstream genes of the 3688 ecdysteroid signalling pathway. Insect Mol Biol 27, 90-98 (2018). 3689 30 lino, S. et al. Neural activity mapping of bumble bee (Bombus ignitus) brains during foraging 3690 flight using immediate early genes. Sci Rep 10, 7887 (2020).

3691 31 Ugajin, A. et al. Identification and initial characterization of novel neural immediate early 3692 genes possibly differentially contributing to foraging-related learning and memory processes 3693 in the honeybee. Insect Mol Biol 27, 154-165 (2018). 3694 32 Lutz, C. C. & Robinson, G. E. Activity-dependent gene expression in honey bee mushroom 3695 bodies in response to orientation flight. J Exp Biol 216, 2031-2038 (2013). 3696 Shah, A., Jain, R. & Brockmann, A. Egr-1: A candidate transcription factor involved in 33 molecular processes underlying time-memory. Frontiers in Psychology 9 (2018). 3697 3698 34 Giurfa, M. Conditioning procedure and color discrimination in the honeybee Apis mellifera. 3699 Naturwissenschaften 91, 228-231 (2004). 3700 35 Dyer, A. G. & Chittka, L. Fine colour discrimination requires differential conditioning in 3701 bumblebees. Naturwissenschaften 91, 224-227 (2004). 3702 36 Avarguès-Weber, A. & Giurfa, M. Cognitive components of color vision in honey bees: how 3703 conditioning variables modulate color learning and discrimination. J Comp Physiol A 200, 449-3704 461 (2014). 3705 37 Avarguès-Weber, A., de Brito Sanchez, M. G., Giurfa, M. & Dyer, A. G. Aversive reinforcement 3706 improves visual discrimination learning in free-flying honeybees. PLoS One 5, e15370 (2010). 3707 38 Menzel, R. The insect mushroom body, an experience-dependent recoding device. Journal of 3708 physiology, Paris 108, 84-95 (2014). 3709 39 de Brito Sanchez, M. G., Serre, M., Avarguès-Weber, A., Dyer, A. G. & Giurfa, M. Learning 3710 context modulates aversive taste strength in honey bees. J Exp Biol 218, 949-959 (2015). 3711 40 Aguiar, J., Roselino, A. C., Sazima, M. & Giurfa, M. Can honey bees discriminate between 3712 floral-fragrance isomers? J Exp Biol 221 (2018). 3713 41 Dyer, A. G. & Chittka, L. Bumblebees (Bombus terrestris) sacrifice foraging speed to solve 3714 difficult colour discrimination tasks. J Comp Physiol A 190, 759-763 (2004). 3715 42 Ings, T. C. & Chittka, L. Speed-accuracy tradeoffs and false alarms in bee responses to cryptic 3716 predators. Curr Biol 18, 1520-1524 (2008). 3717 43 Burns, J. G. & Dyer, A. G. Diversity of speed-accuracy strategies benefits social insects. Curr 3718 Biol 18, R953-R954 (2008). 3719 44 Marchal, P. et al. Inhibitory learning of phototaxis by honeybees in a passive-avoidance task. 3720 Learning & memory (Cold Spring Harbor, N.Y.) 26, 412-423 (2019). 3721 45 Kiya, T. & Kubo, T. Analysis of GABAergic and non-GABAergic neuron activity in the optic 3722 lobes of the forager and re-orienting worker honeybee (Apis mellifera L.). PLoS One 5, e8833 3723 (2010). 3724 46 Ugajin, A. et al. Detection of neural activity in the brains of Japanese honeybee workers 3725 during the formation of a "hot defensive bee ball". *PLoS One* 7, e32902 (2012). 3726 47 Devaud, J. M. et al. Neural substrate for higher-order learning in an insect: Mushroom bodies 3727 are necessary for configural discriminations. Proc Natl Acad Sci U S A 112, E5854-5862 3728 (2015). 3729 48 Guven-Ozkan, T. & Davis, R. L. Functional neuroanatomy of Drosophila olfactory memory 3730 formation. Learning & memory (Cold Spring Harbor, N.Y.) 21, 519-526 (2014). 3731 49 Paulk, A. C., Phillips-Portillo, J., Dacks, A. M., Fellous, J. M. & Gronenberg, W. The processing 3732 of color, motion, and stimulus timing are anatomically segregated in the bumblebee brain. J 3733 Neurosci 28, 6319-6332 (2008). Paulk, A. C., Dacks, A. M., Phillips-Portillo, J., Fellous, J. M. & Gronenberg, W. Visual 3734 50 3735 processing in the central bee brain. J Neurosci 29, 9987-9999 (2009). 3736 51 Menzel, R. & Backhaus, W. in Vision and Visual Dysfunction. The Perception of Colour. (ed P. 3737 Gouras) 262-288 (MacMillan Press, 1991). Mota, T., Yamagata, N., Giurfa, M., Gronenberg, W. & Sandoz, J. C. Neural organization and 3738 52 3739 visual processing in the anterior optic tubercle of the honeybee brain. J Neurosci 31, 11443-3740 11456 (2011). 3741 53 Paulk, A. C., Dacks, A. M. & Gronenberg, W. Color processing in the medulla of the 3742 bumblebee (Apidae: Bombus impatiens). J Comp Neurol 513, 441-456 (2009).

Paulk, A. C. & Gronenberg, W. Higher order visual input to the mushroom bodies in the bee, Bombus impatiens. Arthropod Struct Dev 37, 443-458 (2008). Dyer, A. G., Paulk, A. C. & Reser, D. H. Colour processing in complex environments: insights from the visual system of bees. Proc Biol Sci 278, 952-959 (2011). Kirschner, S. et al. Dual olfactory pathway in the honeybee, Apis mellifera. J Comp Neurol , 933-952 (2006). Ehmer, B. & Gronenberg, W. Segregation of visual input to the mushroom bodies in the honeybee (Apis mellifera). J Comp Neurol 451, 362-373 (2002). Kirkerud, N. H., Schlegel, U. & Giovanni Galizia, C. Aversive learning of colored lights in walking honeybees. Front Behav Neurosci 11, 94 (2017). Plath, J. A. et al. Different roles for honey bee mushroom bodies and central complex in visual learning of colored lights in an aversive conditioning assay. Front Behav Neurosci 11, 98 (2017). Vergoz, V., Roussel, E., Sandoz, J. C. & Giurfa, M. Aversive learning in honeybees revealed by the olfactory conditioning of the sting extension reflex. *PLoS One* **2**, e288 (2007). Tedjakumala, S. R., Aimable, M. & Giurfa, M. Pharmacological modulation of aversive responsiveness in honey bees. Front Behav Neurosci 7 (2014). Wolf, R. et al. Drosophila mushroom bodies are dispensable for visual, tactile, and motor learning. Learning & memory (Cold Spring Harbor, N.Y.) 5, 166-178 (1998). Heisenberg, M., Borst, A., Wagner, S. & Byers, D. Drosophila mushroom body mutants are deficient in olfactory learning. J Neurogenet 2, 1-30 (1985). Ofstad, T. A., Zuker, C. S. & Reiser, M. B. Visual place learning in Drosophila melanogaster. Nature 474, 204-U240 (2011). Vogt, K. et al. Shared mushroom body circuits underlie visual and olfactory memories in Drosophila. Elife 3, e02395 (2014). Szyszka, P., Ditzen, M., Galkin, A., Galizia, C. G. & Menzel, R. Sparsening and temporal sharpening of olfactory representations in the honeybee mushroom bodies. J Neurophysiol , 3303-3313 (2005). Perez-Orive, J. et al. Oscillations and sparsening of odor representations in the mushroom body. Science 297, 359-365 (2002). Laurent, G. J. et al. Odor encoding as an active, dynamical process: experiments, computation, and theory. Annu Rev Neurosci 24, 263-297 (2001). Lin, A. C., Bygrave, A. M., de Calignon, A., Lee, T. & Miesenbock, G. Sparse, decorrelated odor coding in the mushroom body enhances learned odor discrimination. Nat Neurosci 17, 559-568 (2014). Froese, A., Szyszka, P. & Menzel, R. Effect of GABAergic inhibition on odorant concentration coding in mushroom body intrinsic neurons of the honeybee. J Comp Physiol A 200, 183-195 (2014). Papadopoulou, M., Cassenaer, S., Nowotny, T. & Laurent, G. Normalization for sparse encoding of odors by a wide-field interneuron. Science 332, 721-725 (2011). Rybak, J. & Menzel, R. Anatomy of the mushroom bodies in the honey bee brain: The neuronal connections of the alpha-lobe. J Comp Neurobiol 334, 444-465 (1993). Zwaka, H., Bartels, R., Grunewald, B. & Menzel, R. Neural Organization of A3 Mushroom Body Extrinsic Neurons in the Honeybee Brain. Front Neuroanat 12, 57 (2018). Suenami, S., Oya, S., Kohno, H. & Kubo, T. Kenyon cell subtypes/populations in the honeybee mushroom bodies: possible function based on their gene expression profiles, differentiation, possible evolution, and application of genome editing. Front Psychol 9 (2018). Kaneko, K. et al. Novel middle-type Kenyon cells in the honeybee brain revealed by area-preferential gene expression analysis. PLoS One 8, e71732 (2013). Strausfeld, N. J. Organization of the honey bee mushroom body: representation of the calyx within the vertical and gamma lobes. J Comp Neurol 450, 4-33 (2002).

- 379477Shah, A., Jain, R. & Brockmann, A. Egr-1: A Candidate transcription factor involved in3795molecular processes underlying time-memory. Front Psychol **9** (2018).
- 3796 78 Ugajin, A., Kunieda, T. & Kubo, T. Identification and characterization of an Egr ortholog as a
 3797 neural immediate early gene in the European honeybee (*Apis mellifera* L.). *FEBS Letters* 587,
 3798 3224-3230 (2013).
- 3799 79 Gehring, K. B., Heufelder, K., Kersting, I. & Eisenhardt, D. Abundance of phosphorylated *Apis* 3800 *mellifera* CREB in the honeybee's mushroom body inner compact cells varies with age. J
 3801 *Comp Neurol* 524, 1165-1180 (2016).
- 380280Silva, A. J., Kogan, J. H., Frankland, P. W. & Kida, S. CREB and memory. Annual review of3803neuroscience 21, 127-148 (1998).
- 380481Kandel, E. R. The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CPEB.3805Mol Brain 5, 14 (2012).
- 380682Yin, J. C. P. & Tully, T. CREB and the formation of long-term memory. Current opinion in3807neurobiology 6, 264-268 (1996).
- 380883Alberini, C. M. Transcription factors in long-term memory and synaptic plasticity.3809Physiological reviews 89, 121-145 (2009).
- Wüstenberg, D., Gerber, B. & Menzel, R. Long- but not medium-term retention of olfactory
 memory in honeybees is impaired by actinomycin D and anisomycin. *The European journal of neuroscience* 10, 261-261 (1998).
- 381385Villar, M. E., Marchal, P., Viola, H. & Giurfa, M. Redefining single-trial memories in the honey3814bee. Cell Reports **30**, 2603-2613 (2020).
- 381586Avarguès-Weber, A. & Giurfa, M. Conceptual learning by miniature brains. Proc Biol Sci3816(2013).
- 3817 87 Giurfa, M. An Insect's Sense of Number. *Trends Cogn Sci* 23, 720-722 (2019).
- 381888Giurfa, M. Learning of sameness/difference relationships by honey bees: performance,3819strategies and ecological context. Curr Opin Behav Sci 37, 1-6 (2021).
- 382089de Brito Sanchez, M. G., Serre, M., Avargues-Weber, A., Dyer, A. G. & Giurfa, M. Learning3821context modulates aversive taste strength in honey bees. J Exp Biol **218**, 949-959 (2015).
- 382290Bestea, L. *et al.* Peripheral taste detection in honey bees: what do taste receptors respond3823to? *The European journal of neuroscience* (2021).
- 382491Lafon, G., Howard, S. R., Paffhausen, B. H., Avarguès-Weber, A. & Giurfa, M. Motion cues3825from the background influence associative color learning of honey bees in a virtual-reality3826scenario. Sci Rep 11, 21127 (2021).
- 382792R Development Core Team. R: A Language and Environment for Statistical Computing. (The R3828Foundation for Statistical Computing, 2016).
- 3829

Chapter 3

- 3833 The neural signature of visual learning under restrictive virtual-reality
- 3834 conditions



in Behavioral Neuroscience

OPEN ACCESS

frontiers

ORIGINAL RESEARCH published: 16 February 2022 doi: 10.3389/inbeh.2022.846076



The Neural Signature of Visual Learning Under Restrictive Virtual-Reality Conditions

Gregory Lafon^{1‡}, Haiyang Geng^{1,2‡}, Aurore Avarguès-Weber¹, Alexis Buatois^{1†5}, Isabelle Massou¹⁵ and Martin Giurfa^{1,2,3+5}

¹ Research Center on Animal Cognition, Center for Integrative Biology, CNRS, University of Toulouse, Toulouse, France, ² College of Animal Sciences (College of Bee Science), Fujian Agriculture and Forestry University, Fuzhou, China, ³ Institut Universitaire de France, Paris, France

3838

3839 In the second chapter, we found that visual learning under 3D VR conditions led to an upregulation of *Egr1* in the calvees of the mushroom bodies. We thus asked a different question, 3840 namely whether the constraints of the VR environmental impact on neural activation in a color 3841 3842 discrimination task. Specifically, we asked if learning the same color discrimination as in the previous chapter, but this time in a 2D VR, which sets higher movement constraints, results in 3843 similar IEG expression patters as those detected in the 3D environment. Surprisingly, the 3844 3845 comparison between non-learners and learners revealed a completely different pattern of activation from the one found in the second chapter. Here, Egrl was downregulated in the optic 3846 3847 lobes, while Hr38 and kakusei were coincidently downregulated in the mushroom body calyces. 3848 In this chapter, we present these results and offer some possible explanations as to why 2D conditioning leads to a different pattern of IEG expression. In particular, we interpret this 3849 3850 downregulation as a reflect of a higher neural inhibition in the 2D VR discrimination.

3852	The neural signature of visual learning under restrictive virtual-
3853	reality conditions
3854	
3855	Gregory Lafon ^{1*} , Haiyang Geng ^{*1,2} , Aurore Avarguès-Weber ¹ , Alexis
3856	Buatois ^{1,¥,§} , Isabelle Massou ^{1,§} , Martin Giurfa ^{1,2,3§}
3857	
3858 3859	¹ Research Centre on Animal Cognition, Center for Integrative Biology, CNRS, University of Toulouse, 118 route de Narbonne, F-31062 Toulouse cedex 09, France.
3860 3861	² College of Animal Sciences (College of Bee Science), Fujian Agriculture and Forestry University, Fuzhou 350002, China.
3862	³ Institut Universitaire de France, Paris, France (IUF).
3863	
3864	
3865	* Equal contribution
3866	[§] Senior authorship shared
3867	${}^{\mathtt{F}}$ Present address: Institute of Neuroscience and Physiology, Department of Neurochemistry and
3868	Psychiatry, University of Gothenburg, Su Sahlgrenska, 41345 Göteborg, Sweden.
3869	
3870	Corresponding author: Dr. Martin Giurfa
3871	Research Centre on Animal Cognition, CNRS – UPS, 31062 Toulouse cedex 9, France
3872	martin.giurfa@univ-tlse3.fr
3873	
3874	Subject Areas: Behavior, Neuroscience
3875	
3876	

3877 Abstract

Honey bees are reputed for their remarkable visual learning and navigation capabilities. These 3878 capacities can be studied in virtual reality (VR) environments, which allow studying 3879 performances of tethered animals in stationary flight or walk under full control of the sensory 3880 environment. Here we used a 2D VR setup in which a tethered bee walking stationary under 3881 3882 restrictive closed-loop conditions learned to discriminate vertical rectangles differing in color and reinforcing outcome. Closed-loop conditions restricted stimulus control to lateral 3883 3884 displacements. Consistently with prior VR analyses, bees learned to discriminate the trained stimuli. Ex vivo analyses on the brains of learners and non-learners showed that successful 3885 3886 learning led to a downregulation of three immediate early genes in the main regions of the visual 3887 circuit, the optic lobes (OLs) and the calves of the mushroom bodies (MBs). While Egrl was 3888 downregulated in the OLs, Hr38 and kakusei were coincidently downregulated in the calyces of the MBs. Our work thus reveals that color discrimination learning induced a neural signature 3889 3890 distributed along the sequential pathway of color processing that is consistent with an inhibitory trace. This trace may relate to the motor patterns required to solve the discrimination task, which 3891 are different from those underlying pathfinding in 3D VR scenarios allowing for navigation and 3892 3893 exploratory learning and which lead to IEG upregulation.

3894

3895 Keywords

3896 Vision – Visual Learning – Virtual Reality – Honey Bee Brain – Immediate Early Genes –
 3897 Mushroom Bodies – Optic Lobes

Learning relies on changes in neural activity and/or connectivity in the nervous system, which 3900 underlie the acquisition of new, durable information based on individual experience. 3901 3902 Invertebrate models have proved to be extremely influential to characterize learning and memory at multiple levels, not only because they allow determining where and when such 3903 changes occur in the nervous system ¹⁻⁷ but also because their behavioral performances can be 3904 studied in standardized laboratory protocols that allow full control over the sensory variables 3905 3906 that animals should learn and memorize. A paradigmatic example is provided by the honey bee Apis mellifera, where the study of olfactory learning and memory experienced significant 3907 3908 progresses thanks to the availability of a Pavlovian conditioning protocol that offers the 3909 possibility of acquiring consistent behavioral data coupled with the simultaneous use of invasive methods to record neural activity^{5, 8-10}. In this protocol, termed the olfactory 3910 conditioning of the proboscis extension reflex (PER), harnessed bees learn to associate an 3911 odorant with a reward of sucrose solution^{10, 11}. The immobility imposed to the trained bees and 3912 the Pavlovian nature of the association learned (the odorant acts as the conditioned stimulus 3913 and the sucrose reward as the unconditioned stimulus) allows a full control over the stimulations 3914 provided and thus a fine characterization of behavioral changes due to learning and memory. 3915

3916 In the case of visual learning by honey bees, this possibility is reduced as performances are mostly studied in free-flying foragers^{5, 12} under semi-natural conditions. Yet, virtual-reality 3917 (VR) environments have been recently developed to overcome this limitation¹³ as they provide 3918 not only a full control over the visual surrounding of an experimental subject, be it tethered or 3919 3920 not, but also the delivery of physically impossible ambiguous stimuli, which give conflicting visual information¹⁴. In one type of VR that we developed in the last years, a tethered bee walks 3921 stationary on a treadmill while being exposed to a controlled visual environment displayed by 3922 3923 a video projector. Bees can then be trained with virtual targets that are paired with gustatory reward or punishment^{13, 15-19}. To create an immersive environment, closed-loop visual conditions are used in which the variations of the visual panorama are determined by the walking movements of the bee on the treadmill. Under these conditions, bees learn and memorize simple^{15, 19} and higher-order²⁰ visual discriminations, which offers the potential for mechanistic analyses of visually-oriented performances ^{17, 18}.

We have used two different types of closed loop situation so far: a restrictive 2D 3929 situation, in which bees can displace conditioned targets only frontally (i.e. from left to right 3930 and vice versa)^{15, 19, 20}, and a more realistic 3D situation which includes a depth dimension so 3931 that targets expand upon approach and retract upon distancing²¹. Although bees learn to 3932 3933 discriminate color stimuli in both conditions, the processes underlying such learning may differ given the different conditions imposed to the bees in terms of stimulus control. Indeed, while 3934 in 3D scenarios movement translates into a displacement and a recognizable change in the 3935 3936 visual scene, which can then be computed against the available internal information about the displacement, 2D scenarios are restricted to the execution of actions that are dependent on 3937 3938 reinforcement contingency. These two conditions may give rise to different mechanisms of 3939 information acquisition.

In a recent work, we studied color learning in the 3D scenario and quantified immediate 3940 early genes (IEGs) in the brain of learners and non-learners to uncover the regions that are 3941 involved in this discrimination learning²². IEGs are efficient markers of neural activity as they 3942 are transcribed transiently and rapidly in response to specific stimulations inducing neural 3943 activity without *de novo* protein synthesis²³⁻²⁵. Three IEGs were quantified on the basis of 3944 numerous reports that associated them with foraging and orientation activities²⁶⁻³⁰: kakusei, a 3945 nuclear non-coding RNA³¹, the hormone receptor 38 gene (Hr38), a component of the 3946 ecdysteroid signalling pathway³², and the early growth response gene-1 (*Egr1*), which is a 3947 major mediator and regulator of synaptic plasticity and neuronal activity³³. We found that color 3948

learning in the 3D VR environment was associated with an *upregulation* of *Egr1* in the calyces of the mushroom bodies²², a main structure of the insect brain repeatedly associated with the storage and retrieval of olfactory memories^{2, 9}. No other changes of IEG expression were detected in other regions of the brain, thus underlining the relevance of mushroom bodies for color learning and retention²².

3954 Here we asked if color learning in the more restrictive 2D VR environment induces 3955 changes in IEG expression, both at the gene level and at the brain region level, similar to those 3956 detected in the 3D VR system. Asking this question is important to determine if changes in IEG expression differ according to the degrees of freedom of the VR system and the distinct motor 3957 3958 patterns that are engaged in either case. Despite the similarity in behavioral performance (bees learn to discriminate colors in both scenarios), we reasoned that the processes underlying 3959 learning may be different given the restrictive conditions of the 2D VR, which demand a tight 3960 3961 stimulus control while the 3D VR enables exploration of the virtual environment. We thus studied color learning in the 2D VR environment and performed ex vivo analyses to map IEG 3962 3963 expression in brain areas of learners and non-learners, which had the same sensory experience 3964 and only differed in terms of learning success.

3965

3966

3967 **Results**

3968 Behavioral analyses

Honey bee foragers from a hive located in our apiary were captured at an artificial feeder to which they were previously trained. They were enclosed in individual glass vials and brought to the laboratory where they were prepared for the VR experiments. A tether was glued on their thorax (Fig. 1A,B), which allowed to attach them to a holder to adjust their position on a treadmill. The treadmill was a polystyrene ball that was suspended on an air cushion produced by an air pumping system (see Methods for details). The bee suspended from its tether could walk stationary on the treadmill; its movements were recorded by two infrared optic-mouse sensors placed on the ball support perpendicular to each other, which allowed to reconstruct the trajectories and quantify motor parameters. A semi-cylindrical screen made of semitransparent tracing paper was placed in front of the treadmill (i.e. of the walking bee; Fig. 1A). Images were projected onto this screen by a video projector placed behind it.

Bees were trained to discriminate a green from a blue vertical bar against a black 3980 background during ten conditioning trials (Fig. 1C; see Supplementary Fig. 1 for color 3981 characteristics). Experiments were performed under 2D closed-loop conditions so that the 3982 3983 movements of the walking bee displaced the bars laterally on the screen to bring them towards or away from front of the bee. During training, one of the bars (CS+) was rewarded with 1 M 3984 sucrose solution while the other bar (CS-) was punished with an aversive 3M NaCl solution³⁴⁻ 3985 36 . A choice was recorded when the bee moved one rectangle to the center of the screen (i.e., 3986 between -12.5° and $+12.5^{\circ}$ of the bee's central axis; see Fig. 1D, right). 3987

3988 We segregated learners and non-learners according to the bees' performance in a 3989 dedicated unrewarded test at the end of the training. Learners (n=23) were those bees that showed successful discrimination in the test (i.e. which chose the CS+). Non-learners (n=17), 3990 3991 were those bees that did not succeed in the test (i.e. they either chose the CS- or did not make a choice). Importantly, these bees have the same sensory experience in terms of exposure to the 3992 3993 color stimuli and reinforcements as our training procedure froze the CS+ or the CS- stimuli in front of the bee during 8 s upon a choice and delivered the reinforcements accordingly. Bees 3994 3995 that did neither choose the CS+ nor the CS- in at least 5 trials were excluded from the analysis. Acquisition was significant for learners during conditioning trials (Fig. 2A; CS*Trial 3996 3997 effect: $\chi 2=47.746$, df:2, p<0.0001), thus showing that the categorization made based on test performance reflected well learning success. The percentages of bees responding to the CS+ 3998

and to the CS- differed significantly along trials (CS+ vs. CS-: CS*Trial; z=6.845, p<0.0001). 3999 Significant differences were also found between the bees responding to the CS- and the non-4000 4001 responders (CS- vs NC: CS*Trial; z=3.541, p=0.0004) but not between bees responding to the CS+ and non-responders (CS+ vs. NC: CS*Trial; z=-1.201, p=0.23). Non-learners (n=17) did 4002 4003 also show a significant CS*Trial effect (Fig. 2B; χ 2=9.8383, df:2, p=0.007), but this effect was introduced by the non-responders. These bees differed significantly along trials both from the 4004 bees responding to the CS+ (CS+ vs. NC: CS*Trial; z=2.356, p=0.019) and from the bees 4005 4006 responding to the CS- (CS- vs. NC: CS*Trial; z=3.068, p=0.002). On the contrary, the 4007 percentages of bees responding to the CS+ and to the CS- did not vary along trials (CS+ vs. CS-: CS*Trial; z=1.437, p=0.2), consistently with the absence of learning. 4008



Figure 1. Experimental setup, choice criterion and conditioning procedure. A) Global 4010 4011 view of the setup. 1: Semicircular projection screen made of tracing paper. 2: Holding frame 4012 to place the tethered bee on the treadmill. 3: The treadmill was a Styrofoam ball positioned within a cylindrical support (not visible) floating on an air cushion. 4: Infrared mouse optic 4013 sensors allowing to record the displacement of the ball and to reconstruct the bee's trajectory. 4014 4015 5: Air arrival. The video projector displaying images on the screen from behind can be seen on top of the image. B) The tethering system. 1: Plastic cylinder held by the holding frame; the 4016 cylinder contained a glass cannula into which a steel needle was inserted. 2: The needle was 4017 4018 attached to the thorax of the bee. 3: Its curved end was fixed to the thorax by means of melted bee wax. C) Color discrimination learning in the VR setup. The bee had to learn to 4019 discriminate two vertical bars based on their different color and their association with reward 4020 4021 and punishment. Bars were green and blue on a dark background. Color intensities were adjusted to avoid phototactic biases independent of learning. Displacement of the bars was 4022

restricted to the 2D plane in front of the bee. D) Left: view of the stimuli at the start of a trial 4023 or test. The green and the blue virtual bars were a presented at -50° and $+50^{\circ}$ of the bee's 4024 longitudinal axis of the bee. Stimuli could be only displaced by the bee from left to right and 4025 vice versa (double red arrow). The red angles on the virtual surface indicate the visual angle 4026 subtended by each bar at the bee position ($\Box = 31.05^{\circ}$). **Right: Choice of a bar.** A choice was 4027 4028 recorded when the bee kept the center of the object between -12.5° and $+12.5^{\circ}$ in front of it for 4029 1 second. The bar image was then frozen during 8 s and the corresponding reinforcement (US) was delivered. E) Conditioning protocol. Bees were trained along 10 conditioning trials that 4030 lasted a maximum of 1 min and that were spaced by 1 min (intertrial interval). After the end of 4031 conditioning, and following an additional interval of 1 min, bees were tested in extinction 4032 conditions during 1 min. 4033

4034

Learners and non-learners did not differ in their motor activity during training, thus 4035 excluding this factor as determinant of possible changes in neural activity. When walking 4036 speeds and the distances travelled were compared between groups, no significant differences 4037 were detected (*Distance*: Group; $\chi^2=1.93$, df:1, p=0.16; *Speed*: Group; $\chi^2=1.78$, df:1, p=0.18). 4038 In the non-reinforced test, per definition learners (Fig. 2C) chose correctly the CS+ 4039 4040 (100% of the bees) while non-learners (Fig. 2D) did either chose the CS- (35%) or did not 4041 perform any choice (65%). Learners spent more time fixating the CS+ than the CS- consistently with the choice made during the test (Wilcoxon signed rank exact test: V=17, p<0.0001) while 4042 non-learners did not differ in their fixation time for both stimuli in spite of a tendency to fixate 4043 4044 more the CS- (V=26, p=0.05).



4046 Figure 2. Acquisition and test performances of learners and non-learners. A) Acquisition 4047 **performance of learners** (i.e. bees that chose the CS+ in the non-reinforced test; n=23). The red, black and grey curves show the percentages of bees choosing the CS+, the CS- or not 4048 4049 making a choice (NC), respectively. Bees learned the discrimination between CS+ and CS-. B) Acquisition performance of non-learners (i.e. bees that chose the CS- or did not make a 4050 4051 choice in the non-reinforced test; n=17). These bees did not learn to discriminate the CS+ from 4052 the CS-. In A) and B) shaded areas around curves indicate the 95% confidence interval. C) Test performance of learners (% of bees choosing either the CS+, the CS- or not making a choice). 4053 Per definition these bees only chose the CS+ first. D) Test performance of non-learners. (% 4054 4055 of bees choosing either the CS+, the CS- or not making a choice). Per definition these bees chose either the CS- or did not make a choice (NC). In C) and D), error bars represent the 95% 4056 confidence interval. E) Time (s) spent by learners fixating the CS+ and the CS- during the 4057 test. Learners spent more time fixating the CS+ consistently with their stimulus choice. Bars 4058 represent the time spent keeping the object within $-12.5^{\circ}/+12.5^{\circ}$ in front of the bee. Scatter plots 4059 represent individual fixation times. ****: p < 0.0001. F) Time (s) spent by non-learners 4060 fixating the CS+ and the CS- during the test. Non-learners did not differ in their fixation time 4061 of the CS+ and the CS-. Bars represent the time spent keeping the object within $-12.5^{\circ}/+12.5^{\circ}$ 4062 in front of the bee. Scatter plots represent individual fixation times. NS: non-significant. In E) 4063 4064 and **F**), error bars represent the 95% confidence interval.

4065

161

We aimed at determining if visual learning in the 2D VR induces transcriptional changes 4067 revealing the neural trace of the associative learning described in the previous section. To this 4068 4069 end, we performed RT-qPCR in individual brains of learners (n=22; one learner brain was lost during the dissection process) and non-learners (n=17), focusing on three main brain sections 4070 4071 (Fig. 3A): the optic lobes (OLs), the calves of the mushroom bodies (MB) and the remaining 4072 central brain (CB), which included mainly the central complex, the subesophageal zone and the peduncula of the mushroom-bodies (α and β lobes). Brains were collected one hour after the 4073 4074 retention test, which ensures that expression of all three genes was already induced (typically, from 15 to 90 min in the case of kakusei^{31, 37} and 30-60 min in the case of Hr38 and $Egr1^{28, 29}$). 4075 Two reference genes were used for the normalization (see Table 1): Efla (E=106%) and 4076 Actin $(E=110\%)^{38}$. Within-brain structure analyses showed that reference genes did not vary 4077 between learners and non-learners (t test; all comparisons NS; see Suppl Fig. 3) thus enabling 4078 4079 further comparisons between these two categories with respect to the three target IEGs. To this end, the normalization procedure used the geometric mean of the two reference genes. No cross-4080 comparisons between brain regions or genes were performed. 4081

4082 Figures 3 B-D, E-G, and H-J show the relative normalized expression of kakusei, Hr38 and Egr1, respectively, for the three brain regions considered in the case of learners and non-4083 *learners*. Significant variations of normalized expression between learners and non-learners 4084 were found for the three IEGs: in the case of kakusei and Hr38, these differences were restricted 4085 to the MBs (*kakusei:* Fig. 3C; two-sample t test; t = -2.23; df:37; p=0.03; Hr38: Fig. 3F; t = -2.23; df:37; Fig. 3F; Fig. 3 4086 2.39; df:37; p=0.02) while in the case of *Egr1*, they were observed in the optic lobes (*Egr1*: 4087 Fig. 3H; t = -2.32; df:37; p=0.03). All other within-structure comparisons between learners and 4088 non-learners were not significant (p>0.05). Notably, in the three cases in which significant 4089 variations of IEG expression were found, learners exhibited a downregulation of IEG 4090

4091 expression with respect of non-learners. In addition, from the three cases, two referred to the4092 MB calyces, which indicates the important role of this region for visual learning and memory.



Figure 3. Differential IEG expression as a consequence of associative color learning in a 4095 2D VR environment. (A) Honey bee brain with sections used for quantifying IEG 4096 4097 expression. Yellow labels indicate the brain regions used for the analysis: MB: mushroom body; CB: central brain; OL: optic lobes. The dashed lines indicate the sections performed. Ca: 4098 calyx of the mushroom body; li: lip; co: collar; \Box and \Box : \Box and \Box lobes of the mushroom 4099 body; CC: central complex; AL: antennal lobe; SEZ: subesophagic zone; OL: optic lobe; Me: 4100 4101 medulla; lo: lobula. Relative normalized expression of (B-D) kakusei, (E-G) Hr38 and (H-J) Egr1 in three main regions of the bee brain, the optic lobes (B, E, H), the calvces of the 4102 4103 mushroom bodies (C, F, I) and the central brain (D, G, J). The expression of each IEG was normalized to the geometric mean of Actin and Efla (reference genes). IEG expression was 4104

analyzed in individual brains of bees belonging to two categories: learners (L: conditioned bees 4105 that responded correctly and chose the CS+ in their first choice during the non-reinforced test) 4106 and non-learners (NL: conditioned bees that did not choose the CS+ in their first choice during 4107 the non-reinforced test). The range of ordinates was varied between panels to facilitate 4108 4109 appreciation of data scatter. In all panels, n=22 for learners and n=17 for non-learners. Different 4110 letters on top of box plots indicate significate differences (two-sample t test; p < 0.05). Box plots show the mean value in red. Error bars define the 10th and 90th percentiles. Red boxes 4111 indicate cases in which significant variations were detected. 4112

Type of gene	Target	Primer sequence 5' ≽3'	Amplicon length (bp)	E (%)	R ²
Target genes	Kakusei	CTACAACGTCCTCTTCGATT (forward)	149	96.4	0.991
		CCTACCTTGGTATTGCAGTT (reverse)			
	Hr38	TGAGATCACCTGGTTGAAAG (forward)	118	106	0.995
		CGTAGCAGGATCAATTTCCA (reverse)			
	Egrl	GAGAAACCGTTCTGCTGTGA (forward)	138	109	0.991
		GCTCTGAGGGTGATTTCTCG (reverse)			
Reference genes	Efl 🗆	AAGAGCATCAAGAGCGGAGA (forward) CACTC TTAATGACGCCCACA (reverse)	148	106	0.993
	Actin	TGCCAACACTGTCCTTTCTG (forward) AGAATTGACCCACCAATCCA (reverse)	156	110	0.995

4113	Table 1. Primer sequences used to quantify RNA expression of genes of interest and reference
4114	genes by RT-qPCR. Amplicon length (bp), efficiency (E, %) and the coefficient of correlation
4115	obtained for the standard curve (\mathbb{R}^2) are also shown. <i>Hr38</i> : Hormone receptor 38 gene; <i>Egr1</i> :
4116	Early growth response gene-1; <i>Ef1a</i> : Elongation factor 1 α gene.
4117	

4118 **Discussion**

The present work studied visual learning under a restrictive 2D VR environment and confirmed 4119 that bees can learn to discriminate visual stimuli based on their color under these artificial 4120 4121 conditions. Walking parameters did not differ between learners and non-learners so that changes in IEG expression could be ascribed to learning success. We showed that associative 4122 4123 color learning leads to a downregulation of the three IEGs considered in different areas of the visual circuit. While Egr1 was downregulated in the optic lobes, Hr38 and kakusei were 4124 4125 coincidently downregulated in the MB calvces. Our work thus reveals that the neural trace of associative color learning in the bee brain is distributed along the sequential pathway of color 4126 4127 processing and highlights the importance of MBs for color learning in bees.

4128

4129 **IEG downregulation in the bee brain**

We observed an IEG downregulation both in the optic lobes and the calyces of the MBs. This 4130 phenomenon is interesting as increased neural activity resulting from experience-dependent 4131 phenomena is usually reflected by an *upregulation* of IEG expression²⁴. Typically, upon neural 4132 responses, a relatively rapid and transient pulse of gene expression may occur, which 4133 corresponds to an experience-dependent activation of the underlying synaptic circuitry^{23, 39}. In 4134 4135 our case, however, the downregulation observed indicates that a different form of experience-4136 dependent change in neural activity occurred as a consequence of learning. A possible 4137 explanation for this phenomenon may put the accent on an inhibition of neural activity in key visual areas – optic lobes and mushroom bodies - of the learner group. 4138

In the optic lobes, *Erg1* downregulation may correspond to an increased GABAergic inhibitory activity associated with learning. The optic lobes exhibit multiple GABAergic fibers distributed principally in the medulla and the lobula⁴⁰ so that neural activity in these regions is subjected to intense GABAergic inhibitory signaling. Higher GABAergic activity has been

reported in the optic lobes of forager bees *via* quantification of *Amgad*, the honey bee homolog 4143 of the gene responsible for synthesizing the enzyme GAD⁴¹, which catalyzes the 4144 decarboxylation of glutamate to GABA. This increase was accompanied by an increase in 4145 *kakusei*⁴¹, which we did not observe. Yet, we did not study foraging behavior in a natural 4146 context, but associative learning in a controlled laboratory context. Natural foraging may 4147 involve multiple behavioral components and stimulations that may be responsible for the 4148 increase of kakusei that was absent in our study. The interesting finding is, however, that Amgad 4149 expression revealed higher GABAergic neuron activity in the optic lobes of foragers, 4150 confirming the importance of inhibitory feedback for sustaining experience-dependent visual 4151 responses. This conclusion is supported by observed increases of GABA titers in the honey bee 4152 optic lobes upon restart of foraging activities⁴² and by findings in fruit flies indicating that 4153 GABA-ergic neurons in the optic lobes are involved in tuning the sensitivity and selectivity of 4154 different visual channels^{43, 44}. 4155

In the calyces of the MBs, where coincident downregulation of kakusei and Hr38 was 4156 4157 found, neural inhibition is provided by GABAergic feedback neurons (the so-called Av3 4158 neurons)⁴⁵, which are responsible for the sparse coding responses exhibited by Kenyon cells, the constitutive neurons of the MBs. Similar GABAergic neurons exist in fruit flies, which 4159 provide inhibitory feedback to the MBs. These neurons, termed APL (anterior paired lateral) 4160 neurons, are necessary for discrimination learning of similar odorants. When flies are trained 4161 to discriminate odorants in a simple differential conditioning, disrupting the Kenyon cell-APL 4162 feedback loop decreases the sparseness of Kenyon cell odor responses, increases inter-odor 4163 correlations and prevents flies from learning to discriminate similar, but not dissimilar, odors⁴⁶. 4164 Inhibitory feedback onto the calyces of honey bees is needed for solving patterning tasks in 4165 4166 which insects have to suppress summation of responses to single elements previously rewarded when they are presented in an unrewarded compound⁴⁷ (i.e. animals have to learn to respond to 4167

the elements and not to the compound) or for reversal learning⁴⁸. A similar conclusion applies 4168 to fruit flies as GABAergic input to the MBs provided by APL neurons also mediates the 4169 capacity to solve patterning tasks⁴⁹. Increased feedback inhibition at the level of the MBs may 4170 therefore appear as a hallmark of certain learning phenomena, which require enhanced neural 4171 sparseness to decorrelate stimulus representations and thus memory specificity. In our 4172 experiments, both kakusei and Hr38 were subjected to downregulation in the MBs as a 4173 consequence of learning, a phenomenon that may be due to plastic changes in GABAergic 4174 signaling in the calvces of the MBs. Importantly, other visual areas such as the central 4175 complex⁵⁰ or the anterior optic tuberculum^{51, 52}, among others, could exhibit similar variations 4176 4177 undetectable for our methods as sectioning the frozen bee brain for molecular analyses does not allow a fine dissection of these areas. 4178

IEG downregulation is not a common phenomenon as upregulation is usually reported 4179 to indicate the presence of neural activation²². Our hypothesis on neural inhibition being the 4180 cause for this downregulation requires, therefore, to be considered with caution. Further 4181 4182 experiments are necessary to validate it, using - for instance - electrophysiological recordings 4183 in key areas of the visual circuits of learners to verify that neural activity is indeed sparser therein compared to non-learners. In addition, quantifying IEG expression in preparations in 4184 4185 which neural inhibition has been characterized extensively at the cellular level such as in the case of hippocampal and cerebellum slices exhibiting long-term depression (LTD)⁵³ could be 4186 also important. 4187

4188

4189 The neural signature of associative learning differs between different forms of VR

While the main finding in our experiments refer to a downregulation of IEG genes in key
regions of the visual circuit, our previous work using a different 3D VR system yielded a
different result²². In this 3D VR, bees could explore the virtual surroundings around the stimuli

to be learned (not bars, but cuboids that expanded upon forward movements of the bee) and could displace these stimuli laterally and in depth. They explored and learned to discriminate the color stimuli proposed to them and their learning success was comparable, yet slightly lower than that observed in the 2D VR arena (50% vs. 55%, respectively). IEG analyses comparing learners and non-learners in the 3D VR reported an *upregulation* of *Egr1* expression in the MB calyces of learners but not of non-learners. No other change was detected for *kakusei* and *Hr38* in the same three brain regions considered in the present work²².

4200 These differences are difficult to interpret as the 2D and the 3D VR experiments were not done simultaneously but in different years, though in similar seasons. In both cases, 4201 4202 motivated foragers captured at a feeder were used for the experiments. The previous visual experience of these foragers may have differed across individual and between years, thus 4203 leading to differences in performances. This explanation seems, however, rather implausible 4204 4205 given that in bees rely on the most recent appetitive learning as the one guiding predominantly 4206 actual choices. In addition, irrespective of differences in the VR environments and the resulting 4207 difference in VR immersivity, the experiments were done under similar handling conditions 4208 and using strictly the same behavioral criteria. Gene analyses were also performed under the same conditions and using the same materials and methods. Thus, the contrasting results 4209 4210 obtained in the two VR scenarios may be due to the distinct constraints they imposed to achieve discrimination learning and to the fact that the two scenarios may engage different acquisition 4211 mechanisms for learning visual information. In the 3D scenario, bees explored both the stimuli 4212 - the vertical color cuboids - and the imaginary empty surroundings; they could return to the 4213 4214 stimuli if they missed them and walk around them, which added an important exploratory component that was absent in the 2D arena. In the latter case, although bees could also bring 4215 4216 back the stimuli if they missed them by walking too fast, such a control was restricted to the frontal plane and did not allow for three-dimensional inspection. Erg1 upregulation in the 3D 4217

VR upon learning may thus reflect the convergent effects of an exploratory drive and learning in a non-constrained environment. It cannot be due to a pure exploration of the environment as non-learners exhibited the same motor performances and did not show *Egr1* upregulation.

4221 In the 2D VR, bees were forced to control tightly the lateral displacements of the stimuli - the color rectangles – without any further change allowed. This environment and task may 4222 thus impose a higher stimulus and movement control and force the bee to focus exclusively and 4223 artificially on lateral stimulus movements to gain access to sucrose reward and avoid aversive 4224 4225 saline solution. Although in both VR scenarios the background was empty and only the training stimuli were visible, the 2D VR missed the expansion of the images upon approach and thus 4226 4227 lacked of immersivity. In this context, GABA-mediated inhibition may act as a gain control mechanism that enhances response efficiency and stimulus control. In the primary visual cortex 4228 (V1) of vertebrates, GABA inhibition has been proposed to play a fundamental role in 4229 establishing selectivity for stimulus orientation and direction of motion⁵⁴⁻⁵⁶. As the latter is 4230 4231 particularly important in the 2D VR, enhanced GABA inhibition could be associated with 4232 learning to master the visual discrimination in this context.

4233 In addition, a different, yet compatible explanation for the different patterns of IEG expression found in the 3D and the 2D VR refers to a possible difference in the visual 4234 acquisition mechanisms recruited by these two scenarios. In a navigation task, body movement 4235 4236 translates into a displacement and a recognizable change in the visual scene, which can then be computed against the available internal information about the displacement⁵⁷. These 4237 pathfinding, closed-loop actions can be viewed as different from motor actions that are 4238 4239 contingent on reinforcement such as operant behaviors produced when a visual discriminative stimulus is present⁵⁸. In the latter case, vision is also engaged in discrimination learning but in 4240 a context that is not navigational. Visual learning in the 2D VR could be seen as a form of 4241 operant learning in which colors define the action to be produced to obtain the appropriate 4242

reinforcement. Thus, the observed difference in IEG expression between the two types of VRmay reflect a difference in the mechanisms used to reach the rewarded stimulus.

4245

4246 The role of mushroom bodies for visual learning and memory

Our work highlights the participation of mushroom bodies in visual learning and short-term visual retention. Numerous works have demonstrated the necessity of these brain structures for the acquisition, storage and retrieval of olfactory memories in bees^{8, 59, 60} and other insects^{2, 3,} ⁶¹. Yet, less is known about their implication in visually-driven behavioral and neural plasticity^{62, 63}. In our study, the full control over sensory stimulation offered by the VR system allowed a sound comparison between the brain of learners and non-learners, which revealed a neural signature for visual learning that included the mushroom bodies.

The implication of mushroom bodies in visual learning and memory in the bee is expected given the parallels between visual and olfactory inputs at the level of the calyces. While afferent projection neurons convey olfactory information to the lip, a subdivision of the calyces⁶⁴, afferent neurons from the lobula and the medulla, which are part of the optic lobes, convey visual information to other calyx subdivisions, the collar and the basal ring^{65, 66}. In spite of this similarity, studies addressing the role of mushroom bodies in honey bee visual learning and memory remain rare.

Bees learning to associate color lights with the presence or absence of an electric shock in a double-compartment box ^{38, 67} require the ventral lobe of the mushroom bodies to learn to avoid the punished color and spend more time in the safe color⁶⁸. In the same study, pharmacological blockade of one of the four collars (two per MB) had no effect on discrimination learning⁶⁸, which does not exclude a participation of this MB region in this visual learning given that the remaining three calyces could compensate for the loss of the blocked one. In a different study, upregulation of the dopamine receptor *Amdop1* was found in the 4268 calyces of the MBs when bees were trained to inhibit positive phototaxis towards a colored4269 light³⁸.

More recently, the implication of MBs in visual navigation was shown in wood ants 4270 Formica rufa, which are innately attracted to large visual cues (i.e. a large vertical black 4271 4272 rectangle) and which can nevertheless be trained to locate and travel to a food source placed at a specific angle away from the attractive black rectangle⁶⁹. When their MB calyces were 4273 blocked by injection of procaine^{70, 71}, ants reverted their trajectories towards the attractive 4274 4275 rectangle, which suggests a role for mushroom bodies in the dissociation between innate and learned visual responses, and in navigational memory⁶⁹. In another study involving the ant 4276 Myrmecia midas, procaine was again used to block MB function via delivery into the vertical 4277 lobes and evaluate the impact of this blockade in orientation in a familiar environment⁷². 4278 Experienced forager with procaine-inactivated VLs had tortuous paths and were unable to find 4279 their nest, whereas control ants were well directed and successful at returning home⁷². Overall, 4280 these two studies on ant navigation indicate that the vertical lobes of MBs are necessary for 4281 4282 retrieving visual memories for successful view-based navigation.

4283 Studies on the role of MBs for visual learning and memory in fruit flies have yielded contradictory findings. Mushroom body deficits do not affect learning success in the flight 4284 simulator, a setup in which tethered flies in stationary flight learn to avoid quadrants associated 4285 4286 with specific visual landmarks based on the presence of an aversive heat beam pointed towards their thorax⁷³. Similarly, learning to discriminate colors in a cylindrical container made of a 4287 blue-lit and a yellow-lit compartment, one of which was associated with aversive shaking of 4288 the flies, was not affected in mushroom body mutants⁷⁴. Spatial learning of a non-heated spot 4289 in an otherwise heated cylindrical arena displaying surrounding visual landmarks is possible in 4290 the absence of functional mushroom bodies but not of the central complex⁷⁵. Although these 4291 various results points toward a dispensability of MBs for visual learning in fruit flies⁷³, 4292

experiments comparing appetitive and aversive color learning and discrimination question this 4293 view⁷⁶. When blue and green colors were presented from below in an arena, walking flies 4294 learned both the appetitive (based on pairing one color with sugar) and the aversive 4295 discrimination (based on pairing one color with electric shock) but failed if MB function was 4296 blocked using neurogenetic tools⁷⁶. Furthermore, MBs are required for visual context 4297 generalization (e.g. generalizing landmark discrimination in a flight simulator in which 4298 contextual light was switched from blue to green between training and test)⁷⁷⁻⁷⁹. Thus, MBs 4299 4300 participate in different forms of visual learning in fruit flies, although their involvement in these phenomena seems to be less clear than in other insects. 4301

Taken together, our results revealed that learning a visual discrimination under a 2D VR, in which closed-loop conditions restricted stimulus control to lateral displacements, induced a neural signature that spanned the optic lobes and MB calyces and that was characterized by IEG downregulation, consistent with an inhibitory trace. This trace may vary and become excitatory in more permissive VR conditions in which closed-loop conditions allow for 3D exploration during discrimination learning²².

4308

4309

4310 Materials and Methods

4311 Honey bees (*Apis mellifera*) were obtained from our apiary located at the campus of the 4312 University Paul Sabatier – Toulouse III during September 2021. Only foragers caught upon 4313 landing on a gravity feeder filled with a 0.9 M sucrose solution were used in our experiments 4314 to ensure high appetitive motivation. Captured bees were enclosed in individual glass vials and 4315 then transferred to small cages housing ten bees in average; caged bees had access to *ad libitum* 4316 water and to 300 μ l of 1.5 M sucrose solution. They were kept overnight in an incubator at 28 4317 °C and 80% humidity. On the next day, they were placed on ice for five minutes to anesthetize them and facilitate the gluing of a tether to their thorax by means of melted wax (Fig. 1A). After being attached to the tether, each bee was placed on a small (5 cm diameter) Styrofoam ball for familiarization with the treadmill situation. Bees were provided with 5 μ l of 1.5 M sucrose solution and kept for 3 h in this provisory setup in the dark. They were then moved to the VR arena and used for the experiments.

Once in the VR setup, the bee was attached to a holder that allowed adjusting its position
on the treadmill (Fig. 1B), a polystyrene ball (diameter: 5 cm, weight: 1.07 g) held by a 3Dprinted support and floating on a constant airflow produced by an air pump (airflow: 555ml/s;
Aqua Oxy CWS 2000, Oase, Wasquehal, France).

4327

4328 VR setup

The VR setup consisted of the treadmill and of a half-cylindrical vertical screen made of semi-4329 transparent tracing paper, which allowed presentation of a 180° visual environment to the bee 4330 4331 (diameter: 268 mm, height: 200 mm, distance to the bee: 9 cm Fig. 1ABC) and which was placed in front of the treadmill. The visual environment was projected from behind the screen 4332 using a video projector connected to a laptop (Fig. 1A). The video projector was an Acer K135 4333 (Lamp: LED, Maximum Vertical Sync: 120 Hz, Definition: 1280 x 800, Minimum Vertical 4334 Sync: 50 Hz, Brightness: 600 lumens, Maximum Horizontal Sync: 100.10³ Hz, Contrast ratio: 4335 10 000:1, Minimum Horizontal Sync: 30.10^3 Hz)¹⁵. The movements of the walking bee on the 4336 treadmill were recorded by two infrared optic-mouse sensors (Logitech M500, 1000 dpi, 4337 Logitech, Lausanne, Switzerland) placed on the ball support perpendicular to each other. 4338

Experiments were conducted under 2D closed-loop conditions, i.e. rotations of the ball displaced the visual stimuli only laterally. To this end, we used a custom software developed using the Unity engine (version 2018.3.11f1), open-source code available at https://github.com/G-Lafon/BeeVR²¹. The software updated the position of the bee within the
VR every 0.017 s.

4344

4345 Visual stimuli

Bees had to discriminate two vertical rectangles (Fig. 1C) based on their different colors and association with reward and punishment. The colors of the rectangles (see supplementary Fig. S1) were blue (RGB: 0, 0, 255, with a dominant wavelength of 450 nm and an irradiance of 161000 μ W) and green (RGB: 0, 100, 0, with a dominant wavelength of 530 nm and an irradiance of 24370 μ W/cm2). They were displayed on a black background (RGB: 0, 0, 0). These colors were chosen based on previous work showing their successful learning in the VR setup^{15, 21}.

Each rectangle had a 5 cm base and occupied the entire vertical extent of the screen. The rectangles were positioned at -50° and $+50^{\circ}$ from the bee's body axis at the beginning of each trial (Fig. 1D, left). Keeping the object within -12.5° and $+12.5^{\circ}$ in front of the central axis of the bee continuously for 1 s was recorded as a choice (Fig. 1D, right).

4357

4358 Conditioning and testing at the treadmill

Bees were trained using a differential conditioning, which promotes better learning
performances owing to the presence of penalized incorrect color choice that result in an
enhancement of visual attention⁸⁰.

Bees were trained during 10 consecutive trials using a differential conditioning procedure (Fig. 1E) in which one of the rectangles (i.e. one of the two colors, green or blue) was rewarded with 1.5 M sucrose solution (the appetitive conditioned stimulus or CS+) while the other rectangle displaying the alternative color (the aversive conditioned stimulus or CS-) was associated with 3 M NaCl solution. The latter was used to increase the penalty of incorrect choices^{34-36, 81}. To avoid directional biases, the rewarded and the punished color rectangles were
swapped between the left and the right side of the virtual arena in a pseudo random manner
along trials.

4370 At the beginning of the experiment, bees were presented with a dark screen. During training trials, each bee faced the two rectangles (Fig. 1D, left). Choice of the CS+ rectangle 4371 was recorded if the bee kept it at the center of the screen (between -12.5° and $+12.5^{\circ}$ of the 4372 bee's central axis) during 1 s (Fig. 1D, right). Training was balanced in terms of color 4373 contingencies (i.e. blue and green equally rewarded across bees) based on a random assignment 4374 by the VR software. If the bee kept the CS+ in the center of the screen continuously during 1 s 4375 4376 (i.e. if it chose it), the screen was locked on that image for 8 s. This allowed the delivery of sucrose solution in case of a correct choice, or of NaCl in case of an incorrect choice. Solutions 4377 were delivered for 3 s by the experimenter who sat behind the bee and used a toothpick to this 4378 4379 end. The toothpick contacted first the antennae and then the mouthparts while the screen was locked on the visual image fixated by the bee. A different toothpick was used for each tastant. 4380 4381 Each training trial lasted until the bee chose one of the two stimuli or until a maximum of 60 s 4382 (no choice). Trials were separated by an inter-trial interval of 60 s during which the dark screen was presented. Bees that were unable to choose a stimulus (i.e. that did not fulfill the criterion 4383 4384 of a choice defined above) in at least 5 trials were excluded from the analysis. From 50 bees trained, 40 were kept for analysis (~80%). 4385

After the last training trial, each bee was subjected to a non-reinforced test that lasted 60 s (Fig. 1E). Test performance allowed distinguishing *learners* (i.e. bees that chose the CS+ as their first choice in the test) from *non-learners* (i.e. bees that either chose the CS- in their first test choice or that did not make any choice during the test). IEG expression was compared between these two groups, which had the same sensory experience in the VR setup and which differed only in their learning success.

4393 Brain dissection

One hour after the test, the bee was sacrificed and its head was instantly frozen in a nitrogen solution. The frozen head was dissected on dry ice under a binocular microscope. First, the antennae were removed and a window was cut in the upper part of the head capsule, removing the cuticle between the compound eyes and the ocelli. Second, the glands and tracheae around the brain were removed. Third, the retinas of the compound eyes were also removed.

The frozen brain was cut in three main parts for IEG analyses (Fig. 3A): the optic lobes (OL), the upper part of the mushroom bodies (the mushroom-body calyces, MB Ca) and the remaining central brain (CB), which included mainly the peduncula of the mushroom-bodies (α and β lobes), the central complex (CC), the antennal lobes (AL) and the subesophageal zone (SEZ). Samples were stored at -80 °C before RNA extraction. During the dissection process, one *learner* brain was lost so that learner sample sizes differ between the behavioral (n=23) and the molecular analyses (n=22).

4406

4407 **RNA extraction and reverse transcription**

The RNAs from the three sections mentioned above (OL, MB Ca and CB) were extracted using the RNeasy Micro Kit (Qiagen). The final RNA concentration obtained was measured by spectrophotometry (NanoDropTM One, Thermo Scientific). A volume of 10 μ l containing 100 ng of the RNA obtained was used for reverse transcription following the procedure recommended in the Maxima H Minus First Strand cDNA Synthesis kit (Thermo Scientific, 0.25 μ l of random hexamer primer, 1 μ l of 10 mM dNTP mix, 3.75 μ l of nuclease free H₂O, 4 μ l 5X RT Buffer and 1 μ l Maxima H Minus Enzyme Mix).

4416 **Quantitative Polymerase Chain Reaction (RT-qPCR)**

All the primers used for target and reference genes generated amplification products of approximately 150 bp. The efficiencies of all reactions with the different primers used were between 95% and 110 % (Table 1). Their specificity was verified by analyzing melting curves of the RT-qPCR products (see Supplementary Fig. S2). Two reference genes (*Ef1* α and *Actin*) were used for normalization.

4422 Expression was quantified using a SYBR Green real-time PCR method. Real-time PCR were carried out in 384-Well PCR Plates (Bio-Rad) cover with Microseal 'B' PCR Plate Sealing 4423 4424 Film (Bio-Rad). The PCR reactions were performed using the SsoAdvancedTM Universal SYBR® Green Supermix (Bio-Rad) in a final volume of 10 µl containing 5 µl of 2X 4425 SsoAdvancedTM Universal SYBR® Green Supermix, 2 µl of cDNA template (1:3 dilution 4426 from the reverse transcription reaction), 0.5 µl of 10 µmol of each primer and 2 µl of ultrapure 4427 water. The reaction conditions were as follows: 95 °C for 30 s followed by 40 cycles of 95 °C 4428 for 10 s, 55 °C for 30 s and a final step at 95 °C for 10 s followed by a melt curve from 55 °C 4429 to 95 °C with 0.5 °C per second. The reaction was performed in a CFX384 Touch Real-Time 4430 PCR Detection System (Bio-Rad) and analyzed with the software Bio-Rad CFX Manager. 4431

Each sample was run in triplicates. If the triplicates showed too much variability (SD > 4432 0.3), the furthest triplicate was discarded. If the two remaining triplicates still showed too much 4433 variability (SD > 0.3) the sample was discarded. The samples were subjected to a relative 4434 4435 quantification and normalization. First for each sample and for each reference gene per brain region, the relative quantity (Qr) was computed using the difference between the mean Ct value 4436 of each sample and the highest mean Ct value (Δ Ct), using the following formula: Qr= (1+E)^{Δ Ct} 4437 (with E= efficiency of the reaction). Then a normalization factor for each sample was obtained 4438 computing the geometric mean of the relative quantities obtained for the reference genes in the 4439 corresponding samples ($\Delta\Delta$ Ct). 4440

4442 **Data analysis and statistics**

4443 Behavioral data

The first choice of the bees was recorded during the conditioning trials and the non-reinforced test. In this way, we established for each trial and test the percentages of bees choosing first each of the stimuli displayed or not choosing a stimulus (± 95% confidence interval).

Test percentages were analyzed within groups by means of a generalized linear mixed model (GLMM) for binomial family in which the individual identity (Bee) was considered as a random factor (individual effect) while the choice category (CS+, CS-, NC) was fitted as a fixed effect; z values with corresponding degrees of freedom are reported throughout for this kind of analysis.

For the acquisition trials, we recorded motor variables such as the total distance walked during a trial, and the walking speed. The analysis of these continuous variables was done using a linear mixed model (lmer function) in which the individual identity (*Bee ID*) was a random factor and the factors *Group* (i.e. learner or non-learner) and *Trial* were fixed.

4456 Statistical analyses were performed using with R 3.5.1⁸². The package lme4 was used
4457 for GLMMs and LMMs.

4458

4459 Gene expression data

4460 Statistical differences in gene expression were assessed for reference genes to check for stability 4461 and for target genes within a given brain region using One-Factor ANOVA for independent 4462 groups in the case of multiple comparisons or two-sample T test in the case of dual comparisons. 4463 Pots hoc comparisons between groups were performed by means of a Tukey test following 4464 ANOVA. No cross-comparisons between brain regions or genes were performed due to within-
area normalization procedures. Statistical analyses were done either with R 3.5.1 software⁸² or
with Statistica 13 Software (TIBCO® Data Science).

4467

4468 Acknowledgements

We thank the valuable feedback of Profs. Takeo Kubo (University of Tokyo) and Hiroyuki Okuno (Kagoshima University, Japan) on our IEG analyses. We also thank Shiori Iino, Hiroki Kohno, Benjamin H. Paffhausen, Marco Paoli and Dorian Champelovier for valuable discussions and support, and two anonymous reviewers for useful comments on a previous version. This work was supported by an ERC Advanced Grant ('Cognibrains') to M.G, who also thanks the Institut Universitaire de France (IUF), the CNRS and the University Paul Sabatier for support.

4476

4477 **Contributions**

The project was conceived by AB, AAW, IM and MG. GL performed all the behavioral 4478 4479 experiments. HG dissected and sectioned the brains of the bees trained in the VR setup and performed all the molecular analyses. Behavioral experiments were supervised by AAW and 4480 MG. Molecular experiments were supervised by IM and MG. Statistical analyses on behavioral 4481 4482 data were performed by GL. Statistical analyses on gene-expression data were performed by HG and MG. The manuscript was written by MG and was corrected and discussed by all 4483 authors. MG obtained the funding necessary for this work. All authors reviewed and approved 4484 the final version of the manuscript. 4485

4486

4487 **Ethics declarations**

4488 Competing interests

4489 The authors declare no competing interests.

4491 **References**

- Giurfa, M., Cognition with few neurons: higher-order learning in insects. *Trends Neurosci* 2013, 36 (5), 285-294.
- 4494 2. Heisenberg, M., Mushroom body memoir: from maps to models. *Nat. Rev. Neurosci.* 2003, 4 (4),
 266-275.
- 4496 3. Cognigni, P.; Felsenberg, J.; Waddell, S., Do the right thing: neural network mechanisms of 4497 memory formation, expression and update in *Drosophila*. *Curr Opin Neurobiol* **2018**, *49*, 51-58.
- 4498 4. Benjamin, P. R.; Kemenes, G.; Kemenes, I., Non-synaptic neuronal mechanisms of learning and 4499 memory in gastropod molluscs. *Front Biosci* **2008**, *13*, 4051-7.
- 4500 5. Giurfa, M., Behavioral and neural analysis of associative learning in the honeybee: a taste from 4501 the magic well. *J. Comp. Physiol. A* **2007**, *193* (8), 801-824.
- 4502 6. Giurfa, M., Learning and cognition in insects. *Wiley Interdiscip. Rev. Cogn. Sci.* **2015**, *6* (4), 383-4503 395.
- 4504 7. Kandel, E. R., The molecular biology of memory storage: a dialogue between genes and
 4505 synapses. *Science* 2001, *294* (5544), 1030-8.
- 4506 8. Menzel, R., Memory dynamics in the honeybee. J. Comp. Physiol. A **1999**, 185, 323-340.
- 4507 9. Menzel, R., The honeybee as a model for understanding the basis of cognition. *Nature Rev*4508 *Neurosci* 2012, *13*, 758-768.
- 4509 10. Giurfa, M.; Sandoz, J. C., Invertebrate learning and memory: fifty years of olfactory conditioning
 4510 of the proboscis extension response in honeybees. *Learn Mem* **2012**, *19* (2), 54-66.
- 4511 11. Bitterman, M. E.; Menzel, R.; Fietz, A.; Schäfer, S., Classical conditioning of proboscis extension
 4512 in honeybees (*Apis mellifera*). *Journal of comparative psychology (Washington, D.C. : 1983)*4513 1983, 97 (2), 107-119.
- 4514 12. Avargues-Weber, A.; Deisig, N.; Giurfa, M., Visual cognition in social insects. *Annu Rev Entomol*4515 **2011**, *56*, 423-43.
- 4516 13. Schultheiss, P.; Buatois, A.; Avargues-Weber, A.; Giurfa, M., Using virtual reality to study visual
 4517 performances of honeybees. *Curr Opin Insect Sci* **2017**, *24*, 43-50.
- 4518 14. Frasnelli, E.; Hempel de Ibarra, N.; Stewart, F. J., The dominant role of visual motion cues in
 4519 bumblebee flight control revealed through virtual reality. *Front. Physiol.* 2018, *9*, 1038.
- 4520 15. Buatois, A.; Flumian, C.; Schultheiss, P.; Avargues-Weber, A.; Giurfa, M., Transfer of visual
 4521 learning between a virtual and a real environment in honey bees: the role of active vision. *Front.*4522 *Behav. Neurosci.* **2018**, *12*, 139.
- 4523 16. Rusch, C.; Roth, E.; Vinauger, C.; Riffell, J. A., Honeybees in a virtual reality environment learn
 4524 unique combinations of colour and shape. *J Exp Biol* 2017, *220* (Pt 19), 3478-3487.
- 4525 17. Zwaka, H.; Bartels, R.; Lehfeldt, S.; Jusyte, M.; Hantke, S.; Menzel, S.; Gora, J.; Alberdi, R.;
 4526 Menzel, R., Learning and its neural correlates in a virtual environment for honeybees. *Front.*4527 *Behav. Neurosci.* **2018**, *12*, 279.
- 452818. Rusch, C.; Alonso San Alberto, D.; Riffell, J. A., Visuo-motor feedback modulates neural activities4529in the medulla of the honeybee, Apis mellifera. J Neurosci 2021, 41 (14), 3192-3203.
- 4530 19. Buatois, A.; Pichot, C.; Schultheiss, P.; Sandoz, J. C.; Lazzari, C. R.; Chittka, L.; Avargues4531 Weber, A.; Giurfa, M., Associative visual learning by tethered bees in a controlled visual
 4532 environment. *Sci. Rep.* 2017, 7 (1), 12903.
- 4533 20. Buatois, A.; Laroche, L.; Lafon, G.; Avargues-Weber, A.; Giurfa, M., Higher-order discrimination 4534 learning by honeybees in a virtual environment. *Eur J Neurosci* **2020**, *51* (2), 681-694.
- Lafon, G.; Howard, S. R.; Paffhausen, B. H.; Avarguès-Weber, A.; Giurfa, M., Motion cues from
 the background influence associative color learning of honey bees in a virtual-reality scenario. *Sci. Rep.* 2021, *11* (1), 21127.
- 4538 22. Geng, H.; Lafon, G.; Avarguès-Weber, A.; Buatois, A.; Massou, I.; Giurfa, M., Visual learning in a
 4539 virtual reality environment upregulates immediate early gene expression in the mushroom
 4540 bodies of honey bees. *Commun Biol* 2022, (in press).

- 4541 23. Clayton, D. F., The genomic action potential. *Neurobiol Learn Mem* **2000**, *74* (3), 185-216.
- 4542 24. Bahrami, S.; Drabløs, F., Gene regulation in the immediate-early response process. *Advances in*4543 *Biological Regulation* 2016, *62*, 37-49.
- 4544 25. Minatohara, K.; Akiyoshi, M.; Okuno, H., Role of Immediate-Early Genes in Synaptic Plasticity
 4545 and Neuronal Ensembles Underlying the Memory Trace. *Front. Mol. Neurosci.* 2015, *8* (78), 78.
- 4546 26. Kiya, T.; Kubo, T., Dance type and flight parameters are associated with different mushroom
 4547 body neural activities in worker honeybee brains. *PLoS One* **2011**, *6* (4), e19301.
- Singh, A. S.; Shah, A.; Brockmann, A., Honey bee foraging induces upregulation of early growth
 response protein 1, hormone receptor 38 and candidate downstream genes of the ecdysteroid
 signalling pathway. *Insect Mol Biol* **2018**, *27* (1), 90-98.
- 4551 28. lino, S.; Shiota, Y.; Nishimura, M.; Asada, S.; Ono, M.; Kubo, T., Neural activity mapping of
 4552 bumble bee (*Bombus ignitus*) brains during foraging flight using immediate early genes. *Sci. Rep.*4553 2020, 10 (1), 7887.
- Ugajin, A.; Uchiyama, H.; Miyata, T.; Sasaki, T.; Yajima, S.; Ono, M., Identification and initial
 characterization of novel neural immediate early genes possibly differentially contributing to
 foraging-related learning and memory processes in the honeybee. *Insect Mol Biol* 2018, 27 (2),
 154-165.
- 30. Shah, A.; Jain, R.; Brockmann, A., Egr-1: A candidate transcription factor involved in molecular
 processes underlying time-memory. *Front. Psychol.* **2018**, *9* (865), 865.
- 4560 31. Kiya, T.; Kunieda, T.; Kubo, T., Increased neural activity of a mushroom body neuron subtype in
 4561 the brains of forager honeybees. *PLoS One* **2007**, *2* (4), e371.
- 4562 32. Fujita, N.; Nagata, Y.; Nishiuchi, T.; Sato, M.; Iwami, M.; Kiya, T., Visualization of neural activity 4563 in insect brains using a conserved immediate early gene, Hr38. *Curr Biol* **2013**, *23* (20), 2063-70.
- 4564 33. Duclot, F.; Kabbaj, M., The Role of Early Growth Response 1 (EGR1) in Brain Plasticity and
 4565 Neuropsychiatric Disorders. *Front. Behav. Neurosci.* 2017, *11* (35), 35.
- 4566 34. de Brito Sanchez, M. G.; Serre, M.; Avarguès-Weber, A.; Dyer, A. G.; Giurfa, M., Learning 4567 context modulates aversive taste strength in honey bees. *J Exp Biol* **2015**, *218* (Pt 6), 949-59.
- 4568 35. Aguiar, J.; Roselino, A. C.; Sazima, M.; Giurfa, M., Can honey bees discriminate between floral-4569 fragrance isomers? *J Exp Biol* **2018**, *221* (Pt 14).
- 4570 36. Ayestaran, A.; Giurfa, M.; de Brito Sanchez, M. G., Toxic but drank: gustatory aversive
 4571 compounds induce post-ingestional malaise in harnessed honeybees. *PLoS One* 2010, *5* (10),
 4572 e15000.
- 4573 37. Ugajin, A.; Kiya, T.; Kunieda, T.; Ono, M.; Yoshida, T.; Kubo, T., Detection of neural activity in
 4574 the brains of Japanese honeybee workers during the formation of a "hot defensive bee ball".
 4575 *PLoS One* **2012**, *7* (3), e32902.
- 4576 38. Marchal, P.; Villar, M. E.; Geng, H.; Arrufat, P.; Combe, M.; Viola, H.; Massou, I.; Giurfa, M.,
 4577 Inhibitory learning of phototaxis by honeybees in a passive-avoidance task. *Learn Mem* 2019, *26*4578 (10), 412-423.
- 4579 39. Okuno, H., Regulation and function of immediate-early genes in the brain: beyond neuronal
 4580 activity markers. *Neurosci Res* 2011, *69* (3), 175-186.
- 40. Schäfer, S.; Bicker, G., Distribution of GABA-like immunoreactivity in the brain of the honeybee. J
 4582 Comp Neurol 1986, 246 (3), 287-300.
- 4583 41. Kiya, T.; Kubo, T., Analysis of GABAergic and non-GABAergic neuron activity in the optic lobes of 4584 the forager and re-orienting worker honeybee (*Apis mellifera* L.). *PLoS One* **2010**, *5* (1), e8833.
- 4585 42. Chatterjee, A.; Bais, D.; Brockmann, A.; Ramesh, D., Search behavior of individual foragers 4586 involves neurotransmitter systems characteristic for social scouting. *Front Insect Sci* **2021**, *1* (4).
- 4587 43. Keles, M. F.; Hardcastle, B. J.; Stadele, C.; Xiao, Q.; Frye, M. A., Inhibitory interactions and
 4588 columnar inputs to an object motion detector in *Drosophila*. *Cell Rep* 2020, *30* (7), 2115-2124 e5.
- 4589 44. Keles, M. F.; Frye, M. A., Object-detecting neurons in *Drosophila*. *Curr Biol* **2017**, *27* (5), 680-687.
- 4590 45. Rybak, J.; Menzel, R., Anatomy of the mushroom bodies in the honey bee brain: The neuronal connections of the alpha-lobe. *J Comp Neurobiol* **1993**, *334*, 444-465.

- 46. Lin, A. C.; Bygrave, A. M.; de Calignon, A.; Lee, T.; Miesenbock, G., Sparse, decorrelated odor
 coding in the mushroom body enhances learned odor discrimination. *Nat Neurosci* 2014, *17* (4),
 559-68.
- 47. Devaud, J. M.; Papouin, T.; Carcaud, J.; Sandoz, J. C.; Grunewald, B.; Giurfa, M., Neural
 4596 substrate for higher-order learning in an insect: Mushroom bodies are necessary for configural
 4597 discriminations. *Proc Natl Acad Sci U S A* **2015**, *112* (43), E5854-62.
- 4598 48. Boitard, C.; Devaud, J.-M.; Isabel, G.; Giurfa, M., GABAergic feedback signaling into the calyces
 4599 of the mushroom bodies enables olfactory reversal learning in honey bees. *Front. Behav.*4600 *Neurosci.* 2015, *9*.
- 4601 49. Durrieu, M.; Wystrach, A.; Arrufat, P.; Giurfa, M.; Isabel, G., Fruit flies can learn non-elemental
 4602 olfactory discriminations. *Proceedings of the Royal Society B: Biological Sciences* 2020, *287*4603 (1938), 20201234.
- 4604 50. Honkanen, A.; Adden, A.; da Silva Freitas, J.; Heinze, S., The insect central complex and the 4605 neural basis of navigational strategies. *J Exp Biol* **2019**, *222* (Pt Suppl 1), jeb188854.
- 4606 51. Mota, T.; Yamagata, N.; Giurfa, M.; Gronenberg, W.; Sandoz, J. C., Neural organization and
 4607 visual processing in the anterior optic tubercle of the honeybee brain. *J Neurosci* 2011, *31* (32),
 4608 11443-11456.
- 4609 52. Mota, T.; Gronenberg, W.; Giurfa, M.; Sandoz, J. C., Chromatic processing in the anterior optic 4610 tubercle of the honeybee brain. *J Neurosci* **2013**, *33*, 4-16.
- 4611 53. Massey, P. V.; Bashir, Z. I., Long-term depression: multiple forms and implications for brain
 4612 function. *TINS* 2007, *30* (4), 176-184.
- 4613 54. Rose, D.; Blakemore, C., Effects of bicuculline on functions of inhibition in visual cortex. *Nature*4614 **1974**, *249* (455), 375-7.
- 55. Sillito, A. M., Inhibitory mechanisms influencing complex cell orientation selectivity and their
 modification at high resting discharge levels. J. Physiol. **1979**, 289, 33-53.
- 56. Tsumoto, T.; Eckart, W.; Creutzfeldt, O. D., Modification of orientation sensitivity of cat visual
 cortex neurons by removal of GABA-mediated inhibition. *Exp Brain Res* 1979, *34* (2), 351-63.
- 4619 57. von Holst, E.; Mittelstaedt, H., Das Reafferenzprinzip. *Naturwissenschaften* 1950, *37* (20), 4644620 476.
- 4621 58. Skinner, B. F., *The Behavior of Organisms. An Experimental Analysis*. Appleton-Century-Crofts:
 4622 New York, 1938; p 457.
- 4623 59. Menzel, R., The insect mushroom body, an experience-dependent recoding device. *J Physiol*4624 *Paris* 2014, *108* (2-3), 84-95.
- 4625 60. Devaud, J. M.; Papouin, T.; Carcaud, J.; Sandoz, J. C.; Grünewald, B.; Giurfa, M., Neural
 4626 substrate for higher-order learning in an insect: Mushroom bodies are necessary for configural
 4627 discriminations. *Proc Natl Acad Sci U S A* **2015**, *112* (43), E5854-62.
- 4628 61. Guven-Ozkan, T.; Davis, R. L., Functional neuroanatomy of *Drosophila* olfactory memory
 4629 formation. *Learn Mem* 2014, *21* (10), 519-26.
- 4630 62. Avargues-Weber, A.; Mota, T., Advances and limitations of visual conditioning protocols in
 4631 harnessed bees. *J Physiol Paris* **2016**, *110* (3 Pt A), 107-118.
- 4632 63. Avarguès-Weber, A.; Mota, T.; Giurfa, M., New vistas on honey bee vision. *Apidologie* 2012, 14633 25.
- 4634 64. Kirschner, S.; Kleineidam, C. J.; Zube, C.; Rybak, J.; Grünewald, B.; Róssler, W., Dual olfactory
 4635 pathway in the honeybee, *Apis mellifera*. *J Comp Neurol* **2006**, *499* (6), 933-952.
- 463665. Ehmer, B.; Gronenberg, W., Segregation of visual input to the mushroom bodies in the4637honeybee (*Apis mellifera*). J Comp Neurol **2002**, 451 (4), 362-73.
- 4638 66. Paulk, A. C.; Dacks, A. M.; Phillips-Portillo, J.; Fellous, J. M.; Gronenberg, W., Visual processing
 4639 in the central bee brain. *J Neurosci* 2009, *29* (32), 9987-99.
- 4640 67. Kirkerud, N. H.; Schlegel, U.; Giovanni Galizia, C., Aversive learning of colored lights in walking
 4641 honeybees. *Front. Behav. Neurosci.* 2017, 11, 94.

- 4642 68. Plath, J. A.; Entler, B. V.; Kirkerud, N. H.; Schlegel, U.; Galizia, C. G.; Barron, A. B., Different
 4643 roles for honey bee mushroom bodies and central complex in visual learning of colored lights in
 4644 an aversive conditioning assay. *Front. Behav. Neurosci.* 2017, *11*, 98.
- 69. Buehlmann, C.; Wozniak, B.; Goulard, R.; Webb, B.; Graham, P.; Niven, J. E., Mushroom Bodies
 Are Required for Learned Visual Navigation, but Not for Innate Visual Behavior, in Ants. *Curr Biol*2020, *30* (17), 3438-3443 e2.
- 4648
 70. Devaud, J. M.; Blunk, A.; Podufall, J.; Giurfa, M.; Grünewald, B., Using local anaesthetics to
 block neuronal activity and map specific learning tasks to the mushroom bodies of an insect
 brain. *Eur J Neurosci* 2007, *26* (11), 3193-206.
- 4651 71. Muller, D.; Staffelt, D.; Fiala, A.; Menzel, R., Procaine impairs learning and memory
 4652 consolidation in the honeybee. *Brain Res* 2003, *977* (1), 124-127.
- Kamhi, J. F.; Barron, A. B.; Narendra, A., Vertical Lobes of the Mushroom Bodies Are Essential
 for View-Based Navigation in Australian Myrmecia Ants. *Curr Biol* **2020**, *30* (17), 3432-3437 e3.
- 4655 73. Wolf, R.; Wittig, T.; Liu, L.; Wustmann, G.; Eyding, D.; Heisenberg, M., *Drosophila* mushroom
 4656 bodies are dispensable for visual, tactile, and motor learning. *Learn Mem* **1998**, *5* (1-2), 166-78.
- 4657 74. Heisenberg, M.; Borst, A.; Wagner, S.; Byers, D., *Drosophila* mushroom body mutants are
 4658 deficient in olfactory learning. *J Neurogenet* **1985**, *2* (1), 1-30.
- 4659 75. Ofstad, T. A.; Zuker, C. S.; Reiser, M. B., Visual place learning in *Drosophila melanogaster*.
 4660 *Nature* 2011, 474 (7350), 204-U240.
- Vogt, K.; Schnaitmann, C.; Dylla, K. V.; Knapek, S.; Aso, Y.; Rubin, G. M.; Tanimoto, H., Shared
 mushroom body circuits underlie visual and olfactory memories in *Drosophila*. *Elife* 2014, *3*,
 e02395.
- 4664 77. Liu, L.; Wolf, R.; Ernst, R.; Heisenberg, M., Context generalization in Drosophila visual learning
 4665 requires the mushroom bodies. *Nature* **1999**, *400*, 753-756.
- 4666 78. Brembs, B.; Wiener, J., Context and occasion setting in Drosophila visual learning. *Learn. Mem.*4667 **2006**, *13* (5), 618-628.
- Tang, S.; Guo, A., Choice behavior of *Drosophila* facing contradictory visual cues. *Science* 2001, 294, 1543-1547.
- 4670 80. Avarguès-Weber, A.; Giurfa, M., Cognitive components of color vision in honey bees: how
 4671 conditioning variables modulate color learning and discrimination. *J. Comp. Physiol. A* 2014, 200
 4672 (6), 449-461.
- 4673 81. Bestea, L.; Rejaud, A.; Sandoz, J. C.; Carcaud, J.; Giurfa, M.; de Brito Sanchez, M. G., Peripheral
 4674 taste detection in honey bees: What do taste receptors respond to? *Eur J Neurosci* 2021, *54* (2),
 4675 4417-4444.
- 4676 82. R Development Core Team, *R: A Language and Environment for Statistical Computing*. The R
 4677 Foundation for Statistical Computing: Vienna, Austria, 2016.
- 4678



4682 Supplementary Figure 1. Spectral distribution (relative intensity as a function of wavelength)
4683 of the blue light (dominant wavelength 446 nm) and the green light (dominant wavelength 528
4684 nm) used to train the bees in the color discrimination task.



4687 Supplementary Figure 2. Validation selectivity of gene-specific primers. Melting peaks of
4688 RT-qPCR. A) Reference genes. B) Target genes.



4691 **Supplementary Figure 3. Expression levels (Cq values) of the reference genes** *Actin* (upper 4692 row) and *Ef1* (lower row). Expression levels in the brain regions considered (optic lobes, 4693 mushroom body calyces and central brain) of learners and non-learners (n=22 and n=17, 4694 respectively, for both genes). Box plots show the mean value in yellow (*Actin*) or red (*Ef1*). 4695 Sample sizes are indicated within parentheses below each group. Error bars define the 10th and 4696 90th percentiles. Same letters on top of box plots indicate absence of significant differences 4697 (two-sample t test; p < 0.05).

4698

4700	Chapter 4
4701	Comparison of associative visual learning in a 3D virtual reality between
4702	bumblebees and honeybees



4705 Comparison of associative visual learning in a 3D virtual reality between 4706 bumblebees and honey bees.

4707 Gregory Lafon¹, Aurore Avarguès-Weber¹, Martin Giurfa^{1,2,3}

¹Research Centre on Animal Cognition, Center for Integrative Biology, CNRS, University of Toulouse,
118 route de Narbonne, F-31062 Toulouse cedex 09, France.

² College of Animal Sciences (College of Bee Science), Fujian Agriculture and Forestry University,
Fuzhou 350002, China.

4712 ³ Institut Universitaire de France, Paris, France (IUF).

4713

In chapter 2 and 3 we found that color discrimination learning induced a neural signature 4714 distributed along the sequential pathway of color processing by performing ex vivo analysis of 4715 the brains of learner and non-learner bees. To push our investigations further, we believe it 4716 would be interesting to use our VR setup for in vivo analysis. Preliminary experiments revealed 4717 4718 that honey bees engage less with the VR after brain surgery. In order to overcome this 4719 limitation, we decided to investigate the possibility of using bumble bee in VR experiment. Bumble bees are closely related to honey bees and known for being more resilient to surgery, 4720 4721 thus providing a good alternative model for our experiments. By conditioning bumble bees in VR, we found that bumble bees are able to solve the color discrimination task with a success 4722 rate comparable to honey bee. We also found that they engage more and make more choices 4723 than honey bee, which leads to higher amount of useable data. 4724

- 4727 between bumblebees and honeybees.
- 4728

4729 Gregory Lafon¹, Aurore Avarguès-Weber¹, Martin Giurfa^{1,2,3}

- 4730
- 4731 ¹Research Centre on Animal Cognition, Center for Integrative Biology, CNRS, University of Toulouse,
- 4732 118 route de Narbonne, F-31062 Toulouse cedex 09, France.
- ² College of Animal Sciences (College of Bee Science), Fujian Agriculture and Forestry University,
 Fuzhou 350002, China.
- 4735 ³ Institut Universitaire de France, Paris, France (IUF).
- 4736
- 4737 Corresponding author: Dr. Martin Giurfa
- 4738 Research Centre on Animal Cognition, CNRS UPS, 31062 Toulouse cedex 9, France
- 4739 <u>martin.giurfa@univ-tlse3.fr</u>
- 4740

4741 Introduction

Among insects, honeybees represent a valuable model system for cognitive research. Thanks to a rich behavioral repertoire supported by a small brain, they allow easy access to the neural correlate of cognitive processes like learning and memory (Giurfa, 2007). The last decades have also seen the emergence of bumblebees as a model in cognition (Real, 1992; Dyer and Chittka, 2004; Worden et al., 2005; Kulahci et al., 2008)

4747 Bumblebees are eusocial central place forager that despite being closely related to honeybees 4748 differ in a few important points (A. J. Riveros and Gronenberg, 2009). Labor division in bumblebees is strongly influenced by size, with larger bees assuming the role of foragers while 4749 4750 smaller worker stay in the hives (Goulson, 2003). Bumblebees' colonies are smaller than honeybees' with only between 200-400 worker instead of the tens of thousands typically found 4751 in honeybees' hives. This contributes to bumblebees being more easily reared under laboratory 4752 conditions (A. J. Riveros and Gronenberg, 2009). Bumblebees are solitary foragers, unlike 4753 4754 honeybees they do not share intentionally information about location and quality of potential 4755 resources with their nest mates (Leadbeater and Chittka, 2005; Worden et al., 2005; Leadbeater and Chittka, 2007) which makes them an interesting model to study foraging strategies 4756 (Lihoreau et al., 2013). 4757

Related to their central place forager ecology, vision plays a central role in major aspects of bumblebees' life histories, from navigation (Church and Plowright, 2006; Saleh and Chittka, 2007) to floral selection (Dukas and Waser, 1994; Laverty, 1994; Cnaani et al., 2006). In the past decades, bumblebees have been extensively used to investigate cognitive processes under free flight conditions (Heinrich et al., 1977; Real, 1992; Keasar et al., 1996; Chittka and Thomson, 1997; Worden et al., 2005; Cnaani et al., 2006; Kulahci et al., 2008; A. J. Riveros and Gronenberg, 2009; Leonard et al., 2011; Mertes et al., 2014; Foster et al., 2014; Robert et

al., 2018; Li et al., 2018; Frasnelli et al., 2021). Bumblebees were also tested in harnessed 4765 conditions with olfactory conditioning of the PER (Riddell and Mallon, 2006; Andre J. Riveros 4766 and Gronenberg, 2009; A. J. Riveros and Gronenberg, 2009; Toda et al., 2009; Anfora et al., 4767 4768 2011). Initially developed for honeybees (Bitterman et al., 1983), it allows a deep control of the different experimental parameters and makes it possible to couple learning paradigms with 4769 invasive neurobiological techniques (Giurfa and Sandoz, 2012). More recently, visual versions 4770 4771 of the PER conditioning have also been proposed (Riveros and Gronenberg, 2012; Lichtenstein et al., 2015; Muth et al., 2018; Riveros et al., 2020). Not only can bumblebees successfully learn 4772 in harnessed condition but they are also robust and reliable during electrophysiology recordings 4773 4774 (Skorupski et al., 2007; Spaethe et al., 2007; Paulk et al., 2008; Paulk and Gronenberg, 2008; Skorupski and Chittka, 2010a, 2010b; Vähäkainu et al., 2013; Rusanen et al., 2017) or calcium 4775 4776 brain imaging (Mertes et al., 2021).

We have recently developed a Virtual reality (VR) setup that allows to successfully train 4777 restrained honeybees in a visual differential conditioning task (Buatois et al., 2017; Lafon et 4778 4779 al., 2021). Such a setup is intended to open up further possibilities to explore the underlying mechanisms of visual learning by facilitating a live access to the brain of a behaving insect. In 4780 4781 that context, bumblebees appear like a very interesting model since they display some of the 4782 rich behavioral repertoire of the honeybees but with a bigger brain and a more robust body which make them more compatible with invasive protocols like electrophysiology and calcium 4783 imaging. 4784

Here we assessed if and how the bumblebee *Bombus terrestris* can learn a visual discrimination task under VR conditions. We measured bumblebee's performances during the learning phase and a subsequent non-reinforced test and compared them with honeybees conditioned with the same protocol. We also performed two conditioning experiments using either NaCl or Quinine solutions as a punishment, to find the best way to negatively reinforce a stimulus. Since size 4790 plays an important role in bumblebees' labor division (Goulson, 2003), we could expect larger 4791 bumblebees, that are usually foragers, to perform better in the color discrimination task than 4792 smaller workers. Moreover, our 3D VR setup involves a significant motor component as bees 4793 have to move in the virtual environment to reach the rewarded stimuli, so we hypothesize that 4794 the insect's strength could impact bee's performances.

4795 Materials and methods

4796 Study species and collection

Bumblebees were collected each morning around 9 am from twenty commercial colonies of *B*. *terrestris*, sixteen for experiment 2, during July 2021, and four for experiment 3, during
November 2021 (Koppert, Cavaillon, France) by placing a glass vial at the entrance of the box
and collecting the workers that came out into the vial. Each colony contained about 200
workers, brood, and 1 queen. Bumblebees were maintained and tested in the laboratory at 25°C
and 30–40% relative humidity, under a 12:12 h light:dark photocycle.

Honey bee foragers (*Apis mellifera*) were obtained from the CRCA apiary located in the campus of the University Paul Sabatier during July 2021. Foragers were captured the day before the experiment at gravity feeders providing 0.88 M sucrose solution upon landing and before they began feeding. Captured bees were enclosed in individual glass vials and then transferred to small cages housing ten bees in average; where they had access to *ad libitum* water and 300 μ l of 1.5 M sucrose solution. They were then kept overnight in an incubator at 28°C and 80% humidity.

Bees, bumblebees and honeybees, were cooled on ice for 5 minutes to anesthetize them and attach them to their tether. Bees were handled under red light, which ensured a dark environment to the insects.

4813 Tethering procedure

The tethering procedure was the same for both honeybees and bumblebees, referred to as bees
in the following paragraphs. The procedure followed the same protocol as described in chapter
1 (Lafon et al., 2021).

Each bee was tethered by means of a 0.06 g steel needle, 0.5 mm in diameter and 40 mm in 4817 4818 length, which was fixed to the thorax by melted beeswax. The needle was placed within a 3D printed resin tube (Black tough resin Prusa, Prusa Research a. s., Czech republic), 7 mm inner 4819 diameter, 1 cm outer diameter and 55 mm in length, which was fixed on a holding frame placed 4820 above the treadmill (Fig.1 A-B). This system allowed the bee to adjust its position in the vertical 4821 axis once set on the ball, but did not allow rotational movements. The holding frame consisted 4822 4823 of a vertical black, plastic half frame made of two vertical rectangular supports, 105 mm in 4824 length, connected to an upper, horizontal rectangular support, 120 mm in length. The latter held the black cylinder in the middle (Fig. 1B). After being attached to its tether, each bee was placed 4825 on a small (50 mm diameter) Styrofoam ball for familiarization to a provisory set-up and 4826 4827 provided with 5 µl of 1.5 M sucrose solution. Each bee was held for 3 h in this provisory setup, which was kept in the dark and without visual stimulations. 4828



4831 Figure 1. Experimental setup and 3D environment. A) Global view of the VR system. 1: Semicircular projection screen made of tracing paper. 2: Holding frame to place the tethered bee on the treadmill. 4832 4833 3: The treadmill was a Styrofoam ball positioned within a cylindrical support (not visible) floating on 4834 an air cushion. 4: Infrared mouse optic sensors allowing to record the displacement of the ball and to 4835 reconstruct the bee's trajectory. 5: Air arrival. The video projector displaying images on the screen 4836 from behind can be seen on top of the image. B) The tethering system. 1: Plastic cylinder held by the 4837 holding frame; the cylinder contained a glass cannula into which a steel needle was inserted. 2: The 4838 needle was attached to the thorax of the bee. 3: Its curved end was fixed to the thorax by means of 4839 melted bee wax. C) Color discrimination learning in the VR setup. The bee had to learn to discriminate 4840 two vertical cylinders based on their different color and their association with reward and punishment. 4841 Cylinders were green and blue on a dark background. Color intensities were adjusted to avoid phototactic biases independent of learning. 4842

4843

4844 Virtual reality set-up

The bee was then moved to the VR setup (Lafon et al., 2021). The VR relayed on a custom software developed using the Unity engine (version 2020.3.4f1), open-source code available at https://github.com/G-Lafon/BeeVR. The software updated the position of the bee within the VR every 0.017 s.

The VR apparatus consisted of a spherical Styrofoam ball (diameter: 50 mm, weight 1.07 g), which acted as a treadmill. The ball was positioned within a 3D-printed, hollow, cylindrical support (cylinder: 50 mm high, 59 mm diameter). The cylinder allowed distributing an upwards air flow of 33 L.min⁻¹ produced by an AquaOxy 2000 aquarium pump, and released through a small hole at the base of the cylindrical support at a pressure of 1.221 bar. The movements of the ball were recorded by two infrared optic-mouse sensors (Logitech M500, 1000 dpi) placed
at a distance of 7 mm from the sphere and forming an angle of 90° angle relative to each other
(i.e. 45° from the bee body axis).

- 4857 The ball was positioned in front of a half-cylindrical vertical screen, 268 mm in diameter and 200 mm height, which was placed at 9 cm from the bee. The screen was made of semi-4858 transparent tracing paper, which allowed presentation of a 180° visual environment to the bee. 4859 The visual environment was projected from behind the screen using a video projector connected 4860 to a laptop. The video projector was an Acer K135 (Lamp: LED, Maximum Vertical Sync: 120 4861 Hz, Definition: 1280 x 800, Minimum Vertical Sync: 50 Hz, Brightness: 600 lumens, Maximum 4862 Horizontal Sync: 100.10³ Hz, Contrast ratio: 10 000:1, Minimum Horizontal Sync: 30.10³ Hz). 4863 The lag between the motion of the bee and the update of the visual surrounding was of $18.00 \pm$ 4864 2.53 ms (mean \pm S.E.; n = 10) (Lafon et al., 2021). 4865
- Experiment 1: Establishing a balanced pair of green and blue for bumblebee conditioning In order to find a pair of colors that elicited the same amount of attraction we recorded the spontaneous choice of bumblebee presented with two pairs of green and blue stimulus. One pair called Green (R: 0, G:100, B:1.0; irradiance 24 370 μ W/cm²) versus Bright Blue (R: 0, G:80, B:254) and a second pair called Dark Green (R: 0, G:51,B:1.0) versus Blue (R: 0, G:0, B:255; irradiance 161 000 μ W/cm²).
- Each cylinder had a 5 cm diameter base and 1 m height so that it occupied the entire vertical extent of the screen irrespective of the bumblebee's position. At the beginning of each test, it subtended a horizontal visual angle of 6.5° and was positioned either to the left (-50°) or the right ($+50^{\circ}$) of the tethered insect. Approaching the cylinder resulted in an expansion of its horizontal extent (1.7° /cm). A choice was recorded when the bumblebee approached the cylinder within an area of 3 cm surrounding its virtual surface and directly faced its center (Fig. 2A).

4879 Each insect was subjected to two consecutive tests in non-reinforced conditions. One test for
4880 each pair of colors, the order of presentation of the pairs was random. We recorded what color
4881 the bumblebee chose first. Each test lasted 180 s and the inter-test interval was 10 s.



4884 Figure 2. Choice criterion and conditioning protocol for color discrimination learning. A) Choice 4885 criterion. Left: A bee facing the two virtual cylinders. Center: A bee approaching a target cylinder; the 4886 cylinder has not yet been centered by the bee (gray area). Right: A bee having centered the target 4887 cylinder (gray area). A choice was recorded when the bee reached an area of a radius of 3 cm centered 4888 on the cylinder and fixed it frontally. The cylinder image was then frozen during 8 s for honeybees, or 4889 14 s for bumblebees, and the corresponding reinforcement (US) was delivered. B) Conditioning 4890 protocol. Bees were trained along 6 conditioning trials that lasted a maximum of 3 min and that were 4891 spaced by 1 min (inter-trial interval). After the end of conditioning, and following an additional interval 4892 of 1 min, bees were tested in extinction conditions with the two colored cylinders during 3 min.

Having chosen a pair of colors that elicited the same amount of spontaneous attraction in 4895 bumblebees (see above), we trained bumblebees and honeybees, to discriminate between two 4896 vertical colored cylinders, one rewarded and the other not (see Training and testing procedure 4897 4898 below). Both cylinders had the same dimensions of the cylinder employed in the previous experiment. For honeybees, one was blue (RGB: 0, 0, 255, with a dominant wavelength of 446 4899 4900 nm) and the other green (RGB: 0, 51, 0, with a dominant wavelength at 528 nm). Their intensity, 4901 measured at the level of the bee eye, was 161 000 μ W/cm² (blue cylinder) and 24 370 μ W/cm² 4902 (green cylinder). These values were shown to elicit the same level of spontaneous attraction 4903 (Buatois et al., 2017; Lafon et al., 2021). For bumblebees we used the Dark Green (R: 0, G:51, 4904 B:1.0) and Blue pair (RGB: 0, 0, 255, with a dominant wavelength of 446 nm) (see results from experiment 1). 4905

The cylinders were positioned respectively at -50° and $+50^{\circ}$ from the bee's body axis at the beginning of each trial. As in the previous experiment, approaching a cylinder within an area of 3 cm surrounding its virtual surface followed by direct fixation of its center was recorded as a choice (Fig. 2A).

4910 Experiment 3: Conditioning bumblebees with Quinine as punishment

In this experiment we repeated the protocol from the experiment 2 on bumblebees using 1.2g.L⁻
¹ quinine solution (Dyer and Chittka, 2004) instead of 3 M NaCl as punishment. All other
parameters were the same.

4914 Training and testing procedure

Both honeybees and bumblebees, were trained during 6 trials using a differential conditioning
procedure (Fig. 2B) in which one of the cylinders (i.e. one of the two colors, green or blue) was
rewarded with 1.5 M sucrose solution (the appetitive conditioned stimulus or CS+) while the

4918 other cylinder displaying the alternative color (the aversive conditioned stimulus or CS-) was 4919 associated with either 3 M NaCl solution (experiment 2) or 1.2 g.L^{-1} quinine solution 4920 (experiment 3).

4921 At the beginning of the experiment, bees were presented with a dark screen for 60 s. During 4922 training trials, each bee faced the virtual environment with the two cylinders in front of it. The 4923 bee had to learn to choose the CS+ cylinder by walking towards it and centering it on the screen. 4924 Training was balanced in terms of color contingencies (i.e. blue and green equally rewarded across bees) based on a random assignment by the VR software. If the bee reached the CS+ 4925 4926 within an area of 3 cm in the virtual environment (i.e. the chosen cylinder subtended a horizontal 4927 visual angle of 53°) and centered, the screen was locked on that image for 8s for honeybees (Lafon et al., 2021) or 14 s in case of bumblebees. The screen freezing was longer for 4928 bumblebees as we found in pre-experiments that they need more time to take the reward from 4929 the toothpick. This allowed the delivery of sucrose solution in case of a correct choice, or of 4930 NaCl (experiment 2) or quinine (experiment 3) in case of an incorrect choice. Solutions were 4931 4932 delivered for 3 s by the experimenter who sat behind the bee and used a toothpick to this end. The toothpick contacted first the antennae and then the mouthparts while the screen was locked 4933 on the visual image fixated by the bee. 4934

4935 Each training trial lasted until the bee chose one stimuli or until a maximum of 180 s (no choice). Thus, a single choice (or a no choice) was recorded during each training trial. Trials were 4936 4937 separated by an inter-trial interval of 60 s during which the dark screen was presented. The bees that were unable to choose a stimulus in at least 3 trials were excluded from the analysis. From 4938 138 bumblebees trained in experiment 2, 123 were kept for analysis (~89%). From 77 4939 4940 honeybees trained, 31 were kept for analysis (~40%). In the experiment 3, out of 315 trained bumblebees, 235 were kept for analysis (~75%). Every animal was frozen at -20°C after the 4941 experiment to be later weighed and measured. 4942

After the last training trial, each bee was subjected to a non-reinforced test that contrary to 4943 4944 training trials had a fixed duration of 180 s. During this test, two variables were recorded: the first choice (as defined above) and the time spent fixating the rewarded and the non-rewarded 4945 4946 stimulus. Both variables have been used in prior works performed in our VR setup to characterize test performances as they may reveal different aspects of behavioral performances 4947 (Buatois et al., 2020, 2018, 2017; Lafon et al., 2021). Fixation time (s) was defined as the time 4948 4949 spent by each cylinder at the center of the screen (± 2.5 mm) where it was brought by the bee's motor actions. We used the same ray-casting method as in Lafon et al. 2021 (Lafon et al., 2021). 4950

4951 Weight and size measuring

Size was assessed by measuring the distance between the two wing joints using the Toupview
software (ToupTek Photonics, Zhejiang, China). After size measurements insects were placed
in an oven at 70°C for 4 hours in order to evaporate all water from their bodies. Dry weigh was
measured with a precision scale (OHAUS, Nänikon, Switzerland).

4956 Statistical analysis

4957 Statistical analyses were performed using R software (R Core Team, 2020). In Experiment 1 4958 (Color balancing), we counted the number of green and blue choices for each pair. We then 4959 used a chi² test to compare the distribution of green and blue of each pair with a theoretical 4960 distribution of 50% green and 50% blue. We report χ^2 , degrees of freedom and p-values for this 4961 analysis.

In Experiments 2 and 3, the first choice in each trial and test was categorized as choice of the
CS+, choice of the CS- or no choice (NC). Thus, a bee choosing the CS+ was recorded as (1,
0, 0) for choice of the CS+, choice of the CS- and NC, respectively. Data were bootstrapped to
plot the proportion of bees in each category with their corresponding 95% confidence interval.
Performances were analyzed using generalized mixed linear models (GLMM) with a binomial

error structure-logit-link function (glmer function of R package lme4) (Bates et al., 2014). The 4967 4968 independent variables (fixed factors) were the species of bee (Species; Experiment 2), the trial number (*Trial*), the choice category (*Choice*) and the color of the CS+ when applicable (*Color*: 4969 4970 Blue or Green). Bee ID was included as a random factor to account for the repeated-measure design; z values with corresponding degrees of freedom are reported throughout for this kind 4971 4972 of analysis. Post-hoc ANOVA were performed on those models to assess the impact of each 4973 factors. We report χ^2 with corresponding degrees of freedom throughout for this kind of 4974 analysis.

4975 During the tests of Experiments 2 and 3, we also recorded the time spent fixating the test 4976 alternatives (CS+ vs. CS-). Time values were compared using a Wilcoxon signed rank test.

For the acquisition trials, we recorded motor variables such as the total distance walked during 4977 a trial, and walking speed. In addition, we analyzed the latency to make a choice starting from 4978 4979 the beginning of a trial to the moment in which a choice (either for the CS+ or the CS-) was recorded. NC data were excluded from the latency analysis. The analysis of these continuous 4980 4981 variables was done using a linear mixed model (lmer function of R package lme4) in which the 4982 individual identity (Bee ID) was a random factor and the experimental condition (Condition) and trial number (Trial) were fixed factors. Statistical analyses were performed using R version 4983 4984 4.0.2 (R Core Team, 2020).

4985 **Results**

4986 Experiment 1: Establishing a balanced pair of green and blue stimuli

In a first experiment, we recorded the spontaneous choice of bumblebee presented with two pairs of green and blue stimulus. One pair called Green (R: 0, G:100, B:1.0; irradiance 24 370 μ W/cm²) versus Bright Blue (R: 0, G:80, B:254) and a second pair called Dark Green (R: 0, G:51, B:1.0) versus Blue (R: 0, G:0, B:254; irradiance 161 000 μ W/cm²). Each bee was presented once with both pair in a random order. There was no significant effect of the order on the bees' choices (z = 1.094, p = 0.27). Thus we pooled the data of the two test sequences and represented for both pair the number of bee choosing green or blue (Fig. 3).

When presented with a choice between Dark green and Blue, the distribution of choices was not significantly different from 50% (Green: 19, Blue: 23, $\chi^2 = 0.38095$, df = 1, p = 0.54). By contrast, with the Green versus Bright Blue pair, the distribution was significantly different from a random choice (Green: 28, Blue: 11, $\chi^2 = 7.41$, df:1, p = 0.0065) as the bees preferred the green option. The results indicate that only the Dark Green/Blue pair elicited a balanced spontaneous choice. We therefore chose to use this pair of colors in the subsequent experiments.



Figure 3. Experiment 1 - Choosing a balanced pair of green and blue for bumblebee conditioning. Quantification of the spontaneous phototactic responses of bumblebees (N = 51) towards a blue or green cylinder (see Fig. 2A). Two pairs of color were assayed in a random order, one called Green (R: 0, G:100, B:1.0; irradiance 24 370 μ W/cm²) versus Bright Blue (R: 0, G:80, B:254) and a second pair called Dark Green (R: 0, G:51, B:1.0) versus Blue (R: 0, G:0, B:255; irradiance 161 000 μ W/cm²). For each pair, the figure represents percentage of choice of each color. For subsequent experiments, the Dark Green vs Blue pair was used to condition bumblebees.

5009 Experiment 2: Comparing performances of bumblebees and honeybees

5010 in VR

5011 Bees had to learn to discriminate between a green and a blue cylinder presented against a dark

5012 uniform background. They could explore freely the virtual arena for 180 s and a choice was

- recorded when they got within 3 cm of one cylinder and centered it on the screen. Upon making
- a choice the bee received either 1.5 M sucrose solution as reward for choosing the CS+ or 3 M

- 5015 NaCl solution as a punishment associated with the CS-. Conditioning lasted for 6 trials followed
- 5016 by one non-reinforced test.
- 5017 Comparing attendance between honeybees and bumblebees
- 5018 During conditioning we discarded animals that failed to make a choice in 3 trials. Following
- that criterion, the proportion of discarded honeybee (59.74%) was significantly higher than the
- 5020 proportion of discarded bumblebees (10,87%) (Fig. 4) (z = 6.963, p < 0.0001). Overall,
- 5021 bumblebees made more choices and were discarded less than honeybees.



Figure 4. Experiment 2 - Comparing attendance between honeybees and bumblebees. Bee that did
 not make a choice for at least 3 of the 6 training trials were discarded. 59.74% of honeybee were
 discarded while only 10.78% bumblebees were. The difference is significant (z = 6.963, p < 0.0001).

5027 Discrimination learning during training

Figure 5 A-B shows the learning curves of both species of bees trained to discriminate the green from the blue cylinder under VR conditions and the cumulative heat maps displaying the locations of the bees in their trajectories during the six acquisition trials. Learning curves were obtained by recording the percentage of bees choosing correctly the CS+ or the CS- in their first choice, or not choosing any stimulus (NC) during each trial. No significant interaction between species, trial number and the bees' choice was found ($\chi 2 = 4.78$, df:2, p = 0.092), showing that both species followed similar learning dynamic.





5044 Bumblebee learning performances

The color of the CS+ had no effect on the learning of the bumblebees ($\chi^2 = 0.48$, df:2, p = 0.79). 5045 Data were thus represented as a CS+ vs. a CS- discrimination irrespective of color identity (Fig. 5046 5 A). Bumblebees learned to respond more to the CS+ than to the CS- (Fig. 5A). The interaction 5047 between trial number and bee choices was significant ($\chi 2 = 7.51$, df:2, p = 0.023). In the course 5048 of the 6 conditioning trials, the percentages of bees responding to the CS+ and that of bees 5049 5050 responding to the CS- evolved differently (z = 2.06, p = 0.04), thus showing successful discrimination learning. Moreover, the dynamic of CS+ responding bees was also significantly 5051 5052 different from that of the non-responding (NC) bees (z = 2.31, p = 0.021) while the difference between the dynamic of the CS- responding bees and the NC bees was not different (z = 1.083, 5053 p = 0.29). In the corresponding cumulative heat map (Fig. 5C), a clear V shape is visible, 5054 5055 indicating that the bees did interact with both sides in the VR and walked towards the cylinders.

5056 Honeybee learning performances

The color of the rewarded stimulus had no significant effect on honeybees 'ability to solve the discrimination task ($\chi 2 = 1.903$, df:2, p = 0.39). Data were thus pooled irrespective of the color of CS+ (Fig. 5B).

We found a significant interaction between the number of trials and the nature of the honeybees' choice ($\chi 2 = 8.89$, df:2, p = 0.012). Throughout the 6 trials the proportion of CS+ followed a different trajectory from that of CS- (z = 2.903, p = 0.0037), showing that the bees were able to successfully learn the discrimination (Fig. 5B). However, the dynamic of NC was neither different from CS- (z = -1.504, p = 0.133) nor from CS+ (z = 0.467, p = 0.641). The cumulative heat map representing the locations of the bees during their training trajectories (Fig. 5D) shows that, as with bumblebees, honeybees walked and interacted with both sides in the VR.

5067 Motor and temporal components of bee trajectories during training

We analyzed if there was any difference in the displacement of bees during the training trials between bumblebees and honeybees (Fig. 6). To do so, we quantified the distance walked (Fig. 6A), and the walking speed (Fig. 6B) of the bees during each trial. We also measured the choice latency in each trial (Fig. 6C), i.e. the time between the beginning of a trial and the first choice of the animal.

We found no significant interaction between the trial number and the species of the bees (*Trial*Species*: χ^2 0.7290, df:1, p = 0.40) suggesting no difference in the evolution of distance walked across trial between bumblebees and honeybees. The distance was, however, significantly affected by the species of the bee (*Species*: $\chi^2 = 10.91$, df:1, p < 0.001) as bumblebees walked more than honeybees (Fig. 6A). But did not vary significantly over trials (*Trial*: $\chi^2 = 3.2609$, df:1, p = 0.071).

The speed was significantly affected by the interaction between species and trials (*Trial*Species*: χ^2 6.2991, df:1, p = 0.012) and by trials alone (*Trial*: χ^2 = 32.5642, df:1, p < 0.0001). Meaning that both honeybees and bumblebees increased their speed over trials but the increase was steeper for honeybees (Fig. 6B). However, there was no speed difference between species (*Species*: χ^2 = 2.9235, df:1, p = 0.087).

The latency to choose decreased significantly across trials (Fig. 6C) (*Trial*: $\chi^2 = 22.2383$, df:1, p < 0.0001) at a similar rate in both species (*Trial*Species*: χ^2 0.1819, df:1, p = 0.6697) and was not significantly different overall between bumblebees and honeybees (*Species*: $\chi^2 = 0.0009$, df:1, p = 0.98).



Figure 6. Motor and temporal components of bee trajectories during the acquisition trials. For both bumblebees (*N* = 123) and Honeybees (*N* = 31), the evolution of **A**) the distance walked, **B**) the walking speed, and **C**) the choice latency during training trials is shown. The dashed lines above and below the curves represent the 95% confidence interval.

5093 Test Performance

After the last training trial, each bee was subjected to a test in which the green and the blue cylinders were presented in extinction conditions (no reinforcement provided). We recorded the percentage of bees choosing correctly the CS+ or the CS- in their first choice, or not choosing (NC) and the time spent fixating the CS+ and the CS- (Fig. 7). Similarly, to the learning phase bee species had no effect on the choices during the non-reinforced test ($\chi 2 = 0.407$, df:2, p = 0.816).

5100 Bumblebee test performances

- 5101 Contrary to the training phase, bumblebees' choice was affected by the color of the reward
- during the test (*Color*: $\chi^2 = 9.532$, df:2, p = 0.0085). We thus analyzed test results according to
- 5103 the color of reward (Fig. 7A C).

When blue was the rewarded color bumblebees (Fig. 7A) chose the CS+ significantly more than 5104 both CS- (z = 4.714, p < 0.00001) and NC (z = 5.882, p < 0.00001). The proportion of CS-5105 choices was also superior to the proportion of NC (z = -2.575, p = 0.01). Bumblebees were thus 5106 5107 able to solve the discrimination task during the test when the rewarded stimulus was blue (Fig. 7A). However, they did not manage to choose correctly during the test when CS+ was green 5108 (Fig. 7C), as the proportion of CS+ was not significantly different from the proportion of CS-5109 (z = 0.719, p = 0.472). The proportion of NC was significantly less than both CS+ (z = 4.643, z = 0.719)5110 p < 0.00001) and CS- (z = 4.185, p < 0.0001). Similarly, bumblebees did fixate the CS+ 5111 significantly longer than the CS- when blue was rewarded (Fig. 8A; V = 471, p = 0.001) but 5112 not when green was rewarded (Fig. 8C; V = 819, p = 0.271). 5113

5114 Honeybees test performances

5115 Contrary to the training phase, and similarly to bumblebees, color of the rewarded stimulus had 5116 an effect on honeybees' choice during the test (*Color*: $\chi^2 = 15.143$, df:2, p < 0.001). We thus 5117 represented and analyzed green rewarded and blue rewarded separately (Fig.7B D).

When the rewarded cylinder was blue (Fig.7B) bees chose CS+ significantly more than CS- (z 5118 = 3.238, p = 0.001). On the other hand, we found no significant difference neither between the 5119 proportion of NC and CS- (z = -0.007, p = 0.99) nor between NC and CS+ (z = 0.008, p = 0.99). 5120 5121 When the reward was green (Fig.7D) we found no significant difference between any of the 5122 three options (CS+ vs CS-: z = -0.743, p = 0.46; NC vs CS-: z = -1.514, p = 0.13; NC vs CS+: z = 0.819, p = 0.41). Thus, like bumblebees, honeybees were only able to solve the 5123 discrimination during the test when blue was the rewarded stimulus. When considering 5124 5125 centering time (Fig. 8B D), honeybees did not spend significantly more time on CS+ in any of the conditions (Green rewarded: Fig. 8D; V = 39, p = 0.41; Blue rewarded: Fig.8B; V = 44, p 5126 5127 = 0.23).







Figure 7. Test performances in a color discrimination learning task for bumblebees and honeybees. Percentages of bees choosing each stimulus (CS+: rewarded, CS-: punished, NC: no choice) during the non-reinforced test. Results are represented according to the color of the rewarded stimulus. Left Shows 1st choices from bumblebees (N = 123) when A) blue was rewarded (N = 61) and when C) green was rewarded (N = 62). Right Shows 1st choices from honeybees (N = 31) when B) blue was rewarded (N = 16) and when D) green was rewarded (N = 15).





5138Figure 8. Fixation time during the non-reinforced test. Time spent fixating either the CS+ or the CS-5139during the test. Bars represent the mean fixation time. Error bar represent the 95% confidence interval.5140Results are represented according to the color of the rewarded stimulus. Left Shows time from5141bumblebees (N = 123) when A) blue was rewarded (N = 61) and when C) green was rewarded (N = 62).5142Right Shows times from honeybees (N = 31) when B) blue was rewarded (N = 16) and when D) green5143was rewarded (N = 15).

5145 Insect dimensions and performances during the test

5146 After training we measure the inter wing distance and the dry weight of every bee. When then explored if and how did the dimensions of the insects affected their performances during the 5147 test (Fig. 9, 10, 11). Larger bumblebees walked further than smaller ones (Fig. 9A), as distance 5148 5149 walked was positively correlated with the size of the insect (F = 6.4756, df:1, p = 0.012) but not with weight (Fig. 10A) (F = 0.3171, df:1, p = 0.57). On the other hand, distance walked 5150 correlated neither with size (Fig. 9B; F = 1.0977, df:1, p = 0.30) nor with weight (Fig. 10B; F 5151 = 1.1189, df:1, p = 0.30) in honeybees. Larger bumblebees also walked faster than smaller ones 5152 (Fig. 9C) as speed was positively correlated with size in bumblebees (F = 6.4179, df:1, p =5153 0.013) but not with weight (Fig. 10C; F = 0.2985, df:1, p = 0.59) and neither with size (Fig. 9D; 5154 F = 1.0942, df:1, p = 0.30) nor with weight (Fig. 10D; F = 1.1113, df:1, p = 0.3) in honeybees. 5155 Coherent with the effect observed on speed, bumblebees' latency to choose was negatively 5156 5157 correlated with both size (Fig. 9E; F = 19.496, df:1, p < 0.0001) and weight (Fig. 10E; F =6.8365, df:1, p = 0.01). Meaning that bigger bumblebees made their first choice faster, which 5158 makes sense considering they also walked faster (Fig.9 A). For honeybees however, we found 5159 no clear effect of size (Fig. 9F) or weight (Fig. 10F) on the latency to choose (*Size*: F = 0.8738, 5160 df:1, p = 0.36; Weight: F = 0.1025, df:1, p = 0.75). It is likely that the weight and size variations 5161 5162 in honeybees are too small to significantly impact motor performances.







5169 walked. **C D**) Walking speed. **E F**) Latency to make a choice.





Figure 10. Impact of insect's weight on the motor and temporal performances of the bees during the test. After the last trial we measured the dry weight of each bee, reported here as *Weight*. This figure show the various parameter measured during the test, distance walked, speed and latency to choose as a function of insect's size. Each point is a bee and the pink line shows the linear regression.

- 5176 **A C E)** Show bumblebees value (*N* = 123). **B D F)** Show honeybee values (*N* = 31). **A B)** Distance
- 5177 walked. C D) Walking speed. E F) Latency to make a choice.
- 5178
In order to measure bees learning performance during the conditioning we established a learning score. The learning score is computed as the number of CS+ choices minus the number of CS- choices from the second trial to the test. The size of the bee didn't not significantly affect learning success (Fig. 11A, B; *Bumblebee*: $F = 8.10^{-4}$, df:1, p = 0.98; *Honeybee*: F = 2.0229, df:1, p = 0.17). Heavier honeybees had significantly lower learning scores than lighter ones (Fig. 11D; F = 5.3484, df:1, p = 0.03). However, no effect of weight was visible in bumblebees (Fig. 11C; F = 1.125, df:1, p = 0.291).



Figure 11. Impact of insect's dimensions on the learning performances. Shows learning score as a
function of the insect's size (A B) or weight (C D). For each bee we computed the learning score as
the difference between the number CS+ and CS- choices from the second trial to the test. Each point
represents a bee. The pink line shows the linear regression. A C) Bumblebees (N = 123). B D)
Honeybees (N = 31).

5193 Experiment 3: Conditioning bumblebees with Quinine as punishment

In a following experiment we conditioned bumblebees using 1.2 g.L⁻¹ quinine solution as punishment (Dyer and Chittka, 2004). The experiment followed the same protocol from experiment 2, 6 trials of maximum 180 s spaced by 60 s inter-trials followed by one nonreinforced test. The bumblebees had to discriminate between a blue and a green cylinder, we used the colors found in experiment 1, the color of the reward was decided pseudo-randomly, green was rewarded as often as blue.

5200 Discrimination learning during training

- 5201 The evolution of choices across trials was not affected by the color of the reward ($\chi^2 = 5.566$,
- 5202 df:2, p = 0.62). Therefore, we pooled both rewarded condition together and only shows the 5203 choices in term of CS+ and CS- irrespective of the color of the cylinders (Fig. 12).
- 5204 CS+ choices evolved differently from both CS- choices (z = -1.458, p < 0.00001) and non-
- 5205 choices (NC) (Fig. 12A; z = 5.223, p <0.00001) as bees learned to respond more to CS+. On
- 5206 the other hand, CS- choices did no evolved differently from NCs (z = 1.458, p = 0.14). In the
- 5207 corresponding cumulative heat map (Fig. 12D), a clear V shape is visible, indicating that the
- 5208 bees did interact with both sides in the VR and walked towards the cylinders.



5209

Figure 12. Test performances (1st choice and fixation time) of bumblebees in a color discrimination 5210 5211 learning task using quinine as negative reinforcement and sucrose as positive. As there were no 5212 significant differences the color conditions (blue or green rewarded), results were pooled (N = 235). 5213 The graph shows the percentage of bees responding to the CS+ (red), to the CS- (black) or not making 5214 any choice (NC; gray) during the training phase (A) and the retention test (B). C) Shows the time spent fixating each stimulus during the test. Error bars indicate 95% confidence intervals. *: p < 0.05; **: p < 5215 5216 0.01; ***: p < 0.001; ****: p < 0.0001. **D)** Cumulative heat maps of bumblebees' trajectories during 5217 the 6 learning trials (N = 235). E) Cumulative heat maps bumblebees' trajectories during the test ($N = 10^{-1}$ 235). Warmer color higher density of visits (see color scale). 5218

5219 Motor and temporal components of bee trajectories during training

5220 We also analyzed how the trajectory parameters of the bumblebees evolved over trials (Fig.

5221 13). The distanced walked decreased significantly across trials (Fig. 13A; $\chi^2 = 271.62$, df:1, p

- 5222 < 0.00001) as bumblebees took more direct routes toward the cylinders. Speed increased
- significantly with trials (Fig. 13B; $\chi^2 = 10.076$, df:1, p = 0.0015). This increase was concomitant
- 5224 with an increase in the proportion of bees choosing CS+ (Fig. 12A) and may thus reveal an

augmentation of motivation to reach the reward. Finally, coherent with an increase in speed and a decrease in distance the latency to make a choice decreased significantly across trials (Fig. 13C; $\chi^2 = 90.245$, df:1, p < 0.00001).



5228

Figure 13. Motor and temporal components of bumblebee trajectories during the acquisition trials.
 For both bumblebees conditioned in Experiment 3 (*N* = 235), shows the evolution of A) the distance
 walked, B) the walking speed, and C) the choice latency during training trials is shown. The dashed
 lines above and below the curves represent the 95% confidence interval.

5233 Test Performance

5234 As in experiment 2, bumblebees were submitted to a test where cylinders were not reinforced.

5235 We recorded the percentage of bees choosing correctly the CS+ or the CS- in their first choice,

- 5236 or not choosing (NC) and the time spent fixating the CS+ and the CS- (Fig. 12B).
- 5237 Bumblebee successfully learned the discrimination as they chose CS+ significantly more than
- 5238 both CS- and NC (Fig. 12B; CS-: z = 7.585, p < 0.00001; NC: z = 10.962, p < 0.00001). The

- 5240 made the wrong choice were motivated to interact with the VR.
- 5241 Consistent with their first choice, bumblebees spent more time fixating CS+ than CS- (Fig. 12C;
- 5242 V = 5781, p < 0.00001). In the corresponding cumulative heat map (Fig. 12E), we can see that
- 5243 bumblebees did interact with both sides and explored both cylinders.

5245 Discussion

5246 We have shown that bumblebees are able to solve a color discrimination task in a 3D VR setup. After six conditioning trials about 60% of bumblebees were able to correctly chose the rewarded 5247 stimulus during the non-reinforced test, which is similar to the performances displayed by 5248 honeybees, both in this study and in previous ones using a comparable protocol (Buatois et al., 5249 2017; Lafon et al., 2021). This success rate is however lower than what has been observed in 5250 5251 visual conditioning of the PER (Riveros and Gronenberg, 2012) where more than 70% of bumblebees responded to the rewarded stimulus after seven trials. Walking speed and latency 5252 to make a choice were also similar between bumblebees and honeybees, confirming the 5253 5254 suitability of the setup for both species.

When we used NaCl solution as punishment, bees from both species were only able to learn the 5255 discrimination if the rewarded stimulus was blue. Previous experiment in VR with honeybees 5256 have shown that NaCl was not always sufficient to produce a conditioned response (Buatois et 5257 5258 al., 2017). Bumblebees, and honeybees, have an innate preference for short wavelength colors 5259 like blue (Giurfa et al., 1995; Riveros and Gronenberg, 2012) but we did control in Experiment 5260 1 that the blue and green used elicited the same level of spontaneous attraction in bumblebees (Fig. 3), and the color used for honeybees have been controlled in previous studies (Buatois et 5261 5262 al., 2017). This result is confirmed by the fact that naïve bees in Experiment 2 did chose as much green as blue (Fig. 4A). However, bumblebees have also been shown to learn short 5263 wavelength colors faster (Gumbert, 2000). Thus, we can speculate that the strength of the NaCl 5264 reinforcement was not enough to overcome the difference in learning speed between green and 5265 5266 blue. When conditioned with Quinine, however, bumblebees displayed no color bias and were 5267 able to solve the discrimination task whether the reward was associated with blue or green, in accordance with previous findings where free-flying bumblebees trained to discriminate 5268 between two perceptually similar colors, one associated with 1.75 M sucrose solution, and the 5269

5270 other with water or quinine solution 120 mM, perform better if they experience quinine on the 5271 CS- targets rather than water (Chittka et al., 2003). It is also coherent with previous VR results 5272 where honeybees were able to solve the discrimination when CS- was associated with quinine, 5273 but not when it was associated with NaCl (Buatois et al., 2017). In Experiment 3, bumblebees 5274 also show a significant decrease in both the distance walked and the latency to make a choice 5275 (*i.e.* approaching and centering a stimulus), confirming that they acquire the task over trials and 5276 learned to navigate the VR environment.

While learning performances were similar, bumblebees had nevertheless a higher motivation 5277 5278 than honeybees, as 60% of honeybees did not make enough choices to be kept in the analysis 5279 against only 11% in bumblebees. Artificially forcing the insect to walk might conflict less with the natural ecology of *Bombus terrestris* who lives in underground nests than it does for 5280 honeybee foragers who spend most of their time in flight. This contrast would have been 5281 increased further in our study as the honeybees were foragers collected from a feeder in our 5282 apiary while the bumblebees were collected from their commercial nest box that they had never 5283 5284 left in their life. Additionally, the analysis of motor parameters revealed that bumblebees walk longer distances than honeybees, which was likely due to the bigger size of bumblebees since 5285 walking distance and speed were positively correlated with size in bumblebees. Thus, it is 5286 5287 possible that bumblebees had an easier time walking on the treadmill due to their bigger size and ecology (Dahmen et al., 2017). 5288

We found no relation between size and learning performances in our experiment. Several studies have shown that bigger bumblebees have a better visual acuity than smaller bees (Macuda et al., 2001; Spaethe and Chittka, 2003; Wertlen et al., 2008; Taylor et al., 2019).Throughout the experiment the cylinders subtended a visual angle of at least 6° at the beginning of a trial and up to 53° when making a choice, they were thus always above the minimal 5° necessary for color vision in honeybee (Giurfa et al., 1996). It is thus unlikely that

higher visual acuity could help in better solving the discrimination. Larger bumblebees have 5295 been shown to learn faster in free flight condition (Worden et al., 2005). Larger bumblebees 5296 also assumes the role of forager for the colony (Goulson, 2003), a role for which they are better 5297 suited than smaller workers (Heinrich and Heinrich, 1983; Goulson et al., 2002; Spaethe and 5298 Weidenmüller, 2002; Ings et al., 2005; Klein et al., 2017). So it is possible that the learning 5299 abilities of larger bumblebees come from their life experience as foragers (Giurfa et al., 2003; 5300 Cabirol et al., 2017), however in this study all bumblebees were naive as they never left their 5301 nest box prior to the experiment which would have prevented the bigger workers to improve 5302 their learning abilities through higher foraging experience. Finally, this is not the first study to 5303 find no link between size and learning success, in 2008 Raine and Chittka trained free flying 5304 bumblebees to collect a sucrose rewarded from colored feeders and found no correlation 5305 between insect size and learning speed (Raine and Chittka, 2008). 5306

5307 On the other hand, we found a negative effect of body weight on honeybee's learning 5308 performances, it is possible that bees with a higher fat storage had a lower motivation to solve 5309 the task as foraging is coupled with a reduced rate of fat in this species (Toth et al., 2005; Toth 5310 and Robinson, 2005). Consequently, fatter bees might have been less experienced young 5311 foragers, thus explaining relative lower performances.

5313 Conclusion

5314 Our results clearly demonstrate the ability of bumblebees to solve a color discrimination task under VR condition. While learning performances were similar to those of honeybees, 5315 bumblebees engaged more with the VR and thus less individuals were discarded because of a 5316 5317 lack of choices in bumblebees, confirming the suitability of bumblebees as a model for the study of visual learning in VR. We found that bumblebees solved the discrimination better when 5318 5319 the CS- was paired with quinine rather than NaCl. No correlation between bumblebee size and learning success was evidenced which might suggest that the better learning success observed 5320 in larger bumblebees in the literature were linked to a higher foraging experience as the 5321 5322 bumblebees we tested were all naive. These results, associated with their high resilience, point 5323 at the bumblebees as a prime candidate to explore the underlying mechanisms of visual learning by coupling VR experiments with invasive electrophysiological or calcium imaging studies. 5324

5325 Acknowledgements

We thank Mathieu Lihoreau for his help in acquiring and maintaining the bumblebee colonies.
We also thank Benjamin H. Paffhausen, and Marco Paoli for valuable discussions and support.
This work was supported by an ERC Advanced Grant ('Cognibrains') to M.G, who also thanks
the Institut Universitaire de France (IUF), the CNRS and the University Paul Sabatier for
support.

5331 Contributions

The project was conceived by AAW, MG and GL. G.L performed all the behavioral experiments. Naïs Judan, Eva Blot, and Karolina Pecharova also assisted with the behavioral experiments. Behavioral experiments were supervised by M.G. and A.A.-W. Statistical analyses were performed by G.L. The manuscript was written by G.L and was corrected and

- discussed by all authors. M. G. obtained the funding necessary for this work. All authorsreviewed and approved the final version of the manuscript.
- 5338 Ethics declarations

5339 **Competing interests**

5340 The authors declare no competing interests.

5342 References

- 5343 Anfora, G., Rigosi, E., Frasnelli, E., Ruga, V., Trona, F., Vallortigara, G., 2011. Lateralization in the
- Invertebrate Brain: Left-Right Asymmetry of Olfaction in Bumble Bee, Bombus terrestris. PLOS ONE 6,
 e18903. https://doi.org/10.1371/journal.pone.0018903
- Bates, D., Mächler, M., Bolker, B., Walker, S., 2014. Fitting Linear Mixed-Effects Models using Ime4.
 ArXiv14065823 Stat.
- Bitterman, M.E., Menzel, R., Fietz, A., Schäfer, S., 1983. Classical conditioning of proboscis extension
 in honeybees (Apis mellifera). J. Comp. Psychol. Wash. DC 1983 97, 107–119.
- 5350 Buatois, A., Flumian, C., Schultheiss, P., Avarguès-Weber, A., Giurfa, M., 2018. Transfer of Visual
- Learning Between a Virtual and a Real Environment in Honey Bees: The Role of Active Vision. Front.
 Behav. Neurosci. 12, 139. https://doi.org/10.3389/fnbeh.2018.00139
- 5353 Buatois, A., Laroche, L., Lafon, G., Avarguès-Weber, A., Giurfa, M., 2020. Higher-order discrimination
- 5354 learning by honeybees in a virtual environment. Eur. J. Neurosci. 51, 681–694.
- 5355 https://doi.org/10.1111/ejn.14633
- 5356 Buatois, A., Pichot, C., Schultheiss, P., Sandoz, J.-C., Lazzari, C.R., Chittka, L., Avarguès-Weber, A.,
- 5357 Giurfa, M., 2017. Associative visual learning by tethered bees in a controlled visual environment. Sci.
- 5358 Rep. 7. https://doi.org/10.1038/s41598-017-12631-w
- 5359 Cabirol, A., Brooks, R., Groh, C., Barron, A.B., Devaud, J.-M., 2017. Experience during early adulthood
- 5360 shapes the learning capacities and the number of synaptic boutons in the mushroom bodies of honey
- bees (Apis mellifera). Learn. Mem. Cold Spring Harb. N 24, 557–562.
- 5362 https://doi.org/10.1101/lm.045492.117
- 5363 Chittka, L., Dyer, A.G., Bock, F., Dornhaus, A., 2003. Bees trade off foraging speed for accuracy.
 5364 Nature 424, 388–388. https://doi.org/10.1038/424388a
- Chittka, L., Thomson, J.D., 1997. Sensori-motor learning and its relevance for task specialization in
 bumble bees. Behav. Ecol. Sociobiol. 41, 385–398. https://doi.org/10.1007/s002650050400
- 5367 Church, D.L., Plowright, C.M.S., 2006. Spatial encoding by bumblebees (Bombus impatiens) of a
 5368 reward within an artificial flower array. Anim. Cogn. 9, 131–140. https://doi.org/10.1007/s100715369 005-0011-6
- 5370 Cnaani, J., Thomson, J.D., Papaj, D.R., 2006. Flower Choice and Learning in Foraging Bumblebees:
- 5371 Effects of Variation in Nectar Volume and Concentration. Ethology 112, 278–285.
- 5372 https://doi.org/10.1111/j.1439-0310.2006.01174.x
- Dahmen, H., Wahl, V.L., Pfeffer, S.E., Mallot, H.A., Wittlinger, M., 2017. Naturalistic path integration
 of Cataglyphis desert ants on an air-cushioned lightweight spherical treadmill. J. Exp. Biol. 220, 634–
 644. https://doi.org/10.1242/jeb.148213
- 5376 Dukas, R., Waser, N.M., 1994. Categorization of food types enhances foraging performance of 5377 bumblebees. Anim. Behav. 48, 1001–1006. https://doi.org/10.1006/anbe.1994.1332
- 5378 Dyer, A.G., Chittka, L., 2004. Biological significance of distinguishing between similar colours in
- 5379 spectrally variable illumination: bumblebees (Bombus terrestris) as a case study. J. Comp. Physiol.
- 5380 [A] 190, 105–114. https://doi.org/10.1007/s00359-003-0475-2

- 5381 Foster, J.J., Sharkey, C.R., Gaworska, A.V.A., Roberts, N.W., Whitney, H.M., Partridge, J.C., 2014.
- 5382 Bumblebees learn polarization patterns. Curr. Biol. CB 24, 1415–1420.
- 5383 https://doi.org/10.1016/j.cub.2014.05.007

5384 Frasnelli, E., Robert, T., Chow, P.K.Y., Scales, B., Gibson, S., Manning, N., Philippides, A.O., Collett,

- 5385 T.S., Hempel de Ibarra, N., 2021. Small and Large Bumblebees Invest Differently when Learning about 5386 Flowers. Curr. Biol. 31, 1058-1064.e3. https://doi.org/10.1016/j.cub.2020.11.062
- Giurfa, M., 2007. Behavioral and neural analysis of associative learning in the honeybee: a taste from
 the magic well. J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol. 193, 801–824.
 https://doi.org/10.1007/s00359-007-0235-9
- 5390 Giurfa, M., Núñez, J., Chittka, L., Menzel, R., 1995. Colour preferences of flower-naive honeybees. J. 5391 Comp. Physiol. A 177, 247–259. https://doi.org/10.1007/BF00192415
- 5392 Giurfa, M., Sandoz, J.-C., 2012. Invertebrate learning and memory: Fifty years of olfactory
- 5393 conditioning of the proboscis extension response in honeybees. Learn. Mem. 19, 54–66.
- 5394 https://doi.org/10.1101/lm.024711.111
- Giurfa, M., Schubert, M., Reisenman, C., Gerber, B., Lachnit, H., 2003. The effect of cumulative
 experience on the use of elemental and configural visual discrimination strategies in honeybees.
 Behav. Brain Res. 145, 161–169. https://doi.org/10.1016/S0166-4328(03)00104-9
- Giurfa, M., Vorobyev, M., Kevan, P., Menzel, R., 1996. Detection of coloured stimuli by honeybees:
 Minimum visual angles and receptor specific contrasts. J. Comp. Physiol. [A] 178, 699–709.
 https://doi.org/10.1007/BF00227381
- 5401 Goulson, D., 2003. Bumblebees: Their Behaviour and Ecology. Oxford University Press.
- Goulson, D., Peat, J., Stout, J.C., Tucker, J., Darvill, B., Derwent, L.C., Hughes, W.O.H., 2002. Can
 alloethism in workers of the bumblebee, Bombus terrestris, be explained in terms of foraging
 efficiency? Anim. Behav. 64, 123–130. https://doi.org/10.1006/anbe.2002.3041
- 5405 Gumbert, A., 2000. Color choices by bumble bees (Bombus terrestris): innate preferences and 5406 generalization after learning. Behav. Ecol. Sociobiol. 48, 36–43.
- 5407 https://doi.org/10.1007/s002650000213
- 5408 Heinrich, B., Heinrich, M.J.E., 1983. Heterothermia in Foraging Workers and Drones of the
- 5409 Bumblebee Bombus terricola. Physiol. Zool. 56, 563–567.
- 5410 https://doi.org/10.1086/physzool.56.4.30155879
- Heinrich, B., Mudge, P.R., Deringis, P.G., 1977. Laboratory Analysis of Flower Constancy in Foraging
 Bumblebees: Bombus ternarius and B. terricola. Behav. Ecol. Sociobiol. 2, 247–265.
- Ings, T.C., Schikora, J., Chittka, L., 2005. Bumblebees, humble pollinators or assiduous invaders? A
 population comparison of foraging performance in Bombus terrestris. Oecologia 144, 508–516.
 https://doi.org/10.1007/s00442-005-0081-9
- Keasar, T., Motro, U., Shur, Y., Shmida, A., 1996. Overnight memory retention of foraging skills by
 bumblebees is imperfect. Anim. Behav. 52, 95–104. https://doi.org/10.1006/anbe.1996.0155
- Klein, S., Pasquaretta, C., Barron, A.B., Devaud, J.-M., Lihoreau, M., 2017. Inter-individual variability in
 the foraging behaviour of traplining bumblebees. Sci. Rep. 7, 4561. https://doi.org/10.1038/s41598017-04919-8

- Kulahci, I.G., Dornhaus, A., Papaj, D.R., 2008. Multimodal signals enhance decision making in foraging
 bumble-bees. Proc. R. Soc. B Biol. Sci. 275, 797–802. https://doi.org/10.1098/rspb.2007.1176
- Lafon, G., Howard, S.R., Paffhausen, B.H., Avarguès-Weber, A., Giurfa, M., 2021. Motion cues from
- 5424 the background influence associative color learning of honey bees in a virtual-reality scenario. Sci.
- 5425 Rep. 11, 21127. https://doi.org/10.1038/s41598-021-00630-x
- Laverty, T.M., 1994. Bumble bee learning and flower morphology. Anim. Behav. 47, 531–545.
 https://doi.org/10.1006/anbe.1994.1077
- Leadbeater, E., Chittka, L., 2007. The dynamics of social learning in an insect model, the bumblebee
 (Bombus terrestris). Behav. Ecol. Sociobiol. 61, 1789–1796. https://doi.org/10.1007/s00265-0070412-4
- Leadbeater, E., Chittka, L., 2005. A new mode of information transfer in foraging bumblebees? Curr.
 Biol. CB 15, R447-448. https://doi.org/10.1016/j.cub.2005.06.011
- 5433 Leonard, A.S., Dornhaus, A., Papaj, D.R., 2011. Flowers help bees cope with uncertainty: signal
- 5434 detection and the function of floral complexity. J. Exp. Biol. 214, 113–121.
- 5435 https://doi.org/10.1242/jeb.047407
- Li, L., Su, S., Perry, C.J., Elphick, M.R., Chittka, L., Søvik, E., 2018. Large-scale transcriptome changes in
 the process of long-term visual memory formation in the bumblebee, Bombus terrestris. Sci. Rep. 8,
- 5438 534. https://doi.org/10.1038/s41598-017-18836-3
- 5439 Lichtenstein, L., Sommerlandt, F.M.J., Spaethe, J., 2015. Dumb and Lazy? A Comparison of Color
- 5440 Learning and Memory Retrieval in Drones and Workers of the Buff-Tailed Bumblebee, Bombus
- 5441 terrestris, by Means of PER Conditioning. PLOS ONE 10, e0134248.
- 5442 https://doi.org/10.1371/journal.pone.0134248
- Lihoreau, M., Raine, N.E., Reynolds, A.M., Stelzer, R.J., Lim, K.S., Smith, A.D., Osborne, J.L., Chittka, L.,
 2013. Unravelling the mechanisms of trapline foraging in bees. Commun. Integr. Biol. 6, e22701.
 https://doi.org/10.4161/cib.22701
- Macuda, T., Gegear, R., Laverty, T., Timney, B., 2001. Behavioural assessment of visual acuity in
 bumblebees (Bombus impatiens). J. Exp. Biol. 204, 559–564. https://doi.org/10.1242/jeb.204.3.559
- Mertes, M., Carcaud, J., Sandoz, J.-C., 2021. Olfactory coding in the antennal lobe of the bumble bee
 Bombus terrestris. Sci. Rep. 11, 10947. https://doi.org/10.1038/s41598-021-90400-6
- Mertes, M., Dittmar, L., Egelhaaf, M., Boeddeker, N., 2014. Visual motion-sensitive neurons in the
 bumblebee brain convey information about landmarks during a navigational task. Front. Behav.
 Neurosci. 8.
- 5453 Muth, F., Breslow, P.R., Masek, P., Leonard, A.S., 2018. A pollen fatty acid enhances learning and 5454 survival in bumblebees. Behav. Ecol. 29, 1371–1379. https://doi.org/10.1093/beheco/ary111
- Paulk, A.C., Gronenberg, W., 2008. Higher order visual input to the mushroom bodies in the bee,
 Bombus impatiens. Arthropod Struct. Dev. 37, 443–458. https://doi.org/10.1016/j.asd.2008.03.002
- 5457 Paulk, A.C., Phillips-Portillo, J., Dacks, A.M., Fellous, J.-M., Gronenberg, W., 2008. The processing of
- 5458 color, motion, and stimulus timing are anatomically segregated in the bumblebee brain. J. Neurosci.
- 5459 Off. J. Soc. Neurosci. 28, 6319–6332. https://doi.org/10.1523/JNEUROSCI.1196-08.2008

- R Core Team, 2020. R Development Core team (2020). R: A language and environment for statisticalcomputing. Vienna, Austria.
- Raine, N.E., Chittka, L., 2008. The correlation of learning speed and natural foraging success in
 bumble-bees. Proc. R. Soc. B Biol. Sci. 275, 803–808. https://doi.org/10.1098/rspb.2007.1652

Real, L.A., 1992. Information Processing and the Evolutionary Ecology of Cognitive Architecture. Am.
Nat. 140, S108–S145. https://doi.org/10.1086/285399

5466Riddell, C.E., Mallon, E.B., 2006. Insect psychoneuroimmunology: Immune response reduces learning5467in protein starved bumblebees (Bombus terrestris). Brain. Behav. Immun. 20, 135–138.

- 5468 https://doi.org/10.1016/j.bbi.2005.06.008
- Riveros, A. J., Gronenberg, W., 2009. Learning from learning and memory in bumblebees. Commun.Integr. Biol. 2, 437–440.
- 5471 Riveros, A.J., Gronenberg, W., 2012. Decision-making and associative color learning in harnessed
- 5472 bumblebees (Bombus impatiens). Anim. Cogn. 15, 1183–1193. https://doi.org/10.1007/s10071-012-5473 0542-6
- 5474Riveros, Andre J., Gronenberg, W., 2009. Olfactory learning and memory in the bumblebee Bombus5475occidentalis. Naturwissenschaften 96, 851–856. https://doi.org/10.1007/s00114-009-0532-y
- 5476Riveros, A.J., Leonard, A.S., Gronenberg, W., Papaj, D.R., 2020. Learning of bimodal versus unimodal5477signals in restrained bumble bees. J. Exp. Biol. 223, jeb220103. https://doi.org/10.1242/jeb.220103
- 5478 Robert, T., Frasnelli, E., Hempel de Ibarra, N., Collett, T.S., 2018. Variations on a theme: bumblebee
- 5479 learning flights from the nest and from flowers. J. Exp. Biol. 221, jeb172601.
- 5480 https://doi.org/10.1242/jeb.172601
- Rusanen, J., Vähäkainu, A., Weckström, M., Arikawa, K., 2017. Characterization of the first-order
 visual interneurons in the visual system of the bumblebee (Bombus terrestris). J. Comp. Physiol. A
 203, 903–913. https://doi.org/10.1007/s00359-017-1201-9
- Saleh, N., Chittka, L., 2007. Traplining in bumblebees (Bombus impatiens): a foraging strategy's
 ontogeny and the importance of spatial reference memory in short-range foraging. Oecologia 151,
 719–730. https://doi.org/10.1007/s00442-006-0607-9
- 5487 Skorupski, P., Chittka, L., 2010a. Photoreceptor Spectral Sensitivity in the Bumblebee, Bombus
- 5488 impatiens (Hymenoptera: Apidae). PLOS ONE 5, e12049.
- 5489 https://doi.org/10.1371/journal.pone.0012049
- 5490 Skorupski, P., Chittka, L., 2010b. Differences in Photoreceptor Processing Speed for Chromatic and
- 5491 Achromatic Vision in the Bumblebee, Bombus terrestris. J. Neurosci. 30, 3896–3903.
- 5492 https://doi.org/10.1523/JNEUROSCI.5700-09.2010
- 5493 Skorupski, P., Döring, T.F., Chittka, L., 2007. Photoreceptor spectral sensitivity in island and mainland
 5494 populations of the bumblebee, Bombus terrestris. J. Comp. Physiol. A 193, 485–494.
- 5495 https://doi.org/10.1007/s00359-006-0206-6
- 5496 Spaethe, J., Brockmann, A., Halbig, C., Tautz, J., 2007. Size determines antennal sensitivity and
- 5497 behavioral threshold to odors in bumblebee workers. Naturwissenschaften 94, 733–739.
- 5498 https://doi.org/10.1007/s00114-007-0251-1

- 5499 Spaethe, J., Chittka, L., 2003. Interindividual variation of eye optics and single object resolution in 5500 bumblebees. J. Exp. Biol. 206, 3447–3453. https://doi.org/10.1242/jeb.00570
- 5501 Spaethe, J., Weidenmüller, A., 2002. Size variation and foraging rate in bumblebees (Bombus 5502 terrestris). Insectes Sociaux 49, 142–146. https://doi.org/10.1007/s00040-002-8293-z
- 5503 Taylor, G.J., Tichit, P., Schmidt, M.D., Bodey, A.J., Rau, C., Baird, E., 2019. Bumblebee visual allometry
- results in locally improved resolution and globally improved sensitivity. eLife 8, e40613.
 https://doi.org/10.7554/eLife.40613
- Toda, N.R.T., Song, J., Nieh, J.C., 2009. Bumblebees exhibit the memory spacing effect.
 Naturwissenschaften 96, 1185–1191.
- Toth, A.L., Kantarovich, S., Meisel, A.F., Robinson, G.E., 2005. Nutritional status influences socially
 regulated foraging ontogeny in honey bees. J. Exp. Biol. 208, 4641–4649.
 https://doi.org/10.1242/jeb.01956
- 5511 Toth, A.L., Robinson, G.E., 2005. Worker nutrition and division of labour in honeybees. Anim. Behav.
- 5512 69, 427–435. https://doi.org/10.1016/j.anbehav.2004.03.017
- 5513 Vähäkainu, A., Vähäsöyrinki, M., Weckström, M., 2013. Membrane filtering properties of the
- bumblebee (Bombus terrestris) photoreceptors across three spectral classes. J. Comp. Physiol. A 199,
 629–639. https://doi.org/10.1007/s00359-013-0814-x
- 5516 Wertlen, A.M., Niggebrügge, C., Vorobyev, M., Hempel de Ibarra, N., 2008. Detection of patches of 5517 coloured discs by bees. J. Exp. Biol. 211, 2101–2104. https://doi.org/10.1242/jeb.014571
- 5518 Worden, B.D., Skemp, A.K., Papaj, D.R., 2005. Learning in two contexts: the effects of interference
- 5519 and body size in bumblebees. J. Exp. Biol. 208, 2045–2053. https://doi.org/10.1242/jeb.01582

General Discussion

5523 General discussion

5524 With this PhD work, we aimed at improving an existing VR set-up in order to overcome the impossibility of accessing the nervous system of flying bees solving visual problems. In the 5525 5526 pursuit of this goal we established a new 3D VR and showed that increasing the complexity of the virtual background is detrimental to successful conditioning, as the presence of frontal 5527 background motion cues impaired the bees' performance in color discrimination learning. We 5528 5529 then used that VR to condition honeybees in a controlled visual environment which was a prerequisite to quantify variation in IEGs expression specific to visual learning. This revealed 5530 an implication of the optic lobes and the calyces of the mushroom bodies in visual learning, and 5531 5532 that solving a color discrimination task in 2D and 3D involved different neural mechanisms as 5533 both modes lead to different pattern of IEG expression. Finally, we showed that bumblebees would be an ideal alternative model for future VR experiments, since they are easier to 5534 5535 condition in high numbers than honeybees, while being probably more robust to invasive brain recording techniques. 5536

5537 Virtual reality: progress, limitations and future developments

The first chapter of this thesis represents the first publication of a fully 3D virtual setup showing 5538 visual learning in harnessed honeybees. Zwaka et al did publish a setup where the colored 5539 stimuli, a blue and yellow colored stripped, could grow larger as the insect walked toward them 5540 5541 in 2019. But the behavioral results don't show clearly whether or not the bees managed to learn 5542 the color discrimination. Their setup also included gray stripes of different heights in the background, simulating a far-distant skyline, which might have interfered with color learning 5543 5544 as our work showed. Our setup has several advantages over previous attempts. First we paid 5545 more attention to the weight of the treadmill, using a smaller and lighter treadmill of 5cm for 1g which were thus easier to mobilize for the bees and resulted in a more natural behavior. This 5546 is an improvement on previous setups that used a 10 cm Styrofoam ball (Buatois et al., 2017, 5547

2018; Zwaka et al., 2019; Buatois et al., 2020) which would have had a weight of about 7.8g 5548 5549 and thus require from the bee 31 time the force required to move itself on a flat surface compared to only 5 time in our current setup (Dahmen et al., 2017). We recently acquired 5550 5551 specially made hollow balls that might allow to drive that number further down. Our VR software is based on a video game engine (Unity) which offers two main advantages. First, they 5552 5553 have been the focus of intense optimization work in the past decades to reduce as much as 5554 possible the latency between player input and reaction from the software, thus in our current 5555 implementation we measured a latency of 18.00 ± 2.53 ms between a movement of the ball and a reaction in the VR. Keeping the latency as low as possible is very important to create a 5556 5557 believable illusion as honeybee vision has a very high temporal resolution (Srinivasan and Lehrer, 1984). Second, video game engines offer a wide set of tools that make maintaining and 5558 expending the possibilities of our VR software easier. We have made our software open source 5559 5560 (https://github.com/G-Lafon/BeeVR) which should provide a simple and adaptable solution to support a variety a conditioning protocol. Moreover, this gives us the opportunity to keep 5561 5562 growing its possibilities outside of our own projects by allowing other teams to contribute the 5563 functionality they need in their work.

In chapter one, we focused on investigating the influence of motion cues, both frontal and 5564 5565 ventral, on associative color learning in VR. By enriching the VR with a background producing optic flow we were expecting to improve bees' performances as optic flow has been shown to 5566 be crucial for flight navigation in bees (Baird et al., 2006; Frasnelli et al., 2018; Baird et al., 5567 2021), as well as to improve visual learning in harnessed condition (Balamurali et al., 2015). It 5568 5569 was thus surprising to find that motion cues from the background of the VR impaired color 5570 discrimination of objects located in the virtual foreground. Indeed, only when the background was empty were the bees able to properly learn the discrimination. However, in the condition 5571 5572 where optic flow from the background was artificially suppressed, by fixing the grating to the

bee's gaze, honeybees spend more time fixating the rewarded object during the non-rewarded 5573 5574 test, despite not showing a clear preference in term of first choices, which suggest a learning effect albeit less strong than in the empty background condition. Suppressing motion cues 5575 5576 allowed thus to rescue some learning abilities, suggesting that both the motion cues emanating from the background, and its illumination conditions, may have interfered with color learning 5577 5578 in the VR arena. To explain this effect, we hypothesized that both the luminosity and motion 5579 cues from the background might have distracted the insects by decreasing their attention toward the objects of interest. By contrast ventral motion cues had no effect on color learning 5580 performances, in concordance with our previous conclusions as the ventral motion cues 5581 5582 emerged from painted dots on the treadmill and provided thus no additional distracting luminosity while they were not competing spatially with the colored cuboids. However, ventral 5583 motion cues did affect the walking speed of bees. Honeybees were walking slower when the 5584 5585 treadmill motion was generating more optic flow. We know that ventral optic flow is a measure of distance for flying bees, these results thus suggest that bees also use it to evaluate walked 5586 5587 distances.

In the introduction we claimed that the existing VR setup were not at the level of efficiency of PER to trigger learning in bees as the associated learning rate is slower. Unfortunately, increasing the complexity of the VR by introducing a richer background did not prove to be a solution to improve learning performances in VR. So, if enriching the VR is not a solution, what options are left to improve learning rates in VR?

A few technical improvements are still possible: replacing the video projector with a panel of LEDs to increase the refresh rate beyond 200 Hz (limit of the bee's eye temporal resolution), and reducing the weight of the ball even further by using hollow Styrofoam ball. We could also introduce an automatized reward system, as was done in Zwaka et al. 2019, to increase consistency in reward delivery and the experimental productivity by running several

replications in parallel due to a full automatization of the setup. However, concerning the virtual 5598 5599 environment itself, it seems that we have reached the limit of what can be achieved as making the virtual world more complex reduced performances (chapter 1) while moving from 2D to 5600 5601 3D did not improve learning performances (chapter 3), where the same setup with 2D VR, lead to the same 58% of correct choices during the test. Thus, contrary to our expectations, increased 5602 5603 complexity does not produce better results. In the context of visual learning, the future 5604 development of VR should hence focus on actually making the VR simpler. It might not improve learning performances but we now know that it will not impede them, and it will get 5605 us closer to an equivalent of PER for the exploration of visual learning with a simple and easily 5606 5607 reproducible setup that can be used by many teams at little cost.

Now that we've established functioning protocols for differential learning in VR (chapter 1 and
2) (Buatois et al., 2017; Lafon et al., 2021) future developments can focus on adapting protocols
for more complex learning task like reversal learning, or concept learning. This would open the
higher-order cognitive abilities of the bees to deeper investigations.

5612 Investigating the brain regions involved in differential visual learning in VR

In chapter 2, we used the 3D VR environment established in chapter 1 to study visual learning 5613 5614 and determine if it leads to changes in immediate early gene (IEG) expression in specific areas of the bee brain. We focused on 3 IEGs related to bee foraging and orientation, kakusei, Hr38 5615 5616 and *Erg1*. This work represents the first controlled experiments on the variations of IEGs level 5617 in the honeybee brain as a result of visual learning, as previous study focused on foraging and 5618 orientation flights (Kiya et al., 2007; Lutz and Robinson, 2013; Fujita et al., 2013; Singh et al., 2018; Ugajin et al., 2018; Iino et al., 2020). Our experiment revealed an increase in Egr1 5619 5620 expression in the calvees of the mushroom bodies of learners compared to non-learners. Even 5621 though the implication of mushroom bodies in visual learning in the bee is expected given the crucial role of mushroom bodies for the acquisition, storage and retrieval of olfactory memories 5622

(Menzel, 1999, 2014; Devaud et al., 2015) and the parallels between visual and olfactory inputs 5623 5624 at the level of the calyces, studies addressing the role of mushroom bodies in honey bee visual learning and memory remain rare. This finding is coherent with previous results that found an 5625 5626 increase in the dopaminergic receptor gene Amdop1 in the calyces of the mushroom bodies as a result of aversive visual learning (Marchal et al., 2019). The fact that we found no visual 5627 learning induced variations of *kakusei* despite reports of enhanced expression in foragers or in 5628 5629 orienting bees (Kiya et al., 2007; Kiya and Kubo, 2010; Ugajin et al., 2018) suggests that the expression is not necessarily related to learning occurring in these contexts. 5630

5631 We did not find any variation of expression in the central brain despite previous studies 5632 suggesting a role of the CX in visual learning (Plath et al., 2017). This could be due to the limited spatial resolution of our brain dissection. Indeed, our current technique did not allow us 5633 to isolate the CX from adjacent structures, mainly the subesophageal zone and the peduncle of 5634 the mushroom bodies. It is possible that variations in the CX got diluted by unspecific responses 5635 from the adjacent structures that got included in the dissection. Future experiments would 5636 5637 benefit from more precise dissections using, for example, Laser-Capture Microdissection that would allow dissection at cellular resolution. Higher spatial resolution could also allow to 5638 investigate other sub regions of the brain, as previous studies suggested a role of the medial 5639 5640 lobes of the mushroom bodies in visual learning (Plath et al., 2017) for example.

IEGs are transcribed transiently and rapidly in response to specific stimulations inducing neural activity without de novo protein synthesis (Bahrami and Drabløs, 2016), since their expression is part of the early stages of the cell response to an external stimulus, the quantification of IEGs expression level is a good proxy for cell activity. On the other hand, since they are at such an early stage of the cell response, they integrate signals from many different inputs. This means that in order to control the specificity of the measured response, the inputs must be tightly controlled. This is where VR was important, since it allows to completely control the visual inputs presented to the insect during the conditioning. We can thus make sure that all bees had
the same visual experience and that the difference observed between learners and non-learners
is specific of the learning and not caused be a difference in sensory input.

The data from chapter 2 confirmed that the mushroom bodies are involved in visual learning and show that virtual reality can be successfully used to investigate the neural mechanism of visual learning. However, the analysis here was done ex-vivo after the conditioning. In order to go further it would be interesting to be able to perform live recordings from the bees as they learn. We know from previous works that it is possible to record electrophysiological activity from behaving bees in VR (Paulk et al., 2014; Zwaka et al., 2019; Rusch et al., 2021). Coupling our VR setup with electrophysiological recordings thus appear like a logical next step.

5658 In the third chapter we reproduced the previous experiment but using a simpler, more restrictive 5659 2D VR. Surprisingly we found a different pattern of activation by comparison with the 3D VR 5660 despite the fact that the conditioning protocol and the stimuli used were similar.

We showed that, in 2D conditions, associative color learning led to a downregulation of the 5661 three IEGs considered in different areas of the visual circuit in the learner group compared to 5662 the non-learners. While Egrl was downregulated in the optic lobes, Hr38 and kakusei were 5663 coincidently downregulated in the MB calyces. This was doubly unexpected because results in 5664 3D showed an upregulation instead, but also because increased neural activity resulting from 5665 experience-dependent phenomena is usually reflected by an upregulation of IEG expression 5666 5667 (Bahrami and Drabløs, 2016). The downregulation suggests an inhibition of neural activity in key areas involved in visual processing- optic lobes and mushroom bodies - of the learner 5668 5669 group.

5670 Inhibition of the optic lobes, suggested by the downregulation of Egrl in that region, is coherent 5671 with the multiple GABAergic fibers innervating the medulla and lobula (Schäfer and Bicker,

1986) and with the increase in Amgad expression, the gene coding for a key enzyme of the 5672 5673 GABA synthesis, in the optic lobes in foragers (Kiya and Kubo, 2010). In the MB neural inhibition is provided by GABAergic feedback neurons (Av3 neurons) (Rybak and Menzel, 5674 5675 1993), which are responsible for the sparse coding responses exhibited by Kenvon cells. These neurons have been shown to be involved in negative patterning (Devaud et al., 2015). Inhibition 5676 5677 at the level of the MBs may therefore be part of certain learning phenomena, which require 5678 enhanced neural sparseness to de-correlate stimulus representations and thus increase memory specificity. Both kakusei and Hr38 downregulation in the MBs in the 2D VR may be the 5679 consequence of plastic changes in GABAergic signaling in the calyces of the MBs. 5680

5681 Both VR experiments were done under similar handling conditions, used strictly the same behavioral criteria and presented the same colors as stimuli. Genetic analyses were also 5682 performed under the same conditions and using the same materials and methods. The main 5683 difference comes from the way the bees were able to inspect the stimuli: in 3D, the object were 5684 cuboids, could be approach from any side and would expand and retract as the bee got closer 5685 5686 or further away from them; in 2D, the bee could only move the objects laterally and could not get closer or further away from them. Erg1 upregulation in the learner group in the 3D VR 5687 could thus results from an interaction between an exploratory drive of the environment and 5688 5689 learning. In 2D VR, GABA-mediated inhibition may act as a gain control mechanism that enhances response efficiency and stimulus control. It has indeed been proposed to play a 5690 fundamental role in establishing selectivity for stimulus orientation and direction of motion in 5691 mammals (Rose and Blakemore, 1974; Sillito, 1979; Tsumoto et al., 1979). As the latter is 5692 5693 particularly important in the 2D VR, enhanced GABA inhibition could be associated with 5694 learning to master the visual discrimination in this context. It is also possible that the two objects appear more similar in 2D as the insects have less opportunity to inspect them, it could thus 5695 5696 require sparse coding that has been shown to be required to discriminate similar odors in flies

(Lin et al., 2014). However, we measured no difference in learning success between 2D and 3D 5697 which goes against an increased difficulty of the task in 2D. Another possible explanation is a 5698 possible difference in the visual acquisition mechanisms recruited by these two scenarios. On 5699 one hand, the 3D experiment might include a navigation component where body movements 5700 5701 translate into a displacement and a recognizable change in the visual scene, which can then be computed against the available internal information about the displacement. While, on the other 5702 hand, the 2D experiment is closer to a purely operant task where the animal needs to engage in 5703 5704 two different, but stereotyped motor patterns, turn left or turn right, according to the position of CS+ on the screen. The observed differences in IEG expression between the two types of VR 5705 would then reflect the two different mechanisms used to reach the rewarded stimulus. One 5706 involving navigation while the other is purely operant. 5707

Taken together chapter 2 and 3 suggest the existence of a distributed memory trace along the visual system and highlight the importance of MBs for color learning in bees. They also point at the OLs, and the calyces of the MBs as region of interest for further investigation of the neural correlates of visual learning.



Figure 1. The different visual neuronal populations and pathways of the honeybee brain. The black arrow indicates color stimulation. La = lamina, χo = outer chiasm, me = medulla, χI = inner chiasm, lo = lobula, le = lateral calyx of the mushroom bodies, me = median calyx, α = alpha-lobe, β = beta-lobe, al = antennal lobe, ot = anterior optic tuberculum. MB: mushroom bodies; CC: central complex. Courtesy of M. Giurfa.

Throughout chapter 2 and 3 we have relied heavily on two terms, *learner* and *non-learner*. We 5719 5720 have defined the *learners* as the animals that made the correct choice during the non-rewarded test and the non-learners as the rest, meaning those that either chose wrong or did not chose. 5721 During the first trial, naive bees have a 40% chance to choose the CS+ in 3D, as measured in 5722 5723 the first trials of our 3D experiments (chapter 1) and about 50% in 2D (first trial, chapter 3), meaning that in both situations the probability to randomly be a *learner* is very high. Through 5724 the conditioning process the proportion of correct choices in the population increases 5725 significantly, meaning that the bees are changing their behavior and are not choosing randomly 5726 anymore, they are learning. That's why we still use that classification, because we think it is 5727 reasonable to expect that the bees that made a correct choice at the end are the ones that learned 5728 5729 the association, since we clearly see that some bees are learning during the conditioning process. But in truth we still have no certitude over who actually learned something and who still chose 5730 5731 randomly. By using the first choice as a criterion we know that the *learner* group should contain

more individual that learned something but we can't identify them and guarantee that they all 5732 actually learned something. First choice and other similar binomial variables are more 5733 informative about the population than the individuals (Gallistel et al., 2004; Pamir et al., 2011, 5734 5735 2014). However, in studies like the one in chapter 2 and 3, we need individual information to be able to confidently identify the animals that learned the task. Therefore, future studies aiming 5736 5737 at measuring variations induced by learning in the brain of animal will require more quantitative 5738 variables like the latency to display the condition response, the stability of the conditioned response across trials (Gallistel et al., 2004; Pamir et al., 2014) or the time spent choosing the 5739 rewarded stimulus. Using quantitative variables will still contain an element of arbitrariness in 5740 the establishment of thresholds but it will give us more information about each individual and 5741 get us closer to know which one actually learned something. 5742

5743 Bumblebees suitability for VR experiments

Finally, in the last part of this manuscript we explored the possibility of using bumblebees *Bombus terrestris* as a model in VR by measuring their performances in a color discrimination task under VR condition and compared with honeybees. The idea for this experiment came after we observed that honeybees that were injected with PBS solution in the central complex, the calyces of the mushroom bodies or the antennal lobes were not able to solve the discrimination task in VR anymore (data not shown), we thus decided to test a more robust species that could potentially make this kind of experiment more likely to succeed.

First we measured the size and weight of every individual to control for a potential effect of weight on the ability to navigate the VR, since bigger animals should have an easier time in moving the ball (Dahmen et al., 2017). 5754 While weight had no effect on distance or speed we found that larger bumblebees walked more 5755 than smaller ones. This is likely due a difference in the length of each step as larger bumblebees 5756 were also walking faster.

5757 Size and weight did not seem to affect bumblebees' learning performance in our VR experiment. Several studies have shown that bigger bumblebees have a better visual acuity due 5758 to having bigger eyes (Macuda et al., 2001; Spaethe and Chittka, 2003; Taylor et al., 2019; 5759 5760 Wertlen et al., 2008) and have better foraging success (Spaethe and Weidenmüller, 2002; Ings et al., 2005; Klein et al., 2017). But their increased foraging success can be explained by the 5761 5762 fact that larger bees are able to forage in cooler conditions (Heinrich and Heinrich, 1983), may 5763 be able to forage over larger distances, and are perhaps also less vulnerable to predation 5764 (Goulson et al., 2002). Coherent with these observations, larger bumblebee often assume the role of forager for the colony (Goulson, 2003), it is thus possible that their increased learning 5765 performances observed in previous study (Worden et al., 2005) are actually a consequence of 5766 their life experience as foragers (Giurfa et al., 2003; Cabirol et al., 2017). While bumblebees 5767 5768 used in Worden et al. study were allowed to forage freely at a feeder station, in our study we used commercially reared bumblebees that were completely naive and had no foraging 5769 experience prior to our experiment, which would then explain why the larger bumblebees did 5770 5771 not perform better. When investigating correlations between learning speed and foraging success across colonies, Raine and Chittka did not evidence any link between individual forager 5772 size and learning speed when conditioning bumblebees to collect a sucrose reward from colored 5773 5774 feeders in free flying conditions (Raine and Chittka, 2008). The bumblebees used in that study 5775 were also taken from commercial colonies and had no prior exposure to colored stimuli 5776 associated with food, this is coherent with our hypothesis that body size only affects learning speed indirectly through foraging experience. 5777

Bumblebees were able to learn to discriminate cylinders differing in color and reinforcing 5778 5779 outcome in a VR context. After 6 trials, about 60% of bumblebees chose the rewarded stimulus over the punished one. Those results are comparable with honeybees performances both in this 5780 5781 experiments and in previous studies (Buatois et al., 2017; Lafon et al., 2021). We also found that using quinine solution as negative reinforcement gave better results as bumblebee 5782 5783 conditioned with NaCl were only able to solve the discrimination when the rewarded cylinder 5784 was blue, a color they tend to learn faster (Gumbert, 2000). The weaker effect of salt observed here with bumblebees is coherent with previous results found in honeybees where bees were 5785 not able to solve the discrimination when the CS- was paired with NaCl solution (Buatois et al., 5786 2017). 5787

Throughout our work we have applied one consistent criterion to include or not insects in the 5788 study: if a bee doesn't make a choice for at least half the conditioning trials it is discarded. In 5789 this experiment this lead us to discard about 60% of honeybees. While for bumblebees, we only 5790 discarded 11% of individuals. The higher motivation of bumblebees could be explained by their 5791 5792 ecology, since they tend to nest underground they might have less problem walking in the dark for prolonged amount of time compared to honeybees, it could also be explained by a size 5793 difference as larger bumblebees were making longer strides and thus had an easier time moving 5794 5795 in the VR. In any case, this makes conditioning a large number of insects easier in bumblebees as the ratio of useable data over conditioned bees is much higher. 5796

In the past decades bumblebees have proved to be a good model to complement honeybee
research (Riveros and Gronenberg, 2009) thanks to their good cognitive abilities (Laverty,
1994; Laloi et al., 1999; Leadbeater and Chittka, 2005; Worden et al., 2005; Raine and Chittka,
2007; Leadbeater and Chittka, 2007) coupled with their robustness under restrained conditions
and during electrophysiology recordings (Paulk et al., 2008, 2009; Skorupski and Chittka, 2010;
Vähäkainu et al., 2013; Rusanen et al., 2017). In this context our results strongly suggest that

the bumblebee would be an excellent model for investigating further the neural correlate of visual learning in VR using invasive techniques, as they possess the ability to learn successfully in VR and the robustness to endure the required surgeries.

5806 Conclusion

5807 So, is VR the new revolution for the study of bee behavior?

Not quite. While it does allow to study visual learning in tethered animals, the throughput of tested animals is still lower than with olfactory PER conditioning and the technical entry cost is much higher. However, VR offers way more possibilities than PER as the animals are actually moving and thus are more prone to exhibit a richer behavior. VR is a versatile tool, which allows for manipulation of multiple variables and for sophisticated analyses of behavior.

Overall our work improved the field of visual learning by producing a robust 3D VR system 5813 5814 that is inexpensive, open source and supports experiments on both bumblebees and honeybees. We proved that it can reliably be used to condition bees in color discrimination through several 5815 different studies, that also allowed us to refine conditioning protocols in VR. We were able to 5816 use our setup to push our understanding of the neural mechanism of visual learning a little 5817 further by performing the first quantification of IEGs variations in a controlled visual learning 5818 5819 experiment. In order to fully exploit the possibilities that this setup opens we now need to develop a way to couple it with live recording through either calcium imaging or 5820 electrophysiology. As a first step in that direction we also showed that bumblebees, known for 5821 5822 their resilience, are also a good model for the study of visual learning in VR. Taken together our results open the way for a deeper exploration of visual learning through VR 5823 5824 experimentation.

5826 References

5827 Bahrami, S., Drabløs, F., 2016. Gene regulation in the immediate-early response process. Advances in 5828 Biological Regulation 62, 37-49. https://doi.org/10.1016/j.jbior.2016.05.001 5829 Baird, E., Boeddeker, N., Srinivasan, M.V., 2021. The effect of optic flow cues on honeybee flight 5830 control in wind. Proc. R. Soc. B. 288, 20203051. https://doi.org/10.1098/rspb.2020.3051 5831 Baird, E., Srinivasan, M.V., Zhang, S., Lamont, R., Cowling, A., 2006. Visual Control of Flight Speed and 5832 Height in the Honeybee, in: Nolfi, S., Baldassarre, G., Calabretta, R., Hallam, J.C.T., Marocco, 5833 D., Meyer, J.-A., Miglino, O., Parisi, D. (Eds.), From Animals to Animats 9, Lecture Notes in 5834 Computer Science. Springer, Berlin, Heidelberg, pp. 40–51. 5835 https://doi.org/10.1007/11840541_4 Balamurali, G.S., Somanathan, H., Hempel de Ibarra, N., 2015. Motion cues improve the performance 5836 5837 of harnessed bees in a colour learning task. J Comp Physiol A 201, 505–511. https://doi.org/10.1007/s00359-015-0994-7 5838 5839 Buatois, A., Flumian, C., Schultheiss, P., Avarguès-Weber, A., Giurfa, M., 2018. Transfer of Visual 5840 Learning Between a Virtual and a Real Environment in Honey Bees: The Role of Active Vision. 5841 Front Behav Neurosci 12, 139. https://doi.org/10.3389/fnbeh.2018.00139 5842 Buatois, A., Laroche, L., Lafon, G., Avarguès-Weber, A., Giurfa, M., 2020. Higher-order discrimination 5843 learning by honeybees in a virtual environment. European Journal of Neuroscience 51, 681-5844 694. https://doi.org/10.1111/ejn.14633 5845 Buatois, A., Pichot, C., Schultheiss, P., Sandoz, J.-C., Lazzari, C.R., Chittka, L., Avarguès-Weber, A., 5846 Giurfa, M., 2017. Associative visual learning by tethered bees in a controlled visual 5847 environment. Sci Rep 7. https://doi.org/10.1038/s41598-017-12631-w 5848 Cabirol, A., Brooks, R., Groh, C., Barron, A.B., Devaud, J.-M., 2017. Experience during early adulthood 5849 shapes the learning capacities and the number of synaptic boutons in the mushroom bodies 5850 of honey bees (Apis mellifera). Learn Mem 24, 557–562. 5851 https://doi.org/10.1101/lm.045492.117 5852 Dahmen, H., Wahl, V.L., Pfeffer, S.E., Mallot, H.A., Wittlinger, M., 2017. Naturalistic path integration 5853 of Cataglyphis desert ants on an air-cushioned lightweight spherical treadmill. Journal of 5854 Experimental Biology 220, 634–644. https://doi.org/10.1242/jeb.148213 5855 Devaud, J.-M., Papouin, T., Carcaud, J., Sandoz, J.-C., Grünewald, B., Giurfa, M., 2015. Neural 5856 substrate for higher-order learning in an insect: Mushroom bodies are necessary for 5857 configural discriminations. Proc Natl Acad Sci U S A 112, E5854–E5862. 5858 https://doi.org/10.1073/pnas.1508422112 Frasnelli, E., Hempel de Ibarra, N., Stewart, F.J., 2018. The Dominant Role of Visual Motion Cues in 5859 Bumblebee Flight Control Revealed Through Virtual Reality. Front Physiol 9. 5860 5861 https://doi.org/10.3389/fphys.2018.01038 5862 Fujita, N., Nagata, Y., Nishiuchi, T., Sato, M., Iwami, M., Kiya, T., 2013. Visualization of Neural Activity 5863 in Insect Brains Using a Conserved Immediate Early Gene, Hr38. Current Biology 23, 2063-2070. https://doi.org/10.1016/j.cub.2013.08.051 5864 5865 Gallistel, C.R., Fairhurst, S., Balsam, P., 2004. The learning curve: implications of a quantitative analysis. Proc Natl Acad Sci U S A 101, 13124–13131. 5866 5867 https://doi.org/10.1073/pnas.0404965101 Giurfa, M., Schubert, M., Reisenman, C., Gerber, B., Lachnit, H., 2003. The effect of cumulative 5868 experience on the use of elemental and configural visual discrimination strategies in 5869 5870 honeybees. Behavioural Brain Research 145, 161–169. https://doi.org/10.1016/S0166-5871 4328(03)00104-9 5872 Goulson, D., 2003. Bumblebees: Their Behaviour and Ecology. Oxford University Press. 5873 Goulson, D., Peat, J., Stout, J.C., Tucker, J., Darvill, B., Derwent, L.C., Hughes, W.O.H., 2002. Can 5874 alloethism in workers of the bumblebee, Bombus terrestris, be explained in terms of foraging 5875 efficiency? Animal Behaviour 64, 123–130. https://doi.org/10.1006/anbe.2002.3041

- 5876 Gumbert, A., 2000. Color choices by bumble bees (Bombus terrestris): innate preferences and generalization after learning. Behav Ecol Sociobiol 48, 36–43.
 5878 https://doi.org/10.1007/s002650000213
 5870 University P., University M. J. 5., 1082, University in Examples of the second present of the second pres
- 5879 Heinrich, B., Heinrich, M.J.E., 1983. Heterothermia in Foraging Workers and Drones of the
 5880 Bumblebee Bombus terricola. Physiological Zoology 56, 563–567.
 5881 https://doi.org/10.1086/physzool.56.4.30155879
- 5882 lino, S., Shiota, Y., Nishimura, M., Asada, S., Ono, M., Kubo, T., 2020. Neural activity mapping of
 5883 bumble bee (Bombus ignitus) brains during foraging flight using immediate early genes. Sci
 5884 Rep 10, 7887. https://doi.org/10.1038/s41598-020-64701-1
- Ings, T.C., Schikora, J., Chittka, L., 2005. Bumblebees, humble pollinators or assiduous invaders? A
 population comparison of foraging performance in Bombus terrestris. Oecologia 144, 508–
 516. https://doi.org/10.1007/s00442-005-0081-9
- 5888 Kiya, T., Kubo, T., 2010. Analysis of GABAergic and Non-GABAergic Neuron Activity in the Optic Lobes
 5889 of the Forager and Re-Orienting Worker Honeybee (Apis mellifera L.). PLOS ONE 5, e8833.
 5890 https://doi.org/10.1371/journal.pone.0008833
- 5891 Kiya, T., Kunieda, T., Kubo, T., 2007. Increased Neural Activity of a Mushroom Body Neuron Subtype
 5892 in the Brains of Forager Honeybees. PLOS ONE 2, e371.
 5893 https://doi.org/10.1371/journal.pone.0000371
- Klein, S., Pasquaretta, C., Barron, A.B., Devaud, J.-M., Lihoreau, M., 2017. Inter-individual variability in
 the foraging behaviour of traplining bumblebees. Sci Rep 7, 4561.
 https://doi.org/10.1038/s41598-017-04919-8
- Lafon, G., Howard, S.R., Paffhausen, B.H., Avarguès-Weber, A., Giurfa, M., 2021. Motion cues from
 the background influence associative color learning of honey bees in a virtual-reality
 scenario. Sci Rep 11, 21127. https://doi.org/10.1038/s41598-021-00630-x
- Laloi, D., Sandoz, J. c., Picard-Nizou, A. I., Marchesi, A., Pouvreau, A., Taséi, J. n., Poppy, G., Phamdelègue, M. h., 1999. Olfactory conditioning of the proboscis extension in bumble bees.
 Entomologia Experimentalis et Applicata 90, 123–129. https://doi.org/10.1046/j.15707458.1999.00430.x
- 5904Laverty, T.M., 1994. Bumble bee learning and flower morphology. Animal Behaviour 47, 531–545.5905https://doi.org/10.1006/anbe.1994.1077
- 5906Leadbeater, E., Chittka, L., 2007. The dynamics of social learning in an insect model, the bumblebee5907(Bombus terrestris). Behav Ecol Sociobiol 61, 1789–1796. https://doi.org/10.1007/s00265-5908007-0412-4
- 5909Leadbeater, E., Chittka, L., 2005. A new mode of information transfer in foraging bumblebees? Curr5910Biol 15, R447-448. https://doi.org/10.1016/j.cub.2005.06.011
- Lin, A.C., Bygrave, A.M., de Calignon, A., Lee, T., Miesenböck, G., 2014. Sparse, decorrelated odor
 coding in the mushroom body enhances learned odor discrimination. Nat Neurosci 17, 559–
 568. https://doi.org/10.1038/nn.3660
- Lutz, C.C., Robinson, G.E., 2013. Activity-dependent gene expression in honey bee mushroom bodies
 in response to orientation flight. J Exp Biol 216, 2031–2038.
 https://doi.org/10.1242/jeb.084905
- Macuda, T., Gegear, R., Laverty, T., Timney, B., 2001. Behavioural assessment of visual acuity in
 bumblebees (Bombus impatiens). Journal of Experimental Biology 204, 559–564.
 https://doi.org/10.1242/jeb.204.3.559
- Marchal, P., Villar, M.E., Geng, H., Arrufat, P., Combe, M., Viola, H., Massou, I., Giurfa, M., 2019.
 Inhibitory learning of phototaxis by honeybees in a passive-avoidance task. Learning and
 Memory 26, 412–423. https://doi.org/10.1101/lm.050120.119
- Menzel, R., 2014. The insect mushroom body, an experience-dependent recoding device. Journal of
 Physiology-Paris, Neuroethology: A Tribute to Hector Maldonado 108, 84–95.
 https://doi.org/10.1016/j.jphysparis.2014.07.004
- 5926 Menzel, R., 1999. Memory dynamics in the honeybee. J Comp Physiol A 185, 323–340.
- 5927 https://doi.org/10.1007/s003590050392

5932 Pamir, E., Szyszka, P., Scheiner, R., Nawrot, M.P., 2014. Rapid learning dynamics in individual 5933 honeybees during classical conditioning. Frontiers in Behavioral Neuroscience 8. 5934 Paulk, A.C., Dacks, A.M., Gronenberg, W., 2009. Color processing in the medulla of the bumblebee 5935 (Apidae: Bombus impatiens). Journal of Comparative Neurology 513, 441–456. 5936 https://doi.org/10.1002/cne.21993 5937 Paulk, A.C., Phillips-Portillo, J., Dacks, A.M., Fellous, J.-M., Gronenberg, W., 2008. The Processing of 5938 Color, Motion, and Stimulus Timing Are Anatomically Segregated in the Bumblebee Brain. J 5939 Neurosci 28, 6319-6332. https://doi.org/10.1523/JNEUROSCI.1196-08.2008 5940 Paulk, A.C., Stacey, J.A., Pearson, T.W.J., Taylor, G.J., Moore, R.J.D., Srinivasan, M.V., Swinderen, B. 5941 van, 2014. Selective attention in the honeybee optic lobes precedes behavioral choices. PNAS 5942 111, 5006–5011. https://doi.org/10.1073/pnas.1323297111 5943 Plath, J.A., Entler, B.V., Kirkerud, N.H., Schlegel, U., Galizia, C.G., Barron, A.B., 2017. Different Roles 5944 for Honey Bee Mushroom Bodies and Central Complex in Visual Learning of Colored Lights in 5945 an Aversive Conditioning Assay. Front. Behav. Neurosci. 11. 5946 https://doi.org/10.3389/fnbeh.2017.00098 5947 Raine, N.E., Chittka, L., 2008. The correlation of learning speed and natural foraging success in 5948 bumble-bees. Proc Biol Sci 275, 803-808. https://doi.org/10.1098/rspb.2007.1652 5949 Raine, N.E., Chittka, L., 2007. Pollen foraging: learning a complex motor skill by bumblebees (Bombus 5950 terrestris). Naturwissenschaften 94, 459–464. https://doi.org/10.1007/s00114-006-0184-0 5951 Riveros, A.J., Gronenberg, W., 2009. Learning from learning and memory in bumblebees. Commun 5952 Integr Biol 2, 437–440. 5953 Rose, D., Blakemore, C., 1974. Effects of bicuculline on functions of inhibition in visual cortex. Nature 5954 249, 375–377. https://doi.org/10.1038/249375a0 5955 Rusanen, J., Vähäkainu, A., Weckström, M., Arikawa, K., 2017. Characterization of the first-order 5956 visual interneurons in the visual system of the bumblebee (Bombus terrestris). J Comp 5957 Physiol A 203, 903-913. https://doi.org/10.1007/s00359-017-1201-9 5958 Rusch, C., Alonso San Alberto, D., Riffell, J.A., 2021. Visuo-motor feedback modulates neural activities 5959 in the medulla of the honeybee, Apis mellifera. J Neurosci. 5960 https://doi.org/10.1523/JNEUROSCI.1824-20.2021 5961 Rybak, J., Menzel, R., 1993. Anatomy of the mushroom bodies in the honey bee brain: the neuronal 5962 connections of the alpha-lobe. J Comp Neurol 334, 444-465. 5963 https://doi.org/10.1002/cne.903340309 5964 Schäfer, S., Bicker, G., 1986. Distribution of GABA-like immunoreactivity in the brain of the honeybee. 5965 J Comp Neurol 246, 287–300. https://doi.org/10.1002/cne.902460302 Sillito, A.M., 1979. Inhibitory mechanisms influencing complex cell orientation selectivity and their 5966 5967 modification at high resting discharge levels. J Physiol 289, 33–53. 5968 https://doi.org/10.1113/jphysiol.1979.sp012723 5969 Singh, A.S., Shah, A., Brockmann, A., 2018. Honey bee foraging induces upregulation of early growth 5970 response protein 1, hormone receptor 38 and candidate downstream genes of the 5971 ecdysteroid signalling pathway. Insect Molecular Biology 27, 90–98. 5972 https://doi.org/10.1111/imb.12350 5973 Skorupski, P., Chittka, L., 2010. Photoreceptor Spectral Sensitivity in the Bumblebee, Bombus 5974 impatiens (Hymenoptera: Apidae). PLOS ONE 5, e12049. 5975 https://doi.org/10.1371/journal.pone.0012049 5976 Spaethe, J., Chittka, L., 2003. Interindividual variation of eye optics and single object resolution in 5977 bumblebees. Journal of Experimental Biology 206, 3447–3453.

Pamir, E., Chakroborty, N.K., Stollhoff, N., Gehring, K.B., Antemann, V., Morgenstern, L., Felsenberg,

J., Eisenhardt, D., Menzel, R., Nawrot, M.P., 2011. Average group behavior does not

733-741. https://doi.org/10.1101/lm.2232711

represent individual behavior in classical conditioning of the honeybee. Learn. Mem. 18,

5978 https://doi.org/10.1242/jeb.00570

5928

5929

5930

- 5979 Spaethe, J., Weidenmüller, A., 2002. Size variation and foraging rate in bumblebees (Bombus 5980 terrestris). Insectes soc. 49, 142–146. https://doi.org/10.1007/s00040-002-8293-z
- Taylor, G.J., Tichit, P., Schmidt, M.D., Bodey, A.J., Rau, C., Baird, E., 2019. Bumblebee visual allometry
 results in locally improved resolution and globally improved sensitivity. eLife 8, e40613.
 https://doi.org/10.7554/eLife.40613
- 5984Tsumoto, T., Eckart, W., Creutzfeldt, O.D., 1979. Modification of orientation sensitivity of cat visual5985cortex neurons by removal of GABA-mediated inhibition. Exp Brain Res 34, 351–363.5986https://doi.org/10.1007/BF00235678
- 5987 Ugajin, A., Uchiyama, H., Miyata, T., Sasaki, T., Yajima, S., Ono, M., 2018. Identification and initial
 5988 characterization of novel neural immediate early genes possibly differentially contributing to
 5989 foraging-related learning and memory processes in the honeybee. Insect Molecular Biology
 5990 27, 154–165. https://doi.org/10.1111/imb.12355
- Vähäkainu, A., Vähäsöyrinki, M., Weckström, M., 2013. Membrane filtering properties of the
 bumblebee (Bombus terrestris) photoreceptors across three spectral classes. J Comp Physiol
 A 199, 629–639. https://doi.org/10.1007/s00359-013-0814-x
- Wertlen, A.M., Niggebrügge, C., Vorobyev, M., Hempel de Ibarra, N., 2008. Detection of patches of
 coloured discs by bees. Journal of Experimental Biology 211, 2101–2104.
 https://doi.org/10.1242/jeb.014571
- Worden, B.D., Skemp, A.K., Papaj, D.R., 2005. Learning in two contexts: the effects of interference
 and body size in bumblebees. Journal of Experimental Biology 208, 2045–2053.
 https://doi.org/10.1242/jeb.01582
- Zwaka, H., Bartels, R., Lehfeldt, S., Jusyte, M., Hantke, S., Menzel, S., Gora, J., Alberdi, R., Menzel, R.,
 2019. Learning and Its Neural Correlates in a Virtual Environment for Honeybees. Front
 Behav Neurosci 12, 279. https://doi.org/10.3389/fnbeh.2018.00279
- 6003

Curriculum Vitae

Lafon Grégory

PHD · NEUROETHOLOGY

Education	
Université Toulouse III - Paul Sabatier PHD Neuroethology: "Visual learning in bees under virtual reality conditions" • Co-Supervisors: Pr. Martin Giurfa, Dr. Aurore Avarguès-Weber	Toulouse, France 2018 - 2022
ENS Cachan ENS Саснал Diploma • Successfully completed the 4 year program of the ENS Cachan	Paris, France 2013 - 2017
Université Toulouse III - Paul Sabatier Master Neuroscience, Behaviour, Cognition	Toulouse, France 2017 - 2018
ENS Lyon Master Earth and Life Sciences	Lyon, France 2015 - 2016
ENS Cachan Licence Biology	Paris, France 2013 - 2014
Lycée Henri IV CPGE BCPST • Admission at the ENS Cachan, 32th out of 800	Paris, France 2010 - 2013
Research Experience	
Université Toulouse III - CRCA-CBI Co-Supervisors: Pr. Martin Giurfa, Dr. Aurore Avarguès-Weber • PhD: Visual learning in Honey bees under virtual reality conditions	Toulouse, France 2018 - 2022
McGill University - Dept of Biology SUPERVISOR: DR. JON SAKATA • Internship: Role of dopamine within HVC in social context related song modifications	Montreal, Canada Nov.2016 - Aug.2017
Université Pierre et Marie Curie - Dept of Neuroscience SUPERVISOR: DR. CHRISTELLE ROCHEFORT • Internship: Influence of LTP in synapses between parallel fibers and Purkinje cells on place cells	Paris, France Jun - July 2015
University Paris-Saclay, CNRS SUPERVISOR: DR. JEAN-CHRISTOPHE SANDOZ • Internship: Influence of maturity on olfactory attraction between western honey bee drones	Gif-Sur-Yvette, France Jun - July 2014

Teaching Experience _____

2018 - 2021 Ethology, Practical Course, Teaching Assistant	Ethology Practical Course Teaching Assistant	Université
	Linology, Fractical course, reaching Assistant	Toulouse III
2018 - 2021 Behavioural Ecology, Practical Course, Teaching Assistant	Behavioural Ecology Practical Course Teaching Assistant	Université
	Toulouse III	
2010 2020	019 - 2020 Biology of Behaviour, Practical Course, Teaching Assistant	Université
2013 - 2020		Toulouse III
2020 lr	Introduction to the Scientific Method, Practical Course, Teaching Assistant	Université
		Toulouse III
2018	Behavioural Neuroscience, Practical Course, Teaching Assistant	Université
		Toulouse III

Mentoring_____

2021	Naïs Judan (BS), Eva Blot (BS), and Karolina Pecharova (MS), Visual learning in bumble	Université
2021	bees under virtual reality conditions	Toulouse III
2021	Catherine Macri (MS), Marin Nicola (MS), Evaluation of cerebral nanoinjection's effects on	Université
2021	visual learning performances under virtual reality conditions in Apis mellifera	Toulouse III
2021	Clemence Guinnement (MS), Redaction of a review on the neurobiological mechanism of	Université
2021	visual learning in the honey bee	Toulouse III
2019	Rodrigue Fouillet (MS), Juliette Montet (MS), Diane Sam Mine (BS), Emma Giordanengo	Université
	(BS), Influence of Background motion cues in VR on color discrimination in the honey bee	Toulouse III

Skills_____

Beekeeping, Insect conditioning, Basic dissections
3D printing, Arduino, Soldering
French, English
C++, C#, R, Gdscript
Github, Unity, Visual Studio
Gimp, Aseprite

Publications_____

Published

- **Gregory Lafon**^{*}, Haiyang Geng^{*}, Aurore Avarguès-Weber, Alexis Buatois, Isabelle Massou, Martin Giurfa. 2022. The Neural signature of visual learning under restrictive virtual-reality conditions. Frontiers in Behavioral Neuroscience, 16:846076. doi: 10.3389/fnbeh.2022.846076
- Haiyang Geng*, **Gregory Lafon***, Aurore Avarguès-Weber, Alexis Buatois, Isabelle Massou, Martin Giurfa . 2022. Visual learning in a virtual reality environment upregulates immediate early gene expression in the mushroom bodies of honey bees. Communications Biology, 14;5(1):130. doi: 10.1038/s42003-022-03075-8
- **Gregory Lafon**, Scarlett R. Howard, Benjamin H. Paffhausen, Aurore Avarguès-Weber, Martin Giurfa . 2021. Motion cues from the background influence associative color learning of honey bees in a virtual-reality scenario. Scientific Reports, 11(1):21127, doi: 10.1038/s41598-021-00630-x
- Alexis Buatois, Lou Laroche, **Gregory Lafon**, Aurore Avarguès-Weber, Martin Giurfa . 2020. Higher order discrimination learning by honey bees in a virtual environment. European Journal of Neuroscience, 51(2):681-694, doi: 10.1111/ejn.14633
- Florian Bastin, Hanna Cholé, **Grégory Lafon**, Jean-Christophe Sandoz . 2017. Virgin queen attraction toward males in honey bees. Scientific Reports, 7(1):6293. doi: 10.1038/s41598-017-06241-9
Florian Bastin, Fabrice Savarit, **Grégory Lafon**, Jean-Christophe Sandoz . 2017. Age-specific olfactory attraction between Western honey bee drones (Apis mellifera) and its chemical basis. PLoS One, 12(10):e0185949. doi: 10.1371/journal.pone.0185949

* first authorship shared

In Prep

Gregory Lafon, Aurore Avarguès-Weber, Martin Giurfa . Comparison of associative visual learning in a 3D virtual reality between bumble bees and honey bees.

Other Projects ____

OPEN SOURCE DEVELOPMENT

2019-.. Cataclysm: Dark Days Ahead, Core dev team member cataclysmdda.org